

# A New Frontier in Fighting Brain Cancer: Cutting Edge Magnetic Resonance Imaging Techniques

## ABSTRACT

Of the currently available brain imaging techniques for diagnosing tumors, diffusion-weighted MRI and perfusion MRI are cutting-edge techniques and may provide improved diagnostic capacity compared to traditional techniques such as positron emission tomography. Moreover, they are fully non-invasive and avoid the exposure to radiation. While MRI in general has been used in research and medicine for decades, the more recent development of multi-modal and multiparametric imaging in neuro oncology holds much promise for the enhancement of diagnosis, prognosis, and patient-tailored treatments in this field. This review will evaluate how these various imaging techniques provide clinical value above and beyond previous techniques.

## Introduction

Neuro-oncologists are increasingly using diffusion-weighted imaging (DWI) and perfusion MRI to not only diagnose brain tumors, but also guide surgical procedures and monitor the tumor treatment response. DWI allows for the detection of water diffusion. Water diffusion is inversely related to tumor cellularity- meaning the increase of one causes the decrease of the other; in other words, as cellularity decreases due to damage or altered structure, the free diffusion of water in a tissue increases, allowing for the noninvasive detection of tumor cellularity. Perfusion MRI techniques also help oncologists evaluate tumor structure by analyzing tumor vascularity in the context of neoangiogenesis, a process where new blood vessels grow to support the invasion of tumor cells. These advanced techniques are becoming the standard of use for the detection, characterization, and staging of viable tumor lesions. Additionally, these tools become especially useful in malignant tumors such as glioblastomas, metastatic tumors, and lymphomas.

Within DWI and perfusion MRI, different novel MRI analytic techniques further provide unique benefits to oncologists. For example, intravoxel incoherent motion (IVIM), found in DWI, is utilized, with the absence of contrast agents, for the evaluation of tissue perfusion and separation of microcirculation from true water molecular diffusion.<sup>1</sup> Diffusion kurtosis imaging (DKI) can serve a multitude of tasks.<sup>2</sup> DKI locates non-Gaussian diffusion, which may help describe the brain regions' structural components.<sup>2</sup> Perfusion MRI comprises three main methods that are all primarily used to evaluate malignant brain tumors.<sup>3</sup> The three primary perfusion MRI techniques used in neuro oncology—dynamic susceptibility contrast (DSC) perfusion MRI, dynamic contrast-enhanced (DCE) perfusion MRI, and arterial spin labeling (ASL)—gift unique characterizations of the pathophysiology of a patient's pre- and post-treatment brain tumors.<sup>4</sup>

As a biomarker of the glioma outcome, the tumor microvessel area (MVA) could be accurately derived from the relative cerebral blood volume (rCBV) calculated from the DSC perfusion MRI.<sup>5</sup> DCE perfusion MRI differentiates mature from immature tumor vessels.<sup>6</sup> ASL can be used to obtain cerebral blood flow (CBF), which is extremely useful for oncologists because CBF may predict tumor vascular normalization, a therapeutic strategy aimed towards preventing angiogenesis.<sup>7</sup> Cerebral blood flow (CBF) is the rate at which the arterial blood is transported into the brain's capillary bed. Contrast-enhanced susceptibility-weighted imaging (CE-SWI) may further evaluate the cellular and vascular consistency of tumors above and beyond the ability of techniques without contrast agents.<sup>8</sup> This review will discuss each advanced MR imaging techniques separately—diving deep into the clinical usefulness as well as limitations of each method.

## Discussion

Apparent Diffusion Coefficient Characterizes Many Tumor Aspects and Tracks Antiangiogenic Drugs Response

Diffusion-weighted MRI is able to detect the magnitude of this microscopic, subvoxel water motion and this is used to derive several metrics, including the apparent diffusion coefficient (ADC), a measure of the degree of water diffusion within a voxel.<sup>9</sup> Water molecules found in extracellular, intracellular, and intravascular regions are restricted in their movement differently, producing differential ADC signals and allowing for tissue-type segmentation.<sup>10</sup> Specifically, each space is characterized by different microscopic anatomical barriers, which leads to differential perfusion of water, both in terms of the directionality and extent of diffusion.<sup>10</sup> Generally, ADC gives oncologists a fair approximation of the water diffusion within the extracellular and extravascular space.<sup>11</sup> Since tumors are composed of tightly packed cells (they are highly cellular), the extracellular water motion within this tissue is restricted. So when the tumor cellularity is increased, the water diffusion is decreased, which means the ADC is also decreased.<sup>11</sup> Generally, necrosis or cellular lysis, caused by antitumor treatments, can decrease cellularity.<sup>12</sup> Because decreased tumor cell population precedes any measurable tumor size change, DWI can thus predict early treatment outcomes, monitor early treatment response, and detect recurrent tumors.<sup>13</sup>

Shifts in the ADC symbolizes certain long-term patient responses to treatments.<sup>26</sup> For example, the degree of change in ADC after chemotherapy is predictive of patient's overall survival.<sup>27</sup> Changes in ADC might also help identify chemotherapy-resistant tumor types.<sup>27</sup> Therefore, ADC is a valuable tool in monitoring the patients' treatment response.<sup>27</sup> However, when oncologists analyze pretreatment ADC in recurrent glioblastomas, they found that ADC is effective in predicting antiangiogenic therapy's response, but not chemotherapy's response.<sup>28</sup> Therefore, monitoring ADC over the course of treatment serves as a tracker for prescription of antiangiogenic therapy in recurrent glioblastoma but perhaps not when utilizing chemotherapy alone.<sup>28</sup>

## Apparent Diffusion Coefficient's Inaccurate Representation of Tumor Cellularity

However, this technique is not without limitation. Since ADC is affected by capillaries' microcirculation, it may be susceptible to changes in vascularization.<sup>22</sup> Malignant brain tumors express higher tumor cellularity and vascularity compared to benign brain tumors.<sup>23</sup> Unlike the high tumor vascularity, which increases ADC, high tumor cellularity decreases it.<sup>24</sup> This phenomenon proves that the DWI signal attenuation in hypervascular brain tumors can be influenced by two opposing manners, which, as mentioned previously, limits the ADC value for grading hypervascular brain tumors.<sup>25</sup> This under-performance can give rise to contradictory results. To account for this, tumor vascularity can be assessed through a pathology review of tumor sections.<sup>22</sup>

Moreover, different tumor subtypes may interfere with the use of ADC. High-grade gliomas, which are more invasive and harder to treat compared to low-grade gliomas, are heterogeneous regarding their microstructure and genetics.<sup>15</sup> High DWI signals can result from tumor coagulation necrosis or ischemia.<sup>16</sup> However, highly cellular tumor areas in combo with inflammatory processes can dramatically restrict the intensity of the diffusion signal, adding complexity to the characterization of tumor progression.<sup>16</sup> As a result, the usage of mean ADC values to accurately grade tumors can be ineffective due to this heterogeneity,<sup>17</sup> for instance, tumor necrosis can restrict water diffusion and falsely suggest high cellularity; indeed, tumor cell density, metabolic activity, ischemia, and compression are all possible factors responsible for the restriction of diffusion within and around cancerous tissue.<sup>20</sup>

## Intravoxel Incoherent Motion to Differentiate Perfusion from Diffusion

A pseudo diffusion, often observed in abnormally oriented capillary blood flow, can interfere with diffusion measures in DWI images. This occurs only when the *beta* parameter of a DW image are set to relatively small values.<sup>29</sup> To distinguish true diffusion from pseudo diffusion, we can identify the isotropic diffusion in water from the "incoherent motion" present in blood vessels.<sup>30,31</sup> This can be done by taking multiple DWI images while systematically altering their b values corresponding to the weighting of diffusion (where higher b values result in higher-intensity images given a constant degree of diffusion). For each voxel, the b value used across multiple images and the resulting intensity of the voxel can be fit to previously-defined models which differentiate free water diffusion from that observed in blood.

