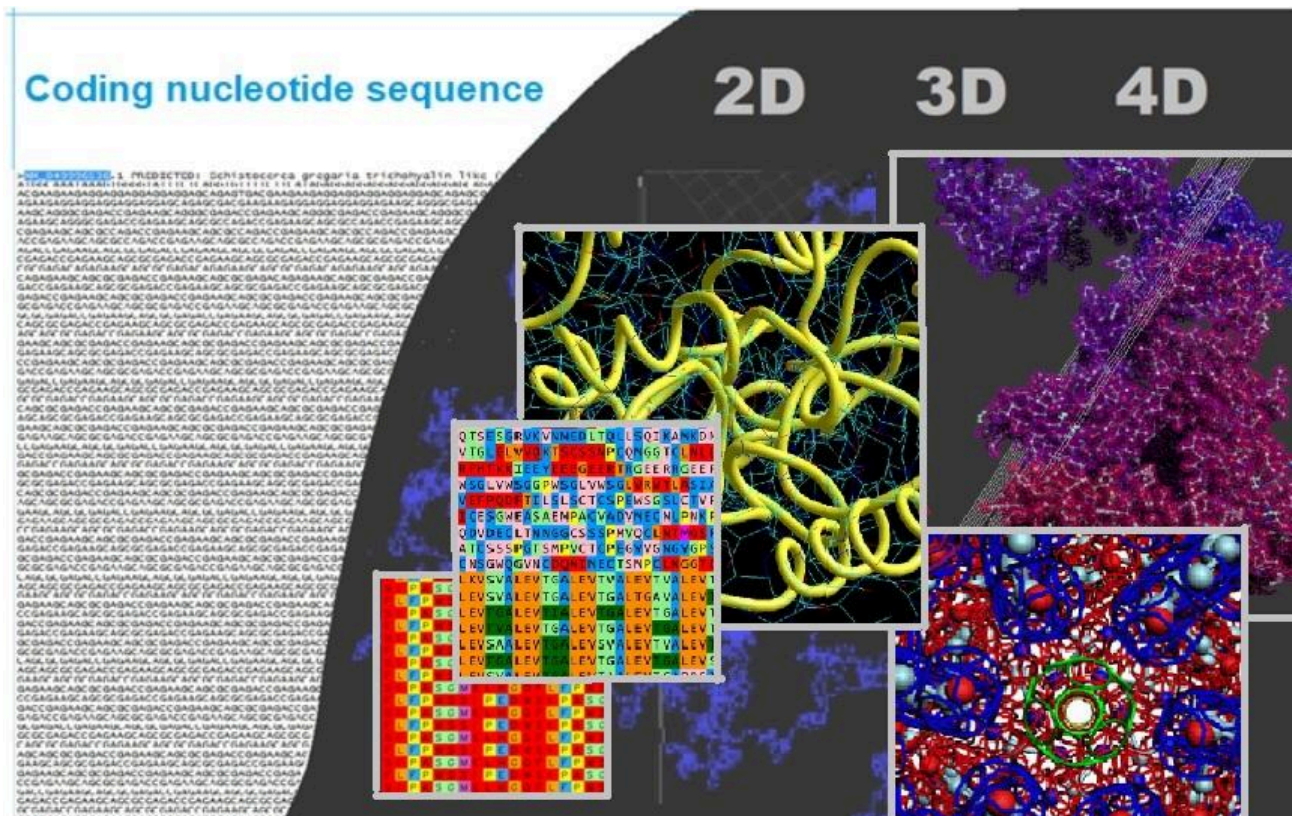
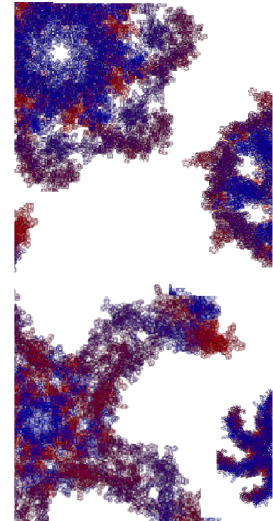
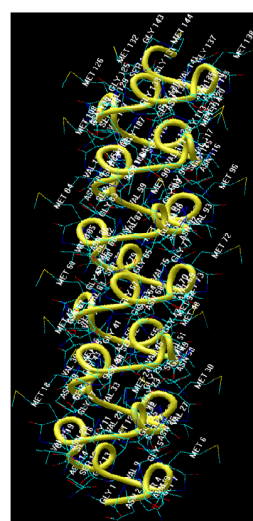


Innovative method of converting nucleotide sequences into 2D, 3D and 4D structures



Software in development





Dear Researchers !

Let us introduce to your attention our development – *Picotechnology of proteins* ([3D Genetic code](#)) - a method of constructing accurate [2D](#) structures, where the initial data is only the nucleotide sequence. Visualization of some [3D](#) and [4D](#) structures is available. This approach was made possible by modeling [electrons as tori \(rings\) of standing waves](#), and [atomic shells as polyhedrons assembled from rings](#).

In [this approach](#) all amino acid residues have the same structural template, and the third letter of the triplet controls the angle of rotation of the next amino acid residue relative to the previous one. Each of the angles is marked on the 2D diagram with its own color and corresponds to the algorithm for forming alpha, beta, pi, 310, methionine and proline helices.

On the 2D color diagram we clearly see the helical sections, individual turns and individual amino acid residues.

We can directly convert the 2D colour diagram to 3D and 4D structure for some structural problems (3D and 4D modeling takes into account [other adjustments](#)). Software for 2D structures has been developed. Software for 3D and 4D structures is under development.

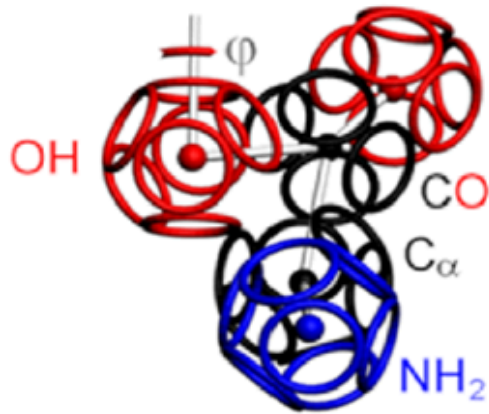
Our 2D diagrams reveal super-secondary structures ([fractal helices](#), [software helices](#)), a coincidence confirmed by published data. The closure of disulfide bridges obtained on 2D and 3D models coincided with experimental data.

