

For VE-cad and α-SMA immunostaining, cells were washed with PBS and fixed with 100% methanol for 15 min in -20 °C. The fixed cells were rinsed three times with PBS for 5 min each and incubated in a blocking buffer containing 5% BSA in PBST for 1 h. Mouse anti-human α-SMA antibody and rabbit anti-VE-cad antibody were added to cells in blocking buffer, and incubated overnight at 4 °C. From this step on, cells were protected from light. After rinsing in PBST three times (10 min each), cells were incubated with a cocktail of Alexa Fluor 488-conjugated goat anti-mouse and Alexa Fluor 555-conjugated donkey anti-rabbit antibody in blocking buffer at room temperature (RT) for 1 h. Samples were washed three times in PBST for 10 min each, then stained with DRAQ5 in PBST at RT for 15 min. The fluorescence images were taken with an Echo revolve fluorescence microscope.