Multiple forms of these models have been employed. Mono-exponential fitting involves the use of a single variable while bi-exponential fitting involves the use of multiple variables. In recent research, scientists, using simple mono-exponential fitting, which doesn't account for the contribution of the perfusion effect, discovered that

lymphoma patients have drastically lower ADC compared to other tumor patient groups.<sup>40</sup> Meanwhile, scientists, using the bi-exponential fitting, which accounts for the contribution of the perfusion effect, discovered that lymphoma patients have similar “true” diffusion parameters compared to other tumor patient groups.<sup>40</sup> This finding suggests that the ADC difference is strongly correlated to the perfusion effect’s contribution and perfusion of blood may sometimes produce spurious differences between groups when not taken into account.<sup>41</sup> It can therefore be concluded that a mono exponentially fitted ADC may not give oncologists the best accuracy in terms of the inverse correlation between the ADC and tumor cellularity.<sup>42,43</sup>

However, IVIM also has its limitations. For example, it requires a high signal-to-noise ratio, when oncologists use it to separate perfusion from diffusion.<sup>36</sup> Furthermore, the phenomena of vascular tubular flow and glandular secretion can produce artifacts.<sup>37</sup> Lastly, different vessel sizes can produce different IVIM signals and result in different levels of sensitivity.<sup>37</sup> All of these challenges makes the differentiation process that much more difficult and future efforts should address these remaining limitations.

## Diffusion Kurtosis Imaging Reflects Gray and White Matter’s Structural Components

The Gaussian law can be easily observed in free, unobstructed, diffusion.<sup>44</sup> Unsurprisingly, a non-Gaussian distribution occurs when the diffusion of water molecules are restrictively affected by the complex microstructure of different tissue types.<sup>44</sup> Therefore, the tissue’s microstructure, namely the cell membranes, organelles, and water compartments, could give rise to specific non-Gaussian diffusion patterns.<sup>44</sup>

DKI is a modified version of DTI. Compared to diffusion tensor imaging (DTI), non-Gaussian diffusion, derived from diffusion kurtosis imaging (DKI), can more accurately evaluate aspects of both normal and pathologic tissue by taking advantage of this property.<sup>45</sup> The microstructural complexity index might thus be represented by the mean kurtosis (mk).<sup>46</sup> Since MK works regardless of whether the tissue is spatially oriented in a certain plain, it proves to be a better venue compared to DTI-derived fractional anisotropy (FA) in the context of neurooncology.<sup>46</sup> For instance, MK can be applied for both the gray and white matter without regard to the directionality of the underlying healthy cytoarchitecture.

## Diffusion Kurtosis Imaging Reliably Grades Glioma

Increased kurtosis is indicative of increased tissue complexity in high-grade glioma.<sup>47</sup> A variety of events such as hemorrhage, tumor invasion, necrosis, endothelial proliferation and more, could be involved in increasing the tissue complexity of tumors.<sup>47</sup> Unsurprisingly, decreased kurtosis parameters are indicative of decreased tissue complexity. This may be used to detect low-grade glioma, which have more homogeneous and less packed cells.<sup>47</sup> Studies have pinpointed that the difference between the intra- and extracellular space can be used to identify low-grade from high-grade gliomas.<sup>48</sup> This difference might be attributed to the different characteristics between these forms of gliomas.<sup>49</sup> High-grade gliomas have more crowding cells and myelin breakdown products, which makes the membrane structure tightly packed.<sup>49</sup> Low-grade gliomas have more differentiated, neoplastic astrocytes, which makes the membrane structure loosely packed.<sup>49</sup> Thus, while both high-grade and low-grade gliomas have more kurtosis than normal tissue, differential kurtosis within tumor tissue may provide clues to which subtype best characterizes a tumor in the absence of a biopsy.

## Dynamic susceptibility contrast perfusion MRI can measure CBV

Dynamic susceptibility contrast (DSC) perfusion MRI operates similarly to the popular T2\*-weighted MR techniques (e.g., ASL, discussed below) that are often used for the estimation of rCBV in functional MRI.<sup>55,56</sup> However, DSC perfusion MRI instead uses gadolinium-based contrast agents in order to increase signal to noise ratio. This is useful because rCBV can be used to predict tumor vascular morphometry, such as MVA (discussed below).<sup>57</sup> However, it should be noted that rCBV can’t accurately predict such measures when the glioblastomas have heterogeneous vessel sizes, which is often the case.

One major problem to consider is contrast agent leakage, which often occurs in tumor vessels when the blood-brain barrier is severely sabotaged.<sup>58</sup> This can amplify the abnormal effects on T1 or T2\*.<sup>59</sup> The effects T2\* exerted as a

consequence of contrast-agent leakage can be influenced by the tumor cells' density and spatial distribution.<sup>59</sup> Moreover, these effects could result in the addition of a susceptibility calibration factor, which could partly, but not fully, compensate for the leakage effects.<sup>59</sup> Due to these limitations of T2, T1 kinetic parameters are more reliable when it comes to analyzing complex tumor vessels with mostly heterogeneous vascular characteristics as these tumors are more likely to present with blood-brain barrier damage.<sup>60</sup>

## DSC Perfusion MRI Eliminates the Problem of Pseudoprogession

After glioblastomas patients were treated with chemoradiotherapy, oncologists took into account the presence of transiently enlarging, contrast-enhanced lesions in order to decide whether to continue or switch to a second-line therapy.<sup>61</sup> Pseudoprogession is the detected expansion of a tumor that is not caused by the tumor's actual growth. In other words, it's false tumor growth. It occurs because during immunotherapy, immune cells surround the tumor and enlarge the region. Inaccurate interpretation of the pseudoprogession in tumors have reduced salvage treatment trials.<sup>62</sup> To prevent the false-positive evaluation of a new drug's effects, it is therefore necessary to exclude pseudoprogession.<sup>63</sup> Oncologists often rely on the histopathologic diagnosis, which is derived from second-look surgery and highly prone to sampling errors, to separate the early tumor progression cases from the pseudoprogession ones.<sup>64</sup> To limit these sampling errors, alternative methods have been integrated, namely the interpretation of MRI findings and clinical manifestations.<sup>65</sup> Dynamic susceptibility contrast perfusion MRI is a specific alternative that researchers have looked to.<sup>66</sup>

Previous studies show that we can predict the patient's chance of one-year survival based on how their rCBV percentage has changed following radiation-temozolomide therapy.<sup>67</sup> Scientists have developed a tool to differentiate pseudoprogession from true progression: a rCBV-derived parametric response map.<sup>68</sup> It was also found that true tumor progression can be expected if the patients experience a rCBV decrease three weeks after therapy.<sup>69</sup> After chemoradiotherapy, early tumor progression patients experience something the pseudoprogession patients did not: negative changes of skewness and kurtosis of rCBV histograms<sup>70</sup> (rCBV histograms shows the distribution of normalized rCBV values over time). In short, as vascular proliferation remains a prominent element indicative of true tumor progression, DSC perfusion MRI likely holds value in the detection of true tumor progression out of many pseudo progressions, despite its caveats.<sup>72</sup>

## Microvessel Density/ Area

Compared to MVD (microvessel density), MVA (microvessel area) shows both the microvessels' density and character, which gives oncologists a better evaluation of the tumor's microvessel morphometric complexity, overall vascular surface area, and stages of angiogenesis.<sup>51</sup> For instance, an increase in MVD suggests a decrease in vessel size. There are many explanations for this phenomena such as the presence of delicate microvessels or glomeruloid vascular structure (made of tumor-derived immature microchannels) in invasive tumors in the brain's grey matter.<sup>52</sup> The field has thus turned to MVA as a better predictor for patient survival than MVD, which has less ability to predict tumor progression.<sup>53</sup> Specifically, in high MVA tumors, glomeruloid vessels are more common, while low MVA tumors possess more delicate (capillary-like) vessels.<sup>53</sup> Importantly, such high MVA tumors are more likely to undergo metastasis or invasion. This suggests that prognosing a glioma patient involves the examination of their tumor morphology and techniques such as MVA, which can do so without surgery, may non-invasively provide guidance for treatment options (e.g., the decision to surgically remove tumors).