We hope that our algorithms will increase the accuracy of data processing both in the field of computational biology and in the field of [X-ray structure prediction](#), [CD](#) and NMR methods. We would appreciate your comments and cooperation, see [DEMO PICOTECH 2D, 3D, 4D, AND ARTICLES](#) .

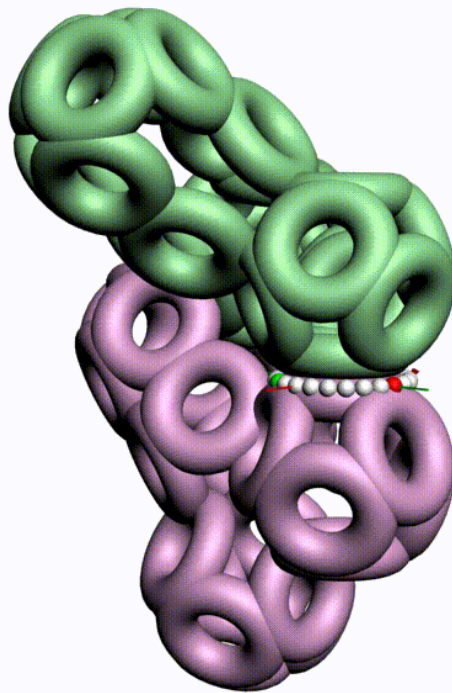
With best wishes,
Alexander Kushelev, Head of Nanoworld Laboratory
Tatyana Ryasina, Referent of Nanoworld Laboratory

The Legend

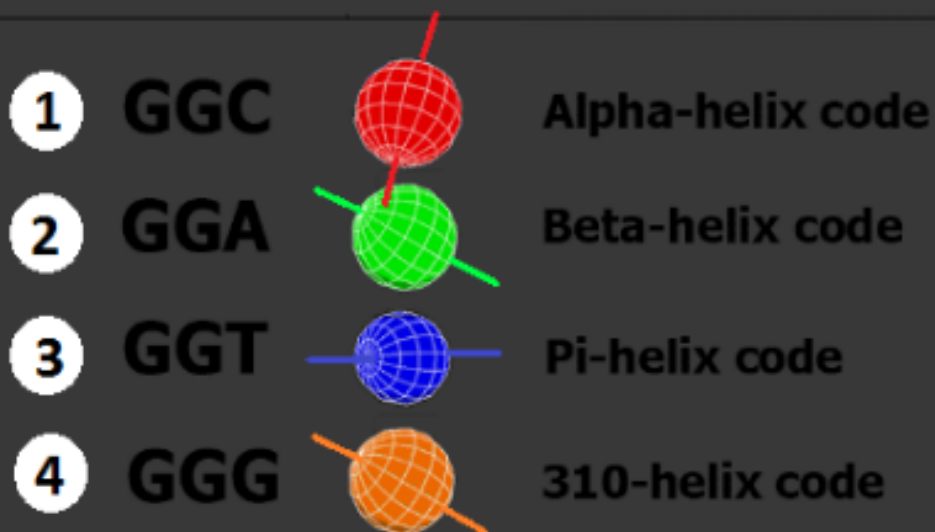
HOW TO READ 2D DIAGRAMS OF 3D GENETIC CODE




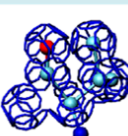
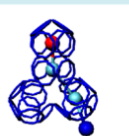
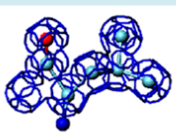
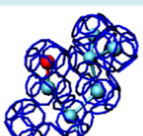
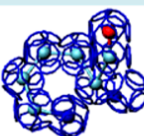
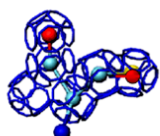
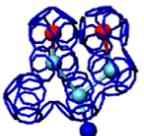
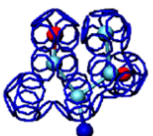

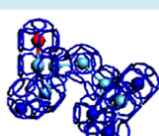
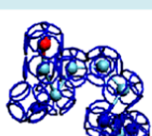
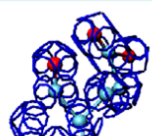
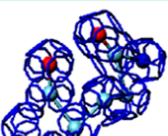
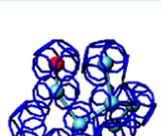
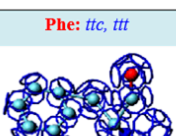
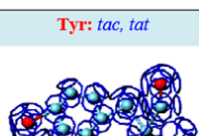

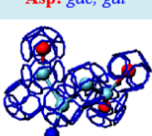
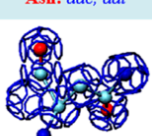
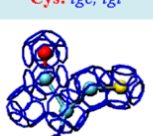
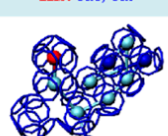
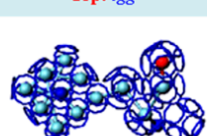

Structural
template of
amino acid
residue



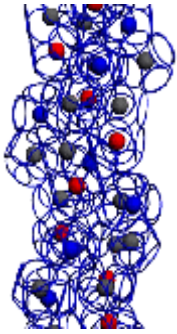
Structure definition by 3D genetic code



3D STRUCTURES OF AMINO ACID RESIDUES

Ala: gcc, gca, gct, gcg	Val: gtc, gta, gtt, gtg	Gly: ggc, gga, ggt, ggg	Leu^a: ctc/cta, ctt, ctg	Leu^b: tta, ttg	Lys: aaa, aag
					
Ser^a: tcc, tca, tct, tcg	Ser^b: agc, agt	Thr: acc, aca, act, acg	Met: atg	Arg^a: cgc, cga, cgt, cgg	Arg^b: aga, agg
					
Glu: gaa, gag	Gln: caa, cag	Ile: atc, ata, att	Phe: ttc, ttt	Tyr: tac, tat	Pro: ccc, cca, cct, ccg
					
Asp: gac, gat	Asn: aac, aat	Cys: tgc, tgt	His: cac, cat	Trp: tgg	Stop-кодон: taa, tag, tga
					

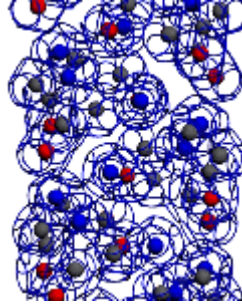
Примечание. Ядра атомов кислорода показаны **красным** цветом, углерода — **голубым**, азота — **синим**, серы — **желтым**. Электроны внешних электронных оболочек атомов, формирующие молекулярную электронную оболочку аминокислот, обозначены **синими** кольцами. Для Arg, Leu и Ser представлены по две модели изомеров по положению радикала, которые кодируются по-разному.



