

# New insights into the boron essentiality on humans and animals

## DATA AVAILABILITY

### PART I

#### In vivo Calcium Fructoborate (CaFB) feeding

*In vivo* CaFB feeding was carried out on two groups of six months old male Wistar rats, each of two animals, with an average weight of 290±10 g:

- Group 1 (B-supplemented) → two rats fed with CaFB as 150 ppm B;
- Group 2 (Reference) → two rats fed the normal diet.

During the first day of the experiment, the rats were chosen and assigned a specific group, while the second day each group started to receive the high boron diet (Group 1) and the normal diet (Group 2), respectively.

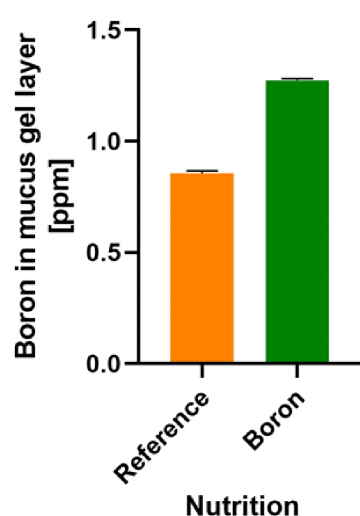
Throughout the experiment, animals were kept under observation in individual cages, in the Animal Facility of the University of Medicine and Pharmacy of Craiova, Romania, under standard conditions of temperature, humidity, lighting (12-hour light/dark cycle – 7/7), food and water (*ad libitum*).

The next seven days, fecal matter was collected from each rat and food was resupplied at four days.

At the end of the seven days, the rats were euthanized, under general anesthesia, and the following parts were stored for later analysis: (i) cecum mucus, (ii) cecum with mucus, (iii) cecum content, (iv) colon mucus, (v) colon with mucus, (vi) colon content, (vii) rectum with mucus, (viii) rectum content.

The general anesthesia was performed by intramuscular injection of an anesthetic cocktail consisting of Ketamine hydrochloride 90 mg/kg body weight (b.w.) (Ketamidol® 100 mg/mL, Richter Pharma AG, Austria) and Xylazine hydrochloride 10 mg/kg b.w. (Xylazin Bio® 2%, Bioveta, Czech Republic).

The experimental protocol was applied according with the European Council Directive No. 86/609 (November 24, 1986), the European Convention for the Protection of Vertebrate Animals used for Experimental and Other Scientific Purposes (December 2, 2005), and the Romanian Parliament Law No. 43 (April 11, 2014) on the protection of animals used for scientific purposes. The study was approved by the Ethics Committee of the University of Medicine and Pharmacy of Craiova.



**Boron detection in the rat's colonic mucus gel layer. Reference: normal diet [Dupre et al., 1994]. Boron: CaFB (NOB) supplemented diet. Boron detection has been performed by UHPLC/MS method [Sah & Brown, 1997].**

**CaFB: Calcium fructoborate; NOB: Naturally organic boron; UHPLC/MS: Ultra-high performance liquid chromatography/mass spectrometry.**

## PART II

### In vitro digestion simulation

*In vitro* digestion simulation was performed using a gastric simulator adapted after Ferrua & Singh (2015), Wang et al. (2021) and Kong & Singh (2010).

Simulated gastric fluid (SGF) was prepared by dissolving 1.38 g NaCl, 0.0612 g KH<sub>2</sub>PO<sub>4</sub>, 0.257 g KCl, 0.0122g MgCl<sub>2</sub>, 1.05 g NaHCO<sub>3</sub> and 0.024 g (NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> per 0.5 L of SGF, adjusting the pH with 0.6 N HCl. CaCl<sub>2</sub>•H<sub>2</sub>O (0.0193 g/0.5 L of SGF) was not added to the SGF solution initially, as it may precipitate, but was added to the final mixture after adjusting the pH. At the final mixture, we added 2.5 g porcine pepsin to simulate enzymatic activity [Minekus et al., 2014].

The experiment was conducted at three distinct pH values, knowing that SGF has different pH values, usually with values between 3–5, depending on the type of food consumed, or below 1.5 on an empty stomach: pH 1.3, to simulate fasted gastric pH, which is commonly below 1.5 [Carrière et al., 1991]; pH 3, when the postprandial pH was simulated in the case of a typical meat and cola meal; pH 4.5, when the postprandial pH was simulated in the case of a typical meal with vegetables.