## Arterial Spin Labeling to Predict Tumor Normalization and Drug Response

As stated, tumor vascular characteristics can be indicative of certain therapeutic responses.<sup>91</sup> This is due, in part, to the fact that particular tumor vessels' patterns play a big role in increasing or decreasing the efficacy of chemotherapeutic drug delivery.<sup>92</sup> For instance, higher tumor blood perfusion is linked to more positive outcomes regarding antiangiogenic treatment.<sup>93</sup> Thus, high CBF, which may be assessed using arterial spin labeling (ASL), signifies low tumor vessels' permeability.<sup>94</sup> This makes the chemotherapeutic drug delivery to tumor cells more efficient and effective, giving a more favorable outcome.<sup>95</sup> Leveraging the endogenous tracer without any contrast agent injection, ASL noninvasively measures CBF<sup>96</sup> (An endogenous tracer is a molecule or subatomic particles that comes from within the system that are used to track another molecule. ASL's endogenous tracer is arterial blood

water protons tagged by a radio-frequency pulse prior to the blood entering the cerebrum). Aside from accurately evaluating the CBF, ASL is also valuable for assessing tumor blood-vessel attenuation and grading gliomas alongside other methods like MVA.<sup>96</sup> Due to all of its capability, ASL is highly helpful in cases where CBF is correlated to clinical outcome measures, such as in the context of drug delivery through the blood stream.<sup>96</sup>

## Arterial Spin Labeling Depicts Efficacy of Drug Delivery

The structure and function of tumor vessels drastically change in angiogenesis.<sup>97</sup> By inhibiting vascular endothelial growth factor signaling, antiangiogenic treatment eradicates mutated vessels and reestablishes the normally-functioning vasculature.<sup>98</sup> This reestablishment of vasculature promotes effective drug delivery as it increases CBF and decreases tumor-induced hypoxia and interstitial fluid pressure, which may kill nearby cells.<sup>99</sup> So, increased tumor perfusion during chemotherapy might serve as a good signal of longer survival.<sup>100</sup> Cytotoxic chemotherapeutic agents showed different effectiveness in high-CBF patients versus low-CBF patients.<sup>101</sup> The high-CBF groups experience a longer median time-to-progression (TTP) compared to their negative-CBF counterparts.<sup>102</sup> High CBF can thus be used to predict favorable TTP and outcomes. Moreover, this is regardless of the MGMT promoter methylation status;<sup>102</sup> MGMT is an enzyme that helps tumors resist chemotherapy and its methylation-dependent expression thus impacts drug-treatment. In sum, high CBF demonstrates hyperperfusion and normalized tumor vessels, which may enhance drug delivery.<sup>102</sup>

## Dynamic Contrast-Enhanced Perfusion MRI Detects Immature Vessels

While leakage of contrast agents may limit techniques like DSC, it may also be used to assess the vascular architecture of tumors<sup>73</sup>. Naturally, the intravascular compartment and the extravascular, extracellular compartment interchange contrast agents with each other at some rate.<sup>74</sup> To quantify the rate of this exchange, Dynamic Contrast-Enhanced (DCE) perfusion MRI uses a pharmacokinetic model, which can derive the transfer coefficient (K<sub>trans</sub>), which is indicative of the tumor's vessel permeability.<sup>75</sup> Brain tumors' contrast agents usually extravasates (leaks out from blood vessels) due to the presence of immature hyperpermeable vessels.<sup>76</sup> Measuring this extravasation is useful in two ways: differentiating mature tumor vessels from immature ones and identifying the tumor-vessel permeability<sup>77</sup>; both can be utilized as a biomarker for brain tumor progression.<sup>77</sup>

DCE perfusion MRI, in order to function, requires complex data acquisition and analysis that the DSC perfusion MRI does not.<sup>78</sup> First, determination of T1 values in brain tissue before contrast injection is required for the calculation of tissue contrast concentration curve with time.<sup>79</sup> After that, scientists need to accurately measure the arterial input function, which is the concentration of tracer (molecules with specific characteristics that allow them to trace a biological process) in the artery's blood.<sup>80</sup> This can be very challenging because inflow might disrupt either the MR signal intensity or absolute contrast concentration.<sup>80</sup>

Despite the required complexity, DCE MRI also has many advantages over other methods like DSC: three-dimensional acquisition of images, a higher signal-to-noise ratio, and a higher spatial resolution compared to DSC MRI.<sup>81</sup> Also, since DCE MRI can more sensitively monitor small vessels compared to DSC MRI, it could better track drug delivery.<sup>81</sup> In theory, DCE perfusion MRI also has other potentials such as the promise of developing more accurate pharmacokinetic models that can be used to modulate tumor drug delivery and patient's response to chemotherapeutic drugs based on estimates of tumor vascularity detected using other imaging techniques.<sup>82</sup> Ultimately, DCE perfusion MRI can be extremely helpful if used correctly and when its limitations are properly taken into account.<sup>83</sup>

## DCE Perfusion MRI Improves Assessment of Patient's Drug Response

Oncologists also use dynamic contrast-enhanced perfusion MRI to evaluate the physiologic aspect of the regeneration of tumor vessels, namely the microcirculation.<sup>84,85</sup> Dynamic contrast-enhanced perfusion MRI is extremely useful because it can serve as a noninvasive tracker of not only tumor progression, but also treatment response.<sup>86,87</sup> Researchers have found that the combo method of DSC/DCE, MRI, and DWI performs better in differentiating recurrent glioblastoma from radiation necrosis compared to the combo the use of only MRI and DWI.<sup>88</sup> In other words, the addition of DSC or DCE improved diagnostic performance. Oncologists are encouraged

to include any form of perfusion MRI to their traditional MRI protocol of MRI and DWI, since perfusion MRI increases accuracy in the recognition of recurrent glioblastoma.<sup>89</sup> More importantly, the specific combination of conventional MRI, DWI, and DCE MRI (not DSC MRI) proves to have the best recurrent glioblastoma diagnostic performance.<sup>90</sup>

## CE-SWI Highlights Tumor Necrosis and Vessels

Susceptibility-weighted images (SWI) are gradient echo sequences that are able to illustrate the cerebral veins and microhemorrhage.<sup>103</sup> SWI describes edema and contrast enhancement, examples of T2 effects related to T1 effects.<sup>104</sup> The capture of the tumor's architecture is necessary to evaluate tumor necrosis.<sup>105</sup> The tumor's architecture shown on SWI is much more useful compared to that shown on contrast-enhanced T1-weighted imaging because contrast-enhanced T1 imaging captures the tumor's architecture based on the presence of necrosis, cysts, and tumor boundaries, while SWI captures the tumor's architecture based on the presence of blood products and tumor vessels.<sup>106,107</sup>

When analyzing brain mass lesions using susceptibility signals, gadolinium-based contrast agents can be integrated into the procedure to improve the analysis.<sup>108</sup> SWI and contrast-enhanced (CE)-SWI can show similar susceptibility signals. But, there are cases when only the CE-SWI shows a particular susceptibility signal, meriting the use of contrast in some patients. Importantly, if oncologists use SWI before and after the contrast agent is applied, oncologists can more easily differentiate hemorrhages from veins.<sup>109</sup> This is because the signal intensity of blood vessels changes from the absence to presence of contrast agent while the signal intensity of hemorrhages, which take much longer to take on contrast agents, remains the same.<sup>109</sup> CE-SWI can be seen as a high-resolution, structural MRI, which has a great potential for contrast-enhancing tumor segmentation.<sup>110</sup>

Previous studies examining the effectiveness of CE-SWI have also shown that it can be used to detect tumor invasion zones.<sup>111</sup> These are zones that experience less tumor-cell density because tumor cells migrate away from these zones into the surrounding brain tissue for invasion.<sup>112</sup> Since this process can be fatal to patients, CE-SWI can be extremely helpful in detecting these processes. In pre-contrast SWI, susceptibility signals may detect highly pathological vessels, micro-hemorrhage, and extensive necrosis; however, CE is necessary for the detection of more subtle qualities such as tumor invasion zones.<sup>113</sup>