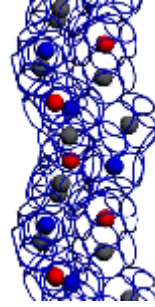
Alpha-helix



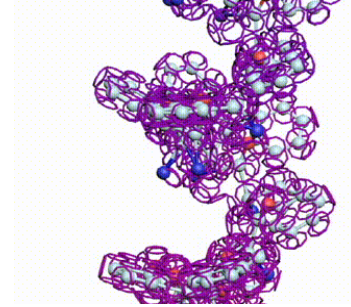
Beta-helix



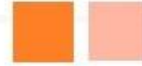
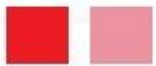
Pi-helix



310-helix



Methionine in a helix



A whole turn of the helix or more



Single amino acid residue



One or more prolines in a row

COMPACT 2D PICOTECHNOLOGY DIAGRAM

Red is an alpha helix.

The orange is a 310-helix.

Pink is a single alpha / 310 helix code.

Blue is a py-helix.

Green is a beta helix.

Lilac - methionine helix. It has a larger pitch of "thread" than the usual alpha-helix.

Black in abbreviated form and white in unfold means either an unknown code or the end of the translation.

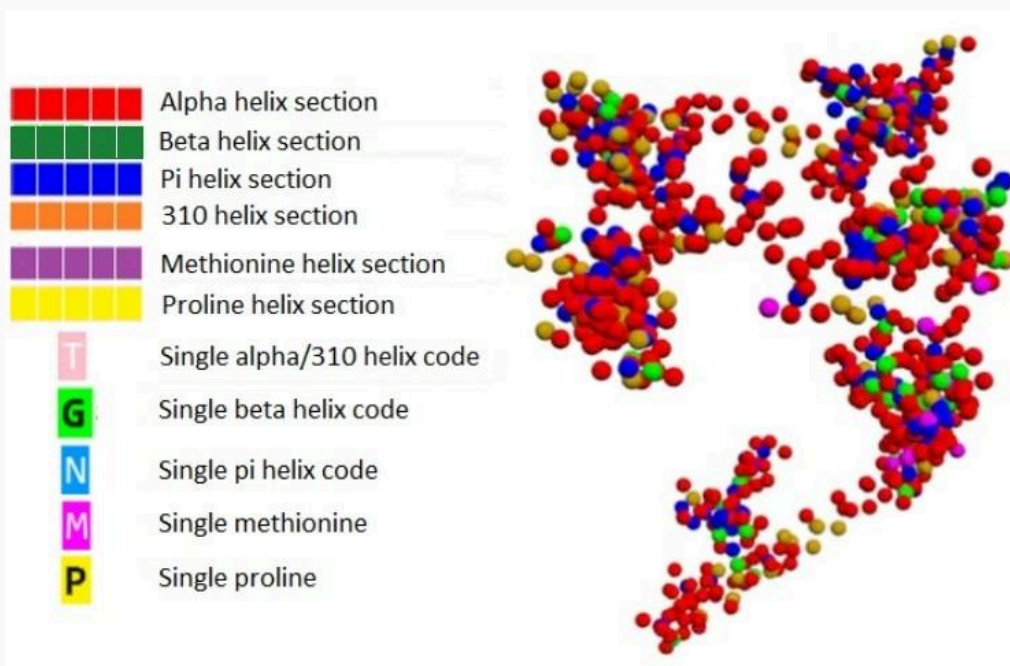
Cyclic repetition of colors - software helix.