Gastric motility was stimulated using a shaker at 100 rpm (stimulating actual stomach contraction). To stimulate the preprandial environment of the stomach (before a meal), 70 mL of simulated gastric juice was first placed in the gastric simulator and brought to 37°C. 500 mg of Fructoborate (FB) was added over it, and an initial FB sample of 2 mL was taken (7.14 mg/mL FB). The addition of FGS was started for three hours at a flow rate of 2.5 mL/min. At regular intervals (30, 60, 90, 120, 150, 180 minutes), equal amounts (2 mL) of solution were extracted and subjected to physico-chemical analysis.

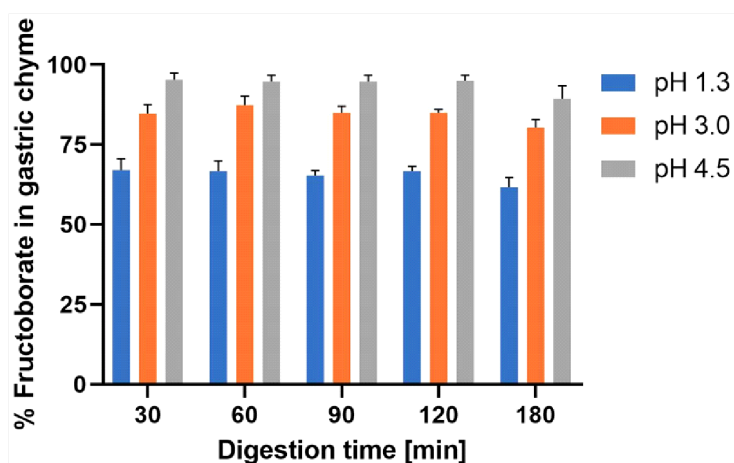
### Parameters for *in vitro* dynamic gastric digestion

Gastric conditions (37°C)	
Simulated gastric fluid (SGF)	Constituent concentration in SGF [mmol/L]
NaCl	47.2
KH <sub>2</sub> PO <sub>4</sub>	0.9
KCl	6.9
MgCl <sub>2</sub> • 6H <sub>2</sub> O	0.12
(NH <sub>4</sub> ) <sub>2</sub> CO <sub>3</sub>	0.5
CaCl <sub>2</sub> • H <sub>2</sub> O	0.14
NaHCO <sub>3</sub>	25
Enzyme	Concentration [g/L]
Porcine pepsin	2.5
<i>pH</i>	1.3, 3, 4.5
<i>Gastric motility</i>	100 rpm
<i>Digestion time</i>	180 minutes
<i>Flow rate</i>	2.5 mL/min

### Measurements of Fructoborate (FruitexB) *in vitro* gastric digestion

Digestion time [min]	% Fructoborate in gastric chyme														
	pH 1.3				±SD	pH 3				±SD	pH 4.5				±SD
	<i>I<sup>st</sup> Dtn.</i>	<i>II<sup>nd</sup> Dtn.</i>	<i>III<sup>rd</sup> Dtn.</i>	<i>±SD</i>		<i>I<sup>st</sup> Dtn.</i>	<i>II<sup>nd</sup> Dtn.</i>	<i>III<sup>rd</sup> Dtn.</i>	<i>±SD</i>		<i>I<sup>st</sup> Dtn.</i>	<i>II<sup>nd</sup> Dtn.</i>	<i>III<sup>rd</sup> Dtn.</i>	<i>±SD</i>	
30	63.34	68.23	70.15	2.87	88.32	83.12	83.02	2.48	97.05	93.45	96.95	1.67			
60	69.45	63.67	68.54	2.54	89.03	89.12	84.78	2.03	97.67	93.94	94.38	1.66			
90	65.78	64.08	67.11	1.24	87.28	83.49	85.11	1.55	97.07	94.04	93.19	1.67			
120	68.21	67.07	65.56	1.09	85.29	84.67	86.38	0.71	94.74	97.12	94.76	1.12			

Dtn.: Determination; SD: Standard deviation.



*In vitro* simulation of gastric digestion of NOB species. NOB: Naturally organic boron.

## References

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