## CE SWI Enhance Tumor Evaluation

SWI is a necessary addition to conventional imaging techniques because it contrasts and detects the tumor's venous vasculature and hemorrhage, which could not be done using conventional imaging techniques.<sup>114</sup> By tracking the intratumoral susceptibility signal (ITSS), oncologists have used SWI to non-invasively grade primary brain tumors.<sup>115</sup> Tumor grading is a process of identifying the levels of malignancy of the tumor base on its characteristics. Glioblastomas are high-grade tumors that consist of hemorrhage and upregulated micro vascularity.<sup>116</sup> SWI is valuable because it can detect the presence of these components that traditional imaging methods can't. There is a strong link between how strong the ITSS is and what the maximum rCBV is in the same tumor segments.<sup>117</sup> However, this correlation of ITSS and maximum rCBV also varies across different patients.<sup>117</sup> Since SWI gives off the strongest ITSS in glioblastoma, ITSS might be a useful tool for oncologists to make accurate glioma diagnosis. Researchers found out that SWI and DSC perfusion MRI have similar diagnostic and grading performance.<sup>118</sup> Again, compared to non-contrast SWI, contrast-enhanced SWI performs better in tumor evaluation.<sup>119</sup>

## Conclusion

As it stands, the use of conventional MRI in neuro-oncology faces diagnostic difficulties that may be solved by emerging imaging techniques which, specifically, may better assess tumor subtypes and, especially, response to treatment.

Nonetheless, the emerging MRI techniques discussed here have unique limitations and challenges. For example, ADC aren't stable for all types of tumors and treatments. For example, it works differently for antiangiogenic

therapy versus chemotherapy. Similarly, IVIM is sensitive to artifacts and different vessel sizes. DSC uses contrast agents, but contrast agents leakage often occurs, thus creating problems. DCE requires more prerequisites compared to DSC to be properly used, which makes its usage highly rigorous. Fortunately, ASL, the third perfusion MRI technique, CE SWI, and DKI all face insignificant obstacles.

Regarding these 7 cutting-edge techniques covered, their usefulness and limitations overlap with or accompany each other, suggesting that a variety of these techniques may provide the best picture of a patient's condition. For instance, ADC reflects diffusion, which is how well water flows in the patient's brain. This is different from perfusion, how well blood flows in the patient's brain. So to differentiate the two, IVIM could be used. It's important to differentiate the two because they provide different suggestions for oncologists. High diffusion means low cellularity, which means less malignant tumors. On the other hand, perfusion gives different measurements such as CBF, CBV, K-trans, and TTP, that reflects characteristics of blood vessels, which reflect how well a drug will perform. The three perfusion MRI techniques discussed here: DSC, DCE, and ASL, are all useful for predicting the efficacy of drug administration. DSC eliminates pseudoprogression which is important because less pseudoprogression means less false-positives evaluation on drug effects. DCE estimates immature vessels which is useful in predicting effectiveness of drug delivery. ASL provides CBF, which evaluates perfusion, which predicts effectiveness of drug delivery. Lastly, DKI and CE SWI can both be used to classify tumors, high-grade or low-grade. Thus, a consortium of advanced MRI techniques is likely to bring about the most effective treatment of brain cancer, from detection to remission.

## Acknowledgements

I would like to thank Joshua Cain for being my mentor and working rigorously alongside me to guide me to the completion of this paper.

## References

1. Luo, M., Zhang, L., Jiang, X. H., & Zhang, W. D. (2017). Intravoxel Incoherent Motion Diffusion-weighted Imaging: Evaluation of the Differentiation of Solid Hepatic Lesions. *Translational oncology*, 10(5), 831–838. <https://doi.org/10.1016/j.tranon.2017.08.003>
2. Jensen, J. H., & Helpert, J. A. (2010). MRI quantification of non-Gaussian water diffusion by kurtosis analysis. *NMR in biomedicine*, 23(7), 698–710. <https://doi.org/10.1002/nbm.1518>
3. Essig, M., Shiroishi, M. S., Nguyen, T. B., Saake, M., Provenzale, J. M., Enterline, D., Anzalone, N., Dörfler, A., Rovira, A., Wintermark, M., & Law, M. (2013). Perfusion MRI: the five most frequently asked technical questions. *AJR. American journal of roentgenology*, 200(1), 24–34. <https://doi.org/10.2214/AJR.12.9543>
4. Lindner, T., Ahmeti, H., Juhasz, J., Helle, M., Jansen, O., Synowitz, M., & Ulmer, S. (2018). A comparison of arterial spin labeling and dynamic susceptibility perfusion imaging for resection control in glioblastoma surgery. *Oncotarget*, 9(26), 18570–18577. <https://doi.org/10.18632/oncotarget.24970>
5. Muto, M., Frauenfelder, G., Senese, R., Zeccolini, F., Schena, E., Giurazza, F., & Jäger, H. R. (2018). Dynamic susceptibility contrast (DSC) perfusion MRI in differential diagnosis between radionecrosis and neoangiogenesis in cerebral metastases using rCBV, rCBF and K2. *La Radiologia medica*, 123(7), 545–552. <https://doi.org/10.1007/s11547-018-0866-7>
6. Padhani A.R. Dynamic contrast-enhanced MRI in clinical oncology: Current status and future directions. *J Magn Reson Imaging*. 2002; 16: 407-422

7. Wierenga, C. E., Hays, C. C., & Zlatar, Z. Z. (2014). Cerebral blood flow measured by arterial spin labeling MRI as a preclinical marker of Alzheimer's disease. *Journal of Alzheimer's disease : JAD*, 42 Suppl 4(Suppl 4), S411–S419. <https://doi.org/10.3233/JAD-141467>
8. Ma, X., Bai, Y., Lin, Y. et al. Amide proton transfer magnetic resonance imaging in detecting intracranial hemorrhage at different stages: a comparative study with susceptibility weighted imaging. *Sci Rep* 7, 45696 (2017). <https://doi.org/10.1038/srep45696>
9. Sener R. N. (2001). Diffusion MRI: apparent diffusion coefficient (ADC) values in the normal brain and a classification of brain disorders based on ADC values. *Computerized medical imaging and graphics : the official journal of the Computerized Medical Imaging Society*, 25(4), 299–326. [https://doi.org/10.1016/s0895-6111\(00\)00083-5](https://doi.org/10.1016/s0895-6111(00)00083-5)
10. Chilla, G. S., Tan, C. H., Xu, C., & Poh, C. L. (2015). Diffusion weighted magnetic resonance imaging and its recent trend-a survey. *Quantitative imaging in medicine and surgery*, 5(3), 407–422. <https://doi.org/10.3978/j.issn.2223-4292.2015.03.01>
11. Surov, A., Meyer, H. J., & Wienke, A. (2017). Correlation between apparent diffusion coefficient (ADC) and cellularity is different in several tumors: a meta-analysis. *Oncotarget*, 8(35), 59492–59499. <https://doi.org/10.18632/oncotarget.17752>
12. Desmoulière, A., Redard, M., Darby, I., & Gabbiani, G. (1995). Apoptosis mediates the decrease in cellularity during the transition between granulation tissue and scar. *The American journal of pathology*, 146(1), 56–66.
13. Karameshev, A., Arnold, M., Schroth, G., Kappeler, L., Stein, P., Gralla, J., Brekenfeld, C., Findling, O., Mono, M. L., De Marchis, G. M., Fischer, U., Mattle, H. P., Nedeltchev, K., & El-Koussy, M. (2011). Diffusion-weighted MRI helps predict outcome in basilar artery occlusion patients treated with intra-arterial thrombolysis. *Cerebrovascular diseases (Basel, Switzerland)*, 32(4), 393–400. <https://doi.org/10.1159/000330644>
14. A, R., M J, T. B., N, C., M, S., M Gh, H., & Gh, A. (2019). Signal Intensity of High B-value Diffusion-weighted Imaging for the Detection of Prostate Cancer. *Journal of biomedical physics & engineering*, 9(4), 453–458. <https://doi.org/10.31661/jbpe.v0i0.811>
15. Salloum, R., McConechy, M. K., Mikael, L. G., Fuller, C., Drissi, R., DeWire, M., Nikbakht, H., De Jay, N., Yang, X., Boue, D., Chow, L., Finlay, J. L., Gayden, T., Karamchandani, J., Hummel, T. R., Olshefski, R., Osorio, D. S., Stevenson, C., Kleinman, C. L., Majewski, J., ... Jabado, N. (2017). Characterizing temporal genomic heterogeneity in pediatric high-grade gliomas. *Acta neuropathologica communications*, 5(1), 78. <https://doi.org/10.1186/s40478-017-0479-8>
16. White, N. S., McDonald, C., Farid, N., Kuperman, J., Karow, D., Schenker-Ahmed, N. M., Bartsch, H., Rakow-Penner, R., Holland, D., Shabaik, A., Bjørnerud, A., Hope, T., Hattangadi-Gluth, J., Liss, M., Parsons, J. K., Chen, C. C., Raman, S., Margolis, D., Reiter, R. E., Marks, L., ... Dale, A. M. (2014). Diffusion-weighted imaging in cancer: physical foundations and applications of restriction spectrum imaging. *Cancer research*, 74(17), 4638–4652. <https://doi.org/10.1158/0008-5472.CAN-13-3534>
17. Azzam, H., Mansour, S., Salem, N. et al. Correlative study between ADC value and grading of invasive breast cancer. *Egypt J Radiol Nucl Med* 51, 12 (2020). <https://doi.org/10.1186/s43055-019-0124-6>
18. Lätt, J., Nilsson, M., van Westen, D., Wirestam, R., Ståhlberg, F., & Brockstedt, S. (2009). Diffusion-weighted MRI measurements on stroke patients reveal water-exchange mechanisms in sub-acute

