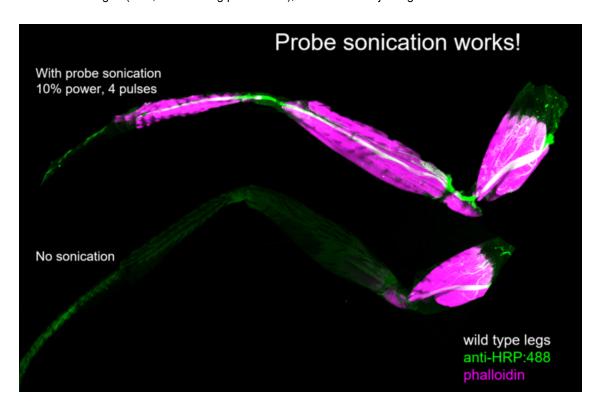
Fly Leg Phalloidin Staining Protocol with Sonication

June 2024 - Anne Sustar - Tuthill Lab

Note:

Sonication makes small tears in the cuticle and allows reagents to penetrate. Typically I work in batches of 20 legs per experiment, and I see a mix of results: about a third of the legs are over-damaged (thus bad morphology), a third are under-damaged (thus, bad staining penetration), and a third are just right.



Dissection and fixing

Dissect legs from an anesthetized fly. Put legs in a 1.5 mL tube with fixative. Nutate in fixative for 20 mins.

Fixative

 $390~\mu L$ PBS

10 μL PBS-Tx (PBS with 0.2% Triton-X)

48µL 38% aqueous formaldehyde (Macron 5016-02)

Wash

Wash with PBS-Tx (0.2% triton-X) 3x over 30-60 mins at RT, then 2x with PBS. End with ~ 500 mL volume of PBS.

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Probe sonication

For a Branson Sonifier 450:

- Set to 10% output control (higher power will shred the legs)
- Dip the probe into the tube. Assuming legs are all at bottom of the tube, the probe tip should be lowered to \sim 3mm above the legs.
- Do 4 short (~2 second each) pulses. Place tube on ice between the pulses
- Clean probe with ethanol.



Phalloidin Stain with extra penetrants

Make a big batch, freeze in 0.5mL aliquots (based on a recipe form Igor Siwanowicz at Janelia)

phalloidin stain			final concentraion
75	μL	triton X-100 (VWR 0694-1L)	1%
37.5	μL	DMSO (Sigma 154938)	0.5%
0.375	mg	Escin (Sigma E1378)	0.05 mg/mL
225	μL	normal goat serum (Sigma G9023)	3%
150	μL	Alexa Fluor™ 647 Phalloidin (Thermofisher A22287)	2%
1.5	μL	sodium azide (Sigma, S2002)	0.02%
7	mL	<u>PBS</u>	
7.5	mL	TOTAL	

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Incubate legs for ~ 3 days at room temperature in a small tube. Briefly vortex twice per day.

Clear and mount (FocusClear/ Focus Mount)

Wash 3x with PBS-Tx over one day, RT, Wash 1x in PBS.

Remove PBS and replace with ~100 μ L FocusClear (CelExplorer FC-101). Incubate in FocusClear at RT for about 20-30 minutes. Flick tube periodically to make sure legs are submerged (Legs tend to float in FocusClear)

Transfer leg pieces to a slide with a small drop of MountClear (CelExplorer MC-301). Cover with size #0 coverslips with 3M Scotch doublestick tape (1 layer) as spacers.