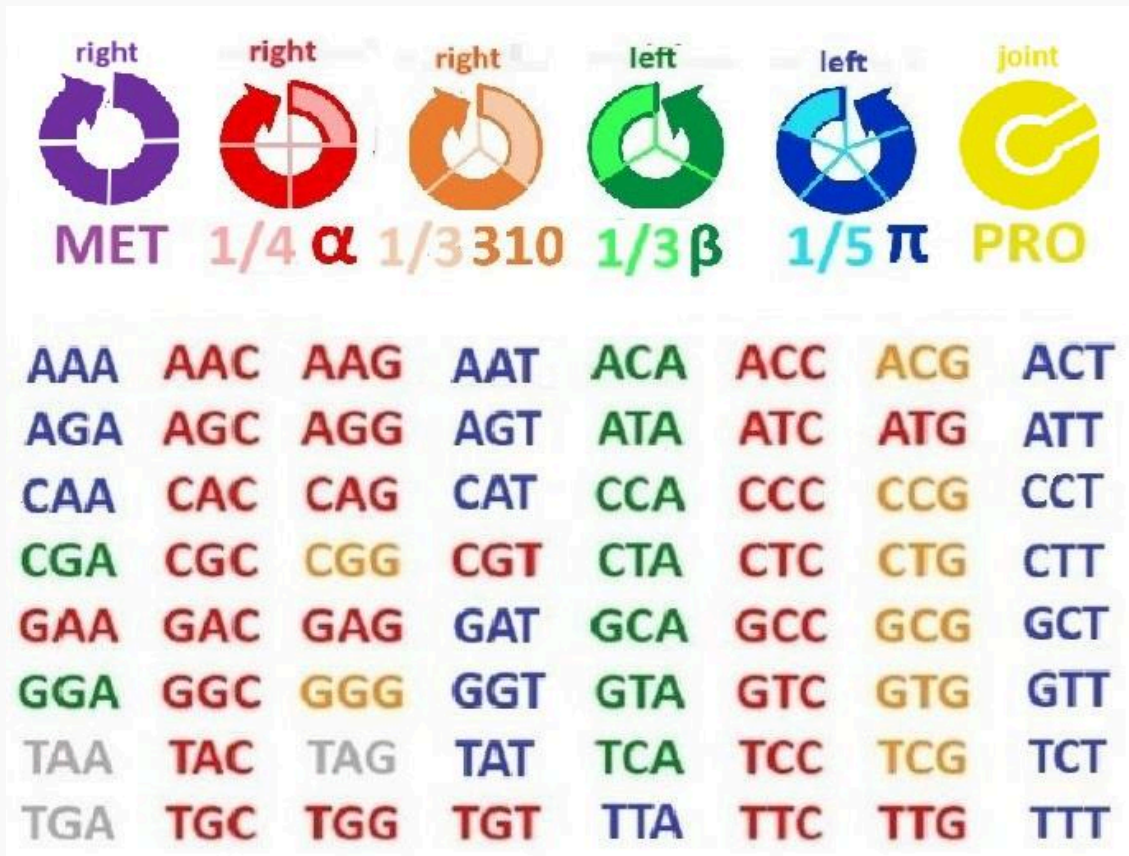
Alpha helix - right,

Pi helix - left,

Beta helix - left,

310 helix - right.





How does the 3D Genetic code table work?

		Second base position								
		U		C		A		G		
First base position	U	UUU	P π	UCU	S π	UAU	Y π	UGU	C π	U
		UUC	P α	UCC	S α	UAC	Y α	UGC	C α	C
		UUA	L π	UCA	S β	UAA	Stop	UGA	Stop	A
		UUG	L α	UCG	S 3_{10}	UAG	Stop	UGG	W α	G
	C	CUU	L π	CCU	P π	CAU	H π	CGU	R π	U
		CUC	L α	CCC	P α	CAC	H α	CGC	R α	C
		CUA	L β	CCA	P β	CAA	Q π	CGA	R β	A
		CUG	L 3_{10}	CCG	P 3_{10}	CAG	Q α	CGG	R 3_{10}	G
	A	AUU	I π	ACU	T π	AAU	N π	AGU	S π	U
		AUC	I α	ACC	T α	AAC	N α	AGC	S α	C
		AUA	I β	ACA	T β	AAA	K π	AGA	R π	A
		AUG	M α	ACG	T 3_{10}	AAG	K α	AGG	R α	G
	G	GUU	V π	GCU	A π	GAU	D π	GGU	G π	U
		GUC	V α	GCC	A α	GAC	D α	GGC	G α	C
		GUA	V β	GCA	A β	GAA	E π	GGA	G β	A
		GUG	V 3_{10}	GCG	A 3_{10}	GAG	E α	GGG	G 3_{10}	G

α - 1/4 turn of the alpha helix (alpha turn)

β - 1/3 turn of beta helix (beta turn)

π - 1/5 turn of pi-helix (pi-turn)

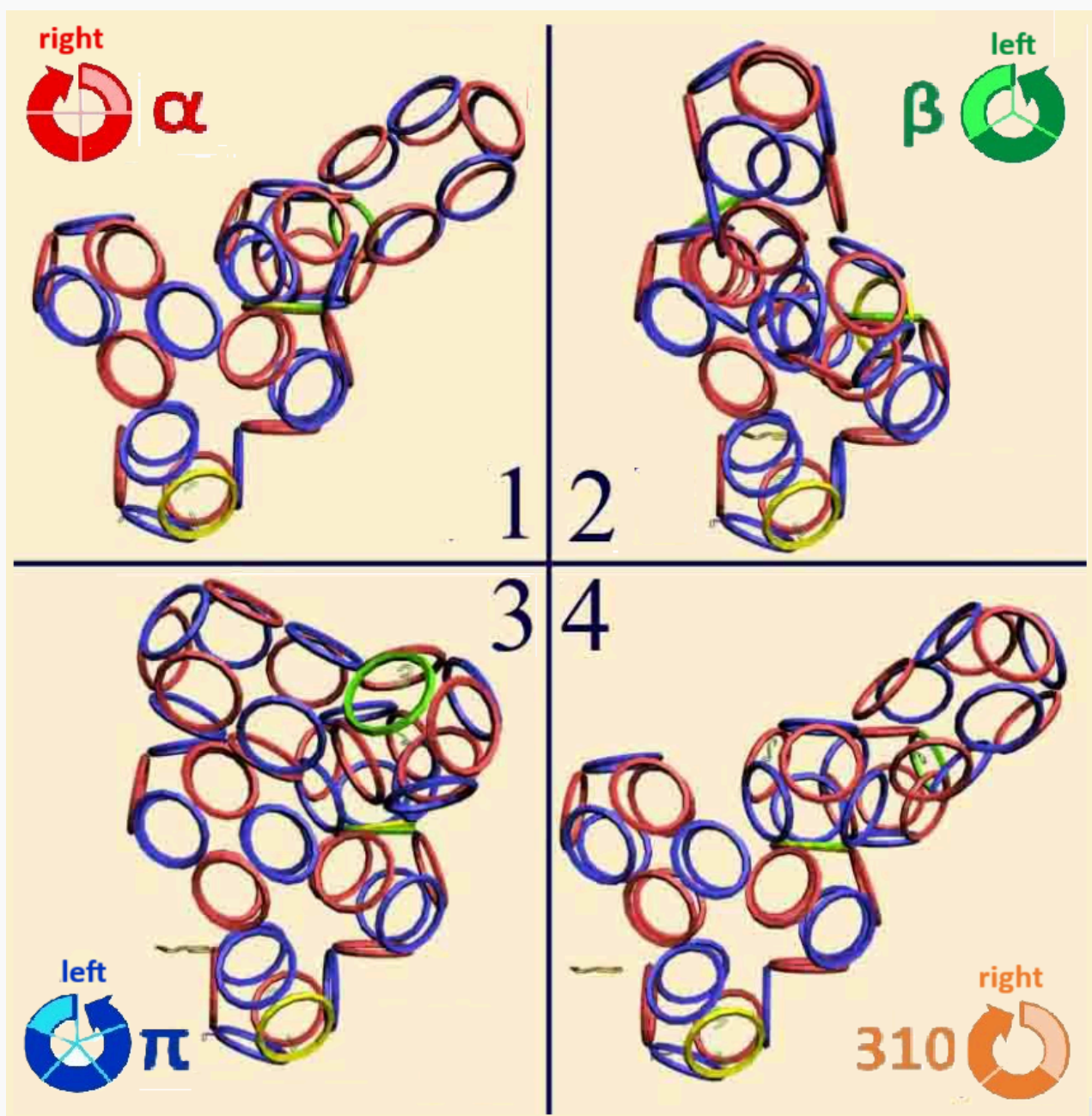
3_{10} - 1/3 turn 3_{10} -helix (3_{10} -turn)

Input data: GGC GGC GGC GGC GGT GGT GGA GGA ...

1 2 3 4 5 6 7 8

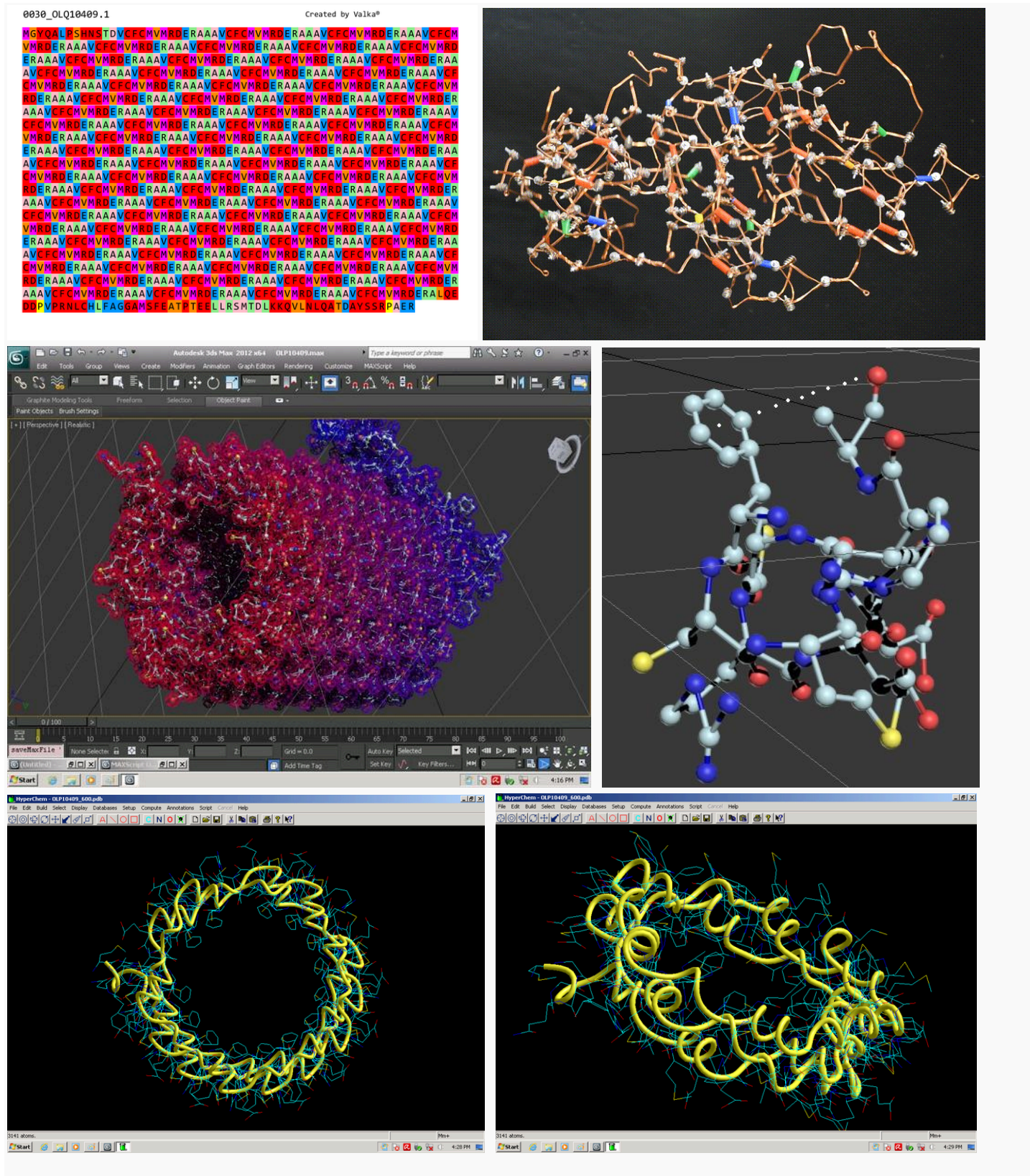
Output data: GGGGGGGG

X	Y	C	a.o.	A	a.o.	T	a.o.	G	a.o.	Z	РВПС	ω
C		CCC	P	CAC	H	CTC	L	CGC	R	C	R	35°
		CCA		CAA	Q	CTA		CGA		A	0	155°
		CCT		CAT	H	CTT		CGT		T	L	275°
		CCG		CAG	Q	CTG		CGG		G	R	35°
A		ACC	T	AAC	N	ATC	I	AGC	S	C	R	35°
		ACA		AAA	K	ATA		AGA	R	A	0	155°
		ACT		AAT	N	ATT		AGT	S	T	L	275°
		ACG		AAG	K	ATG	M	AGG	R	G	R	35°
T		TCC	S	TAC	Y	TTC	F	TGC	C	C	R	35°
		TCA		TAA	Stop	TTA	L	TGA	Stop	A	0	155°
		TCT		TAT	Y	TTT	F	TGT	C	T	L	275°
		TCG		TAG	Stop	TTG	L	TGG	W	G	R	35°
G		GCC	A	GAC	D	GTC	V	GGC	G	C	R	35°
		GCA		GAA	E	GTA		GGA		A	0	155°
		GCT		GAT	D	GTT		GGT		T	L	275°
		GCG		GAG	E	GTG		GGG		G	R	35°



[illegible]

While the 3D picotech program is in development, [we can model proteins manually](#) and then digitize them to deliver the result as a 3D images and .pdb files.



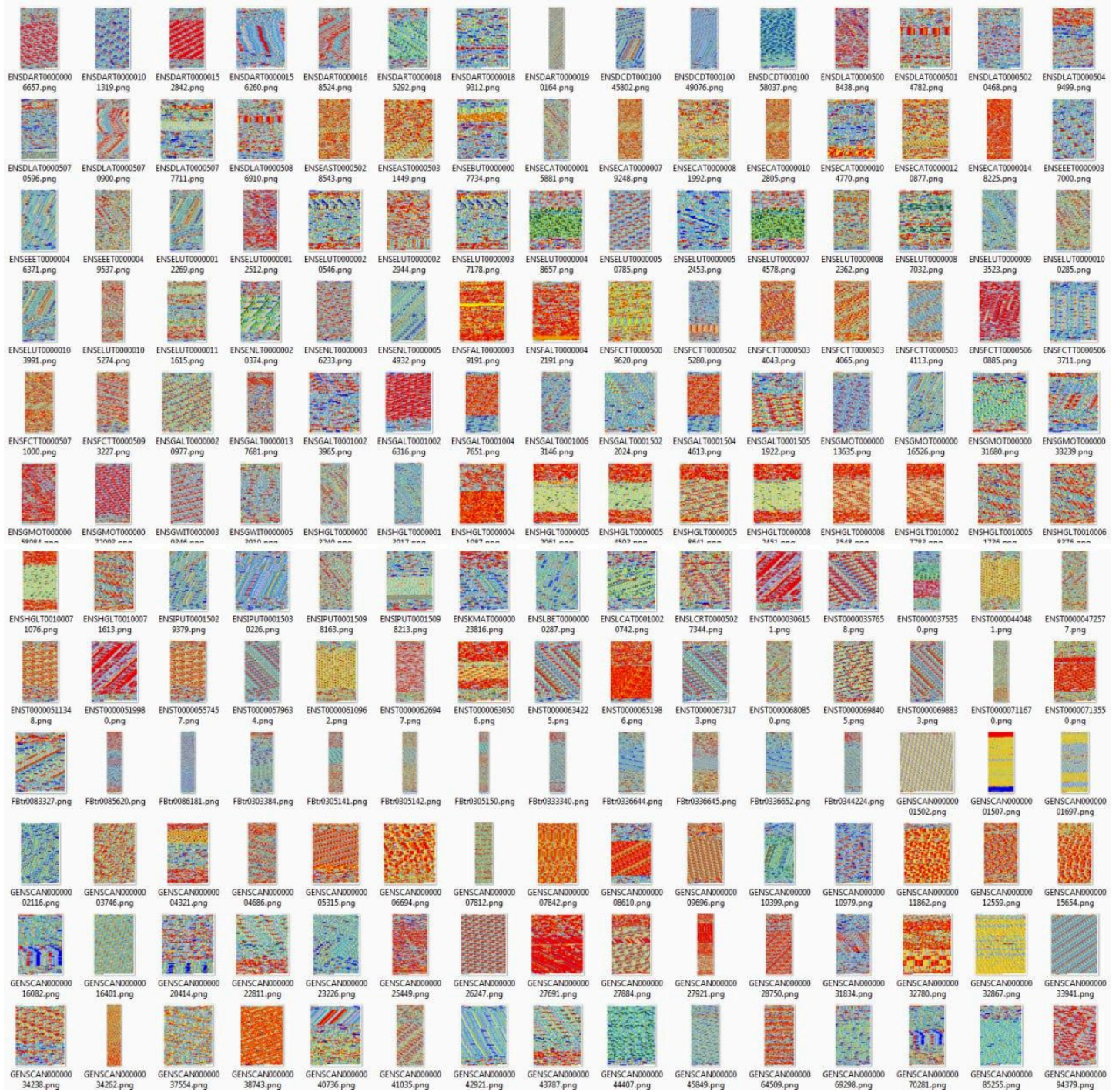
1992

Код	Остаток	Вариант
AAA	Lys	3
AAC	Asn	1
AAG	Lys	1
AAT	Asn	3
ACA	Thr	2
ACC	Thr	1
ACG	Thr	4
ACT	Thr	3
AGA	Arg	3
AGC	Ser	1
AGG	Arg	1
AGT	Ser	3
ATA	Ile	2
ATC	Ile	1
ATG	Met	1
ATT	Ile	3
CAA	Gln	3
CAC	His	1
CAG	Gln	1
CAT	His	3
CCA	Pro	2
CCC	Pro	1
CCG	Pro	4
CCT	Pro	3
CGA	Arg	2
CGC	Arg	1
CGG	Arg	4
CGT	Arg	3
CTA	Leu	2
CTC	Leu	1
CTG	Leu	4
CTT	Leu	3
GAA	Glu	3
GAC	Asp	1
GAG	Glu	1
GAT	Asp	3
GCA	Ala	2
GCC	Ala	1
GCG	Ala	4
GCT	Ala	3
GGA	Gly	2
GGC	Gly	1
GGG	Gly	4
GGT	Gly	3
GTA	Val	2
GTC	Val	1
GTG	Val	4
GTT	Val	3
TAC	Tyr	-
TAA	Tyr	1
TAG	TKD	-
TAT	Tyr	3
TCA	Ser	2
TCC	Ser	1
TCG	Ser	4
TCT	Ser	3
TGA	TKD	-
TGC	Cys	1
TGG	Trp	1
TGT	Cys	3
TTA	Leu	3
TTC	Phe	1
TTG	Leu	1
TTT	Phe	3