ischaemic lesions. *NMR in biomedicine*, 22(6), 619–628. <https://doi.org/10.1002/nbm.1376>

19. Institute of Medicine (US) Committee on Military Nutrition Research. The Role of Protein and Amino Acids in Sustaining and Enhancing Performance. Washington (DC): National Academies Press (US); 1999. 14, Amino Acid and Protein Requirements: Cognitive Performance, Stress, and Brain Function. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK224629/>
20. Finelli P. F. (2012). Diagnostic approach to restricted-diffusion patterns on MR imaging. *Neurology. Clinical practice*, 2(4), 287–293. <https://doi.org/10.1212/CPJ.0b013e318278bee1>
21. Hayashida, Y., Hirai, T., Morishita, S., Kitajima, M., Murakami, R., Korogi, Y., Makino, K., Nakamura, H., Ikushima, I., Yamura, M., Kochi, M., Kuratsu, J. I., & Yamashita, Y. (2006). Diffusion-weighted imaging of metastatic brain tumors: comparison with histologic type and tumor cellularity. *AJNR. American journal of neuroradiology*, 27(7), 1419–1425.
22. Abdullah, O. M., Gomez, A. D., Merchant, S., Heidinger, M., Poelzing, S., & Hsu, E. W. (2016). Orientation dependence of microcirculation-induced diffusion signal in anisotropic tissues. *Magnetic resonance in medicine*, 76(4), 1252–1262. <https://doi.org/10.1002/mrm.25980>
23. Klatzo I. Presidential address: neuropathological aspects of brain edema. *J Neuropathol Exp Neurol* 1967;26:1–14
24. Messina, C., Bignone, R., Bruno, A., Bruno, A., Bruno, F., Calandri, M., Caruso, D., Coppolino, P., Robertis, R., Gentili, F., Grazzini, I., Natella, R., Scalise, P., Barile, A., Grassi, R., & Albano, D. (2020). Diffusion-Weighted Imaging in Oncology: An Update. *Cancers*, 12(6), 1493. <https://doi.org/10.3390/cancers12061493>
25. Pauleit D, Langen KJ, Floeth F, et al. Can the apparent diffusion coefficient be used as a noninvasive parameter to distinguish tumor tissue from peritumoral tissue in cerebral gliomas? *J Magn Reson Imaging* 2004;20:758–64
26. Zhang, M., Chen, Y., Cong, X., & Zhao, X. (2018). Utility of intravoxel incoherent motion MRI derived parameters for prediction of aggressiveness in urothelial bladder carcinoma. *Journal of magnetic resonance imaging : JMRI*, 48(6), 1648–1656. <https://doi.org/10.1002/jmri.26165>
27. Suh, J. Y., Cho, G., Song, Y., Lee, C. K., Kang, J. S., Kang, M. R., Park, S. B., Kim, Y. R., & Kim, J. K. (2012). Is apparent diffusion coefficient reliable and accurate for monitoring effects of antiangiogenic treatment in a longitudinal study?. *Journal of magnetic resonance imaging : JMRI*, 35(6), 1430–1436. <https://doi.org/10.1002/jmri.23574>
28. Schneider, M. J., Cyran, C. C., Nikolaou, K., Hirner, H., Reiser, M. F., & Dietrich, O. (2014). Monitoring early response to anti-angiogenic therapy: diffusion-weighted magnetic resonance imaging and volume measurements in colon carcinoma xenografts. *PloS one*, 9(9), e106970. <https://doi.org/10.1371/journal.pone.0106970>
29. Wu, W., & Miller, K. L. (2017). Image formation in diffusion MRI: A review of recent technical developments. *Journal of magnetic resonance imaging : JMRI*, 46(3), 646–662. <https://doi.org/10.1002/jmri.25664>
30. Li, H., Liang, L., Li, A., Hu, Y., Hu, D., Li, Z., & Kamel, I. R. (2017). Monoexponential, biexponential, and stretched exponential diffusion-weighted imaging models: Quantitative biomarkers for differentiating renal clear cell carcinoma and minimal fat angiomyolipoma. *Journal of magnetic resonance imaging : JMRI*, 46(1), 240–247. <https://doi.org/10.1002/jmri.25524>

31. Yang DM, Kim HC, Kim SW, Jahng GH, Won KY, Lim SJ, et al. Prostate cancer: correlation of intravoxel incoherent motion MR parameters with gleason score. *Clin Imaging*. (2016) 40:445–50. doi: 10.1016/j.clinimag.2016.01.001
32. Zhang YD, Wang Q, Wu CJ, Wang XN, Zhang J, Liu H, et al. The histogram analysis of diffusion-weighted intravoxel incoherent motion (IVIM) imaging for differentiating the gleason grade of prostate cancer. *Eur Radiol*. (2015) 25:994–1004. doi: 10.1007/s00330-014-3511-4
33. Beyhan M, Sade R, Koc E, Adanur S, Kantarci M. The evaluation of prostate lesions with IVIM DWI and MR perfusion parameters at 3T MRI. *Radiol Med*. (2019) 124:87–93. doi: 10.1007/s11547-018-0930-3
34. Calamante F. (2013). Arterial input function in perfusion MRI: a comprehensive review. *Progress in nuclear magnetic resonance spectroscopy*, 74, 1–32. <https://doi.org/10.1016/j.pnmrs.2013.04.002>
35. Hectors, S. J., Wagner, M., Besa, C., Bane, O., Dyvorne, H. A., Fiel, M. I., Zhu, H., Donovan, M., & Taouli, B. (2016). Intravoxel incoherent motion diffusion-weighted imaging of hepatocellular carcinoma: Is there a correlation with flow and perfusion metrics obtained with dynamic contrast-enhanced MRI?. *Journal of magnetic resonance imaging : JMIR*, 44(4), 856–864. <https://doi.org/10.1002/jmri.25194>
36. Maximov, I. I., & Vellmer, S. (2019). Isotropically weighted intravoxel incoherent motion brain imaging at 7T. *Magnetic resonance imaging*, 57, 124–132. <https://doi.org/10.1016/j.mri.2018.11.007>
37. Wang, J., Fernández-Seara, M. A., Wang, S., & St Lawrence, K. S. (2007). When perfusion meets diffusion: in vivo measurement of water permeability in human brain. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism*, 27(4), 839–849. <https://doi.org/10.1038/sj.jcbfm.9600398>
38. Pang Y, Turkbey B, Bernardo M, Kruecker J, Kadoury S, Merino MJ, et al. Intravoxel incoherent motion MR imaging for prostate cancer: an evaluation of perfusion fraction and diffusion coefficient derived from different b-value combinations. *Magn Reson Med*. (2013) 69:553–62. doi: 10.1002/mrm.24277
39. Mao, W., Zhou, J., Zeng, M., Ding, Y., Qu, L., Chen, C., Ding, X., Wang, Y., Fu, C., & Gu, F. (2018). Intravoxel incoherent motion diffusion-weighted imaging for the assessment of renal fibrosis of chronic kidney disease: A preliminary study. *Magnetic resonance imaging*, 47, 118–124. <https://doi.org/10.1016/j.mri.2017.12.010>
40. Jiang J, Fu Y, Hu X, Cui L, Hong Q, Gu X, et al. The value of diffusion-weighted imaging based on monoexponential and biexponential models for the diagnosis of benign and malignant lung nodules and masses. *Br J Radiol*. (2020) 93:20190400. doi: 10.1259/bjr.20190400
41. Song M, Yue Y, Jin Y, Guo J, Zuo L, Peng H, et al. Intravoxel incoherent motion and ADC measurements for differentiating benign from malignant thyroid nodules: utilizing the most repeatable region of interest delineation at 3.0 T. *Cancer Imaging*. (2020) 20:9. doi: 10.1186/s40644-020-0289-2
42. Chatterjee A, Watson G, Myint E, Sved P, McEntee M, Bourne R. Changes in epithelium, stroma, and lumen space correlate more strongly with gleason pattern and are stronger predictors of prostate ADC changes than cellularity metrics. *Radiology*. (2015) 277:751–62. doi: 10.1148/radiol.2015142414
43. Dolgorsuren, E. A., Harada, M., Kanazawa, Y., Abe, T., Otomo, M., Matsumoto, Y., Mizobuchi, Y., & Nakajima, K. (2019). Correlation and Characteristics of Intravoxel Incoherent Motion and Arterial Spin Labeling Techniques Versus Multiple Parameters Obtained on Dynamic Susceptibility Contrast Perfusion MRI for Brain Tumors. *The journal of medical investigation : JMI*, 66(3.4), 308–313. <https://doi.org/10.2152/jmi.66.308>

44. Cerebral gliomas: diffusional kurtosis imaging analysis of microstructural differences. Raab P, Hattingen E, Franz K, Zanella FE, Lanfermann H. *Radiology*. 2010 Mar; 254(3):876-81.
45. Gliomas: diffusion kurtosis MR imaging in grading. Van Cauter S, Veraart J, Sijbers J, Peeters RR, Himmelreich U, De Keyser F, Van Gool SW, Van Calenbergh F, De Vleeschouwer S, Van Hecke W, Sunaert S. *Radiology*. 2012 May; 263(2):492-501.
46. Wu, E. X., & Cheung, M. M. (2010). MR diffusion kurtosis imaging for neural tissue characterization. *NMR in biomedicine*, 23(7), 836–848. <https://doi.org/10.1002/nbm.1506>
47. Jiang, Rifeng et al. "Diffusion kurtosis imaging can efficiently assess the glioma grade and cellular proliferation." *Oncotarget* vol. 6,39 (2015): 42380-93. doi:10.18632/oncotarget.5675
48. Grading of astrocytomas. A simple and reproducible method. Daumas-Duport C, Scheithauer B, O'Fallon J, Kelly P. *Cancer*. 1988 Nov 15; 62(10):2152-65.
49. Larsen, Jennifer et al. "Low grade glioma: an update for radiologists." *The British journal of radiology* vol. 90,1070 (2017): 20160600. doi:10.1259/bjr.20160600
50. Momeni, Farideh et al. "Differentiating Between Low- and High-grade Glioma Tumors Measuring Apparent Diffusion Coefficient Values in Various Regions of the Brain." *Oman medical journal* vol. 36,2 e251. 31 Mar. 2021, doi:10.5001/omj.2021.59
51. Seifi, S., Shafaie, S., & Ghadiri, S. (2011). Microvessel density in follicular cysts, keratocystic odontogenic tumours and ameloblastomas. *Asian Pacific journal of cancer prevention : APJCP*, 12(2), 351–356.
52. Rojiani, A. M., & Dorovini-Zis, K. (1996). Glomeruloid vascular structures in glioblastoma multiforme: an immunohistochemical and ultrastructural study. *Journal of neurosurgery*, 85(6), 1078–1084. <https://doi.org/10.3171/jns.1996.85.6.1078>
53. Hu, L S et al. "Correlations between perfusion MR imaging cerebral blood volume, microvessel quantification, and clinical outcome using stereotactic analysis in recurrent high-grade glioma." *AJNR. American journal of neuroradiology* vol. 33,1 (2012): 69-76. doi:10.3174/ajnr.A2743
54. Grimm, Sean A, and Marc C Chamberlain. "Anaplastic astrocytoma." *CNS oncology* vol. 5,3 (2016): 145-57. doi:10.2217/cns-2016-0002
55. Thomsen, H., Steffensen, E., & Larsson, E. M. (2012). Perfusion MRI (dynamic susceptibility contrast imaging) with different measurement approaches for the evaluation of blood flow and blood volume in human gliomas. *Acta radiologica (Stockholm, Sweden : 1987)*, 53(1), 95–101. <https://doi.org/10.1258/ar.2011.110242>
56. Lee D. H. (1991). Mechanisms of contrast enhancement in magnetic resonance imaging. *Canadian Association of Radiologists journal = Journal l'Association canadienne des radiologistes*, 42(1), 6–12.
57. Chakhoyan, Ararat et al. "Validation of vessel size imaging (VSI) in high-grade human gliomas using magnetic resonance imaging, image-guided biopsies, and quantitative immunohistochemistry." *Scientific reports* vol. 9,1 2846. 26 Feb. 2019, doi:10.1038/s41598-018-37564-w
58. Leu, K., Boxerman, J. L., Lai, A., Nghiemphu, P. L., Pope, W. B., Cloughesy, T. F., & Ellingson, B. M. (2016). Bidirectional Contrast agent leakage correction of dynamic susceptibility contrast (DSC)-MRI improves cerebral blood volume estimation and survival prediction in recurrent glioblastoma treated with bevacizumab. *Journal of magnetic resonance imaging : JMRI*, 44(5), 1229–1237. <https://doi.org/10.1002/jmri.25227>