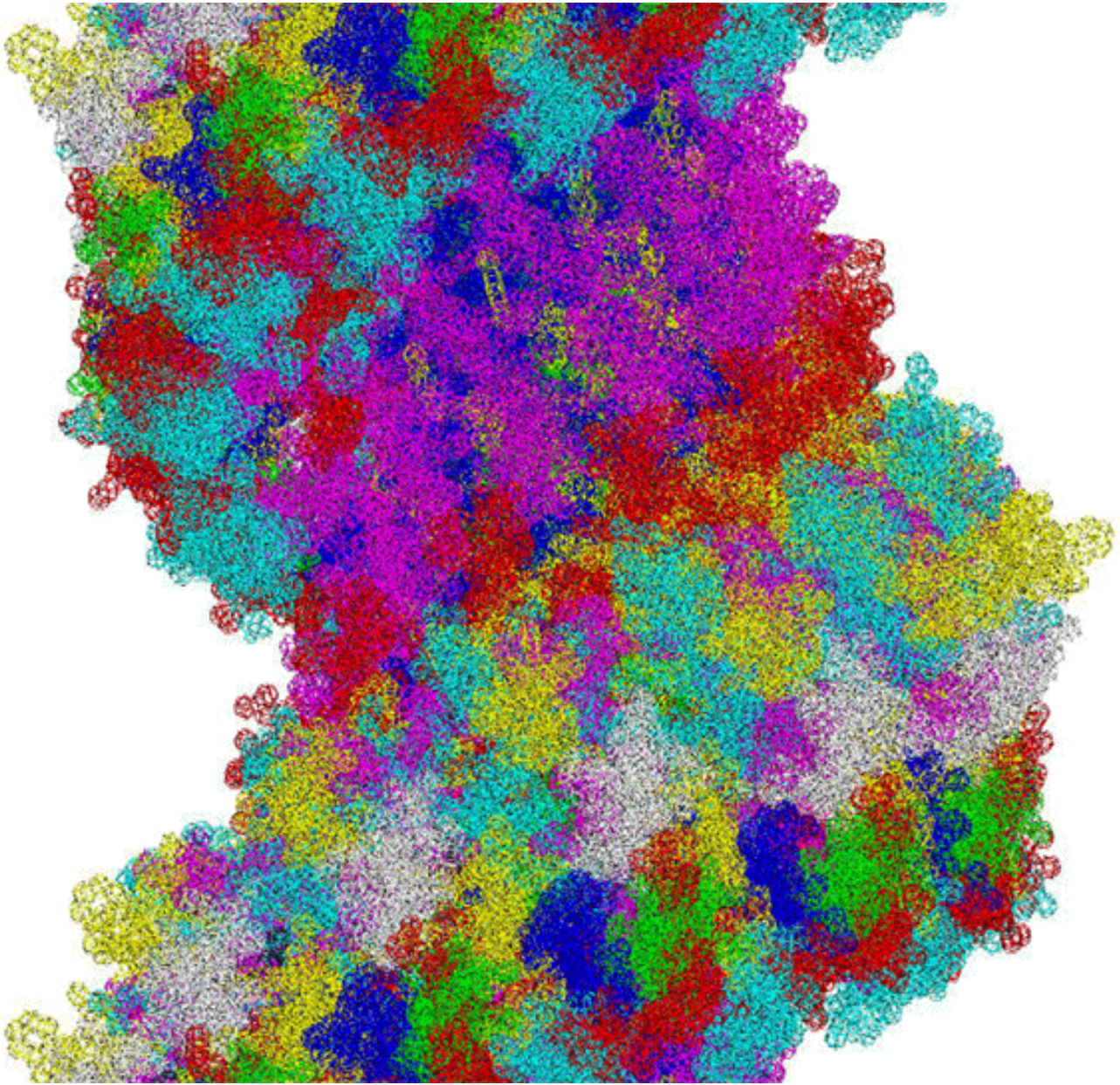
2019

No	DNA	ami	AMI		9var	Score	
1	AAA	K	LYS		3	F#1	
2	AAC	N	ASN		5	H1	
3	AAG	K	LYS		5	F#1	
4	AAT	N	ASN		3	H1	
5	ACA	T	THR		2	E2	
6	ACC	T	THR		5	E2	
7	ACG	T	THR		5	E2	
8	ACT	T	THR		3	E2	
9	AGA	R	ARG		3	D1	
10	AGC	S	SER		5	A2	
11	AGG	R	ARG		5	D1	
12	AGT	S	SER		3	A2	
13	ATA	I	ILE		2	C2	
14	ATC	I	ILE		5	C2	
15	ATG	M	MET		6	G1	
16	ATT	I	ILE		3	C2	
17	CAA	Q	GLN		3	G1	
18	CAC	H	HIS		5	A1	
19	CAG	Q	GLN		5	G1	
20	CAT	H	HIS		3	A1	
21	CCA	P	PRO		92	A3	
22	CCC	P	PRO		91	A3	
23	CCG	P	PRO		95	A3	
24	CCT	P	PRO		93	A3	
25	CGA	R	ARG		2	D1	
26	CGC	R	ARG		5	D1	
27	CGG	R	ARG		5	D1	
28	CGT	R	ARG		3	D1	
29	CTA	L	LEU		2	H1	
30	CTC	L	LEU		5	H1	
31	CTG	L	LEU		5	H1	
32	CTT	L	LEU		3	H1	
33	GAA	E	GLU		3	G1	
34	GAC	D	ASP		5	H1	
35	GAG	E	GLU		5	G1	
36	GAT	D	ASP		3	H1	
37	GCA	A	ALA		2	A3	
38	GCC	A	ALA		5	A3	
39	GCG	A	ALA		5	A3	
40	GCT	A	ALA		3	A3	
41	GGA	G	GLY		2	G0	
42	GGC	G	GLY		5	G0	
43	GGG	G	GLY		5	G0	
44	GGT	G	GLY		3	G0	
45	GTA	V	VAL		2	G2	
46	GTC	V	VAL		5	G2	
47	GTG	V	VAL		5	G2	
48	GTT	V	VAL		3	G2	
49	TAA		TKD		0		
50	TAC	Y	TYR		5	D1	
51	TAG		TKD		0		
52	TAT	Y	TYR		3	D1	
53	TCA	S	SER		2	A2	
54	TCC	S	SER		5	A2	
55	TCG	S	SER		5	A2	
56	TCT	S	SER		3	A2	
57	TGA		TKD		0		
58	TGC	C	CYS		5	E2	
59	TGG	W	TRP		5	D1	
60	TGT	C	CYS		3	E2	
61	TTA	L	LEU		3	H1	
62	TTC	F	PHE		5	G1	
63	TTG	L	LEU		5	H1	
64	TTT	F	PHE		3	G1	

The result of processing 62 genomes (approximately 6 million proteins)
was obtained in an hour '2025



3D MODEL OF COLLAGEN



Software helices

Fractal helices

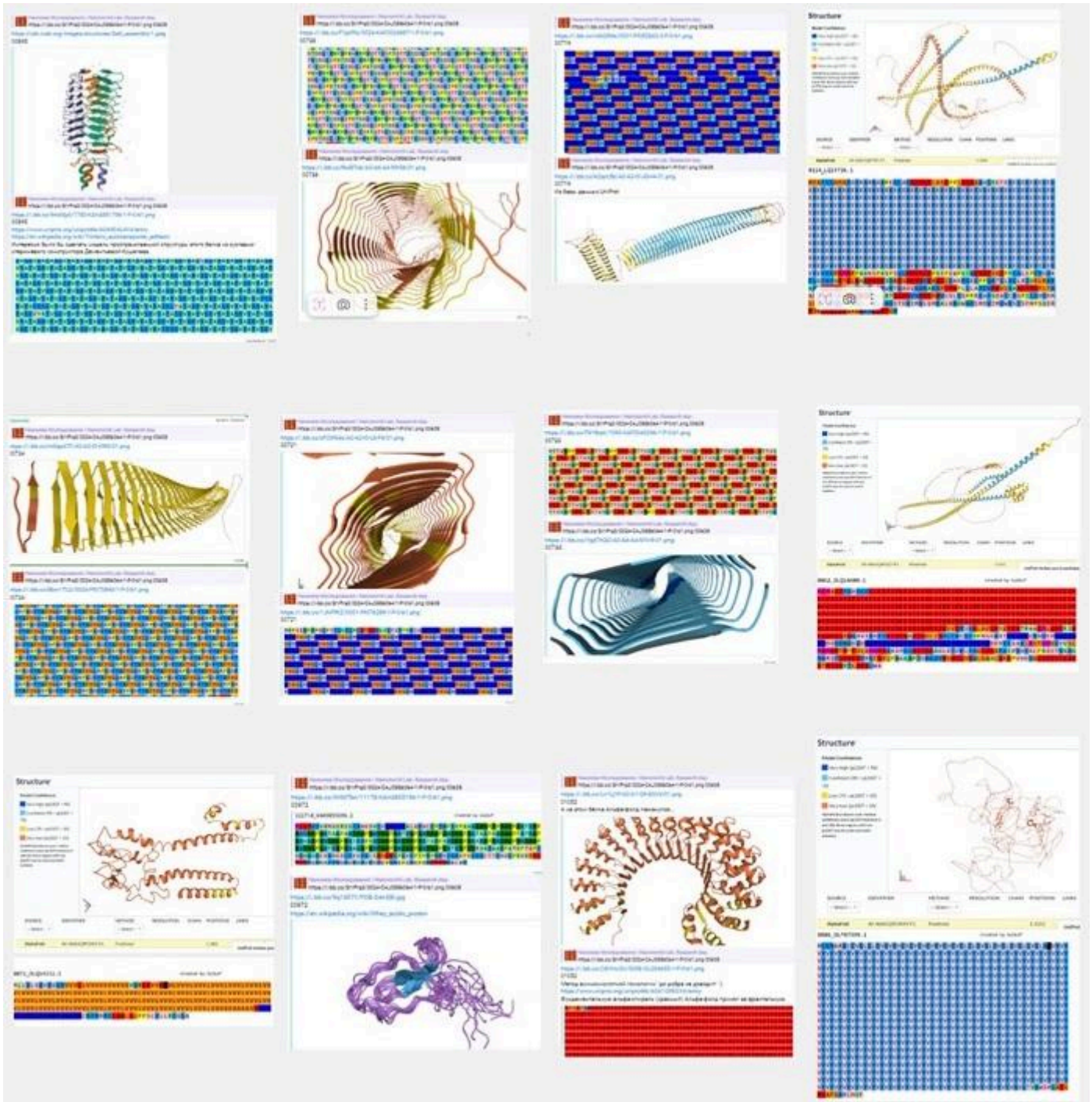
Q-helices

X-ray diffraction and 3D Genetic code

Examples of super-secondary structures

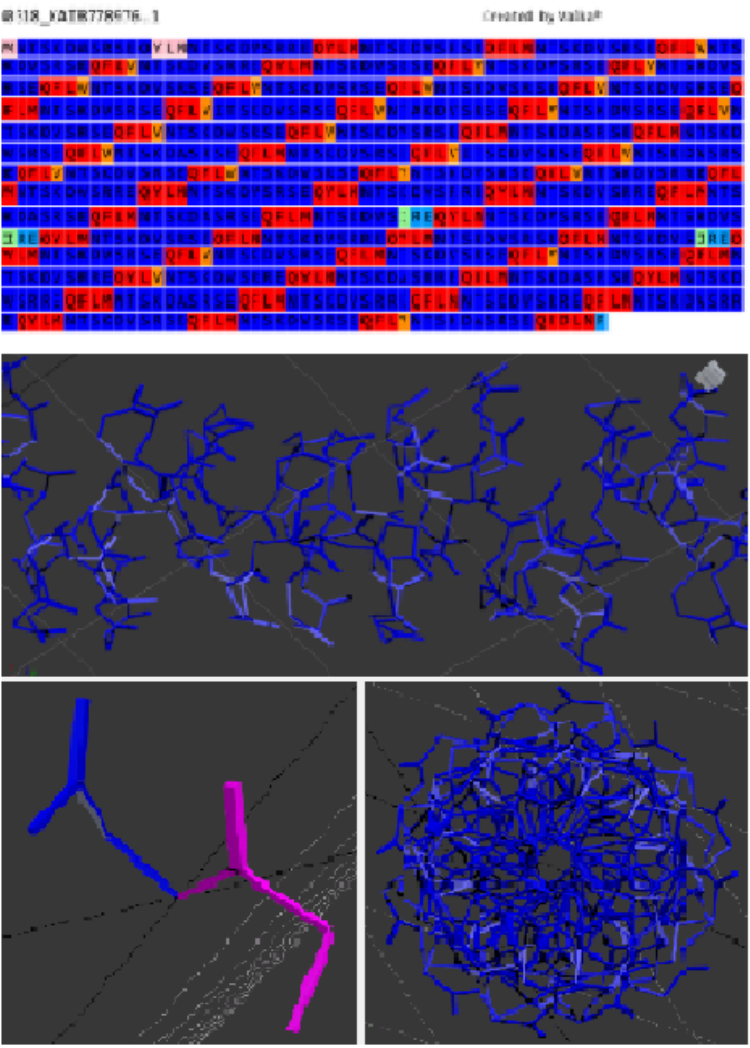