59. Boxerman, J L et al. "The Role of preload and leakage correction in gadolinium-based cerebral blood volume estimation determined by comparison with MION as a criterion standard." *AJNR. American journal of neuroradiology* vol. 33,6 (2012): 1081-7. doi:10.3174/ajnr.A2934
60. Law, Meng et al. "Comparison of cerebral blood volume and vascular permeability from dynamic susceptibility contrast-enhanced perfusion MR imaging with glioma grade." *AJNR. American journal of neuroradiology* vol. 25,5 (2004): 746-55.
61. Huber, P. E., Hawighorst, H., Fuss, M., van Kaick, G., Wannenmacher, M. F., & Debus, J. (2001). Transient enlargement of contrast uptake on MRI after linear accelerator (linac) stereotactic radiosurgery for brain metastases. *International journal of radiation oncology, biology, physics*, 49(5), 1339–1349.  
[https://doi.org/10.1016/s0360-3016\(00\)01511-x](https://doi.org/10.1016/s0360-3016(00)01511-x)
62. Ma, Yiming et al. "How to differentiate pseudoprogression from true progression in cancer patients treated with immunotherapy." *American journal of cancer research* vol. 9,8 1546-1553. 1 Aug. 2019
63. Nichols R. G. (1997). Drug proficiency test false positives: a lack of critical thought. *Science & justice : journal of the Forensic Science Society*, 37(3), 191–196. [https://doi.org/10.1016/S1355-0306\(97\)72173-7](https://doi.org/10.1016/S1355-0306(97)72173-7)
64. Melguizo-Gavilanes, Isaac et al. "Characterization of pseudoprogression in patients with glioblastoma: is histology the gold standard?." *Journal of neuro-oncology* vol. 123,1 (2015): 141-50.  
doi:10.1007/s11060-015-1774-5
65. Thust, Stefanie C et al. "Pseudoprogression of brain tumors." *Journal of magnetic resonance imaging : JMRI*, vol. 48,3 571–589. 7 May. 2018, doi:10.1002/jmri.26171
66. Young, Robert J et al. "MRI perfusion in determining pseudoprogression in patients with glioblastoma." *Clinical imaging* vol. 37,1 (2013): 41-9. doi:10.1016/j.clinimag.2012.02.016
67. Spampinato, M V et al. "Correlation between cerebral blood volume measurements by perfusion-weighted magnetic resonance imaging and two-year progression-free survival in gliomas." *The neuroradiology journal* vol. 26,4 (2013): 385-95. doi:10.1177/197140091302600404
68. Tsien, Christina et al. "Parametric response map as an imaging biomarker to distinguish progression from pseudoprogression in high-grade glioma." *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* vol. 28,13 (2010): 2293-9. doi:10.1200/JCO.2009.25.3971
69. Surapaneni, Krishna et al. "Early Cerebral Blood Volume Changes Predict Progression After Convection-Enhanced Delivery of Topotecan for Recurrent Malignant Glioma." *World neurosurgery* vol. 84,1 (2015): 163-72. doi:10.1016/j.wneu.2015.03.008
70. Yoo, Roh-Eul, and Seung Hong Choi. "Recent Application of Advanced MR Imaging to Predict Pseudoprogression in High-grade Glioma Patients." *Magnetic resonance in medical sciences : MRMS : an official journal of Japan Society of Magnetic Resonance in Medicine* vol. 15,2 (2016): 165-77.  
doi:10.2463/mrms.rev.2015-0053
71. Hakyemez, B., Erdogan, C., Ercan, I., Ergin, N., Uysal, S., & Atahan, S. (2005). High-grade and low-grade gliomas: differentiation by using perfusion MR imaging. *Clinical radiology*, 60(4), 493–502.  
<https://doi.org/10.1016/j.crad.2004.09.009>
72. Lee, J., Wang, N., Turk, S. et al. Discriminating pseudoprogression and true progression in diffuse infiltrating glioma using multi-parametric MRI data through deep learning. *Sci Rep* 10, 20331 (2020).  
<https://doi.org/10.1038/s41598-020-77389-0>

73. Zhou, Zhuxian, and Zheng-Rong Lu. "Gadolinium-based contrast agents for magnetic resonance cancer imaging." Wiley interdisciplinary reviews. Nanomedicine and nanobiotechnology vol. 5,1 (2013): 1-18. doi:10.1002/wnan.1198
74. Jost, Gregor et al. "Penetration and distribution of gadolinium-based contrast agents into the cerebrospinal fluid in healthy rats: a potential pathway of entry into the brain tissue." European radiology vol. 27,7 (2017): 2877-2885. doi:10.1007/s00330-016-4654-2
75. Cheng Q, Huang J, Liang J, Ma M, Ye K, Shi C, et al. The diagnostic performance of DCE-MRI in evaluating the pathological response to neoadjuvant chemotherapy in breast cancer: a meta-analysis. Front Oncol. (2020) 10:93. doi: 10.3389/fonc.2020.00093
76. Zach, Leor et al. "Delayed contrast extravasation MRI: a new paradigm in neuro-oncology." Neuro-oncology vol. 17,3 (2015): 457-65. doi:10.1093/neuonc/nou230
77. Ritchie, David J et al. "Intraparenchymal extravasation of gadolinium mimicking an enhancing brain tumor." The neuroradiology journal vol. 32,4 (2019): 273-276. doi:10.1177/1971400919853789
78. Liang J, Liu D, Gao P, Zhang D, Chen H, Shi C, et al. Diagnostic values of DCE-MRI and  
a. DSC-MRI for differentiation between high-grade and low-grade gliomas: a comprehensive meta-analysis. Acad Radiol. (2018) 25:338–48. doi: 10.1016/j.acra.2017.10.001
79. Wake, Nicole et al. "Accuracy and precision of quantitative DCE-MRI parameters: How should one estimate contrast concentration?." Magnetic resonance imaging vol. 52 (2018): 16-23. doi:10.1016/j.mri.2018.05.007
80. Duhamel, G., Schlaug, G., & Alsop, D. C. (2006). Measurement of arterial input functions for dynamic susceptibility contrast magnetic resonance imaging using echoplanar images: comparison of physical simulations with in vivo results. Magnetic resonance in medicine, 55(3), 514–523. <https://doi.org/10.1002/mrm.20802>
81. Gao P, Shi C, Zhao L, Zhou Q, Luo L. Differential diagnosis of prostate cancer and noncancerous tissue in the peripheral zone and central gland using the quantitative parameters of DCE-MRI: a meta-analysis. Medicine. (2016) 95:e5715. doi: 10.1097/MD.00000000000005715
82. Yankeelov, T. E., Luci, J. J., Lepage, M., Li, R., Debusk, L., Lin, P. C., Price, R. R., & Gore, J. C. (2005). Quantitative pharmacokinetic analysis of DCE-MRI data without an arterial input function: a reference region model. Magnetic resonance imaging, 23(4), 519–529. <https://doi.org/10.1016/j.mri.2005.02.013>
83. Zhang, Junfeng et al. "Clinical Applications of Contrast-Enhanced Perfusion MRI Techniques in Gliomas: Recent Advances and Current Challenges." Contrast media & molecular imaging vol. 2017 7064120. 20 Mar. 2017, doi:10.1155/2017/7064120
84. Gordon, Yaron et al. "Dynamic contrast-enhanced magnetic resonance imaging: fundamentals and application to the evaluation of the peripheral perfusion." Cardiovascular diagnosis and therapy vol. 4,2 (2014): 147-64. doi:10.3978/j.issn.2223-3652.2014.03.01
85. Shen, F. U., Lu, J., Chen, L., Wang, Z., & Chen, Y. (2016). Diagnostic value of dynamic contrast-enhanced magnetic resonance imaging in rectal cancer and its correlation with tumor differentiation. Molecular and clinical oncology, 4(4), 500–506. <https://doi.org/10.3892/mco.2016.762>
86. Krüger-Genge, Anne et al. "Vascular Endothelial Cell Biology: An Update." International journal of molecular sciences vol. 20,18 4411. 7 Sep. 2019, doi:10.3390/ijms20184411

87. Aquino, Domenico et al. "MRI in Glioma Immunotherapy: Evidence, Pitfalls, and Perspectives." *Journal of immunology research* vol. 2017 (2017): 5813951. doi:10.1155/2017/5813951
88. Verma, Nishant et al. "Differentiating tumor recurrence from treatment necrosis: a review of neuro-oncologic imaging strategies." *Neuro-oncology* vol. 15,5 (2013): 515-34. doi:10.1093/neuonc/nos307
89. van Dijken, Bart R J et al. "Perfusion MRI in treatment evaluation of glioblastomas: Clinical relevance of current and future techniques." *Journal of magnetic resonance imaging : JMRI* vol. 49,1 (2019): 11-22. doi:10.1002/jmri.26306
90. Kim, H. S., Goh, M. J., Kim, N., Choi, C. G., Kim, S. J., & Kim, J. H. (2014). Which combination of MR imaging modalities is best for predicting recurrent glioblastoma? Study of diagnostic accuracy and reproducibility. *Radiology*, 273(3), 831–843. <https://doi.org/10.1148/radiol.14132868>
91. Siemann, Dietmar W. "The unique characteristics of tumor vasculature and preclinical evidence for its selective disruption by Tumor-Vascular Disrupting Agents." *Cancer treatment reviews* vol. 37,1 (2011): 63-74. doi:10.1016/j.ctrv.2010.05.001
92. Dewhirst, Mark W, and Timothy W Secomb. "Transport of drugs from blood vessels to tumour tissue." *Nature reviews. Cancer* vol. 17,12 (2017): 738-750. doi:10.1038/nrc.2017.93
93. Mankoff, David A et al. "Blood flow-metabolism mismatch: good for the tumor, bad for the patient." *Clinical cancer research : an official journal of the American Association for Cancer Research* vol. 15,17 (2009): 5294-6. doi:10.1158/1078-0432.CCR-09-1448
94. Jain, R., Ellika, S. K., Scarpace, L., Schultz, L. R., Rock, J. P., Gutierrez, J., Patel, S. C., Ewing, J., & Mikkelsen, T. (2008). Quantitative estimation of permeability surface-area product in astroglial brain tumors using perfusion CT and correlation with histopathologic grade. *AJNR. American journal of neuroradiology*, 29(4), 694–700. <https://doi.org/10.3174/ajnr.A0899>
95. Baron J. C. (2001). Perfusion thresholds in human cerebral ischemia: historical perspective and therapeutic implications. *Cerebrovascular diseases* (Basel, Switzerland), 11 Suppl 1, 2–8. <https://doi.org/10.1159/000049119>
96. Borogovac, Ajna, and Iris Asllani. "Arterial Spin Labeling (ASL) fMRI: advantages, theoretical constraints, and experimental challenges in neurosciences." *International journal of biomedical imaging* vol. 2012 (2012): 818456. doi:10.1155/2012/818456
97. Ziyad, Safiyyah, and M Luisa Iruela-Arispe. "Molecular mechanisms of tumor angiogenesis." *Genes & cancer* vol. 2,12 (2011): 1085-96. doi:10.1177/1947601911432334
98. Yao, V. J., Ozawa, M. G., Varner, A. S., Kasman, I. M., Chanthery, Y. H., Pasqualini, R., Arap, W., & McDonald, D. M. (2006). Antiangiogenic therapy decreases integrin expression in normalized tumor blood vessels. *Cancer research*, 66(5), 2639–2649. <https://doi.org/10.1158/0008-5472.CAN-05-1824>
99. Salehi, Arjang et al. "Response of the cerebral vasculature following traumatic brain injury." *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism* vol. 37,7 (2017): 2320-2339. doi:10.1177/0271678X17701460
100. Barker, Holly E et al. "The tumour microenvironment after radiotherapy: mechanisms of resistance and recurrence." *Nature reviews. Cancer* vol. 15,7 (2015): 409-25. doi:10.1038/nrc3958
101. Rotchanapanya, Wannaphorn et al. "Clinical Outcomes Based on Measurable Residual Disease Status in Patients with Core-Binding Factor Acute Myeloid Leukemia: A Systematic Review and Meta-Analysis."

102. Beauchemin, C et al. "Relationship between progression-free survival and overall survival in chronic lymphocytic leukemia: a literature-based analysis." *Current oncology* (Toronto, Ont.) vol. 22,3 (2015): e148-56. doi:10.3747/co.22.2119
103. Young, A., Poretti, A., Bosemani, T., Goel, R., & Huisman, T. (2017). Sensitivity of susceptibility-weighted imaging in detecting developmental venous anomalies and associated cavernomas and microhemorrhages in children. *Neuroradiology*, 59(8), 797–802. <https://doi.org/10.1007/s00234-017-1867-2>
104. Thomas, B., Somasundaram, S., Thamburaj, K., Kesavadas, C., Gupta, A. K., Bodhey, N. K., & Kapilamoorthy, T. R. (2008). Clinical applications of susceptibility weighted MR imaging of the brain - a pictorial review. *Neuroradiology*, 50(2), 105–116. <https://doi.org/10.1007/s00234-007-0316-z>
105. Löbel, Ulrike et al. "Three-dimensional susceptibility-weighted imaging and two-dimensional T2\*-weighted gradient-echo imaging of intratumoral hemorrhages in pediatric diffuse intrinsic pontine glioma." *Neuroradiology* vol. 52,12 (2010): 1167-77. doi:10.1007/s00234-010-0771-9
106. Stadnik, T W et al. "Diffusion-weighted MR imaging of intracerebral masses: comparison with conventional MR imaging and histologic findings." *AJNR. American journal of neuroradiology* vol. 22,5 (2001): 969-76.
107. Sehgal, V., Delproposto, Z., Haddar, D., Haacke, E. M., Sloan, A. E., Zamorano, L. J., Barger, G., Hu, J., Xu, Y., Prabhakaran, K. P., Elangovan, I. R., Neelavalli, J., & Reichenbach, J. R. (2006). Susceptibility-weighted imaging to visualize blood products and improve tumor contrast in the study of brain masses. *Journal of magnetic resonance imaging : JMRI*, 24(1), 41–51. <https://doi.org/10.1002/jmri.20598>
108. Ozturk, K., & Nascene, D. (2021). Susceptibility-Weighted Imaging of the Pediatric Brain after Repeat Doses of Gadolinium-Based Contrast Agent. *AJNR. American journal of neuroradiology*, 42(6), 1136–1143. <https://doi.org/10.3174/ajnr.A7143>
109. Xia, X. B., & Tan, C. L. (2013). A quantitative study of magnetic susceptibility-weighted imaging of deep cerebral veins. *Journal of neuroradiology = Journal de neuroradiologie*, 40(5), 355–359. <https://doi.org/10.1016/j.neurad.2013.03.005>
110. Barnes, Samuel R S, and E Mark Haacke. "Susceptibility-weighted imaging: clinical angiographic applications." *Magnetic resonance imaging clinics of North America* vol. 17,1 (2009): 47-61. doi:10.1016/j.mric.2008.12.002
111. Kim, Baek Gil et al. "Laminin-332-rich tumor microenvironment for tumor invasion in the interface zone of breast cancer." *The American journal of pathology* vol. 178,1 (2011): 373-81. doi:10.1016/j.ajpath.2010.11.028
112. Martin TA, Ye L, Sanders AJ, et al. Cancer Invasion and Metastasis: Molecular and Cellular Perspective. In: *Madame Curie Bioscience Database* [Internet]. Austin (TX): Landes Bioscience; 2000-2013. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK164700/>
113. Aydın, Ö., Büyükkaya, R., & Hakyemez, B. (2017). Contrast medium enhanced susceptibility imaging signal mechanism; should we use contrast medium?. *Acta radiologica* (Stockholm, Sweden : 1987), 58(1), 107–113. <https://doi.org/10.1177/0284185116637246>
114. Haacke, E. M., Mittal, S., Wu, Z., Neelavalli, J., & Cheng, Y. C. (2009). Susceptibility-weighted imaging: technical aspects and clinical applications, part 1. *AJNR. American journal of neuroradiology*, 30(1),

19–30. <https://doi.org/10.3174/ajnr.A1400>

115. Kong, LW., Chen, J., Zhao, H. et al. Intratumoral Susceptibility Signals Reflect Biomarker Status in Gliomas. *Sci Rep* 9, 17080 (2019). <https://doi.org/10.1038/s41598-019-53629-w>
116. McGahan, Ben G et al. “Assessment of vascularity in glioblastoma and its implications on patient outcomes.” *Journal of neuro-oncology* vol. 132,1 (2017): 35-44. doi:10.1007/s11060-016-2350-3
117. Saini, Jitender et al. “Comparative evaluation of cerebral gliomas using rCBV measurements during sequential acquisition of T1-perfusion and T2\*-perfusion MRI.” *PloS one* vol. 14,4 e0215400. 24 Apr. 2019, doi:10.1371/journal.pone.0215400
118. Mohammed, Wasif et al. “Clinical applications of susceptibility-weighted imaging in detecting and grading intracranial gliomas: a review.” *Cancer imaging : the official publication of the International Cancer Imaging Society* vol. 13,2 186-95. 24 Apr. 2013, doi:10.1102/1470-7330.2013.0020
119. Eisele, P., Fischer, K., Szabo, K., Platten, M., & Gass, A. (2019). Characterization of Contrast-Enhancing and Non-contrast-enhancing Multiple Sclerosis Lesions Using Susceptibility-Weighted Imaging. *Frontiers in neurology*, 10, 1082. <https://doi.org/10.3389/fneur.2019.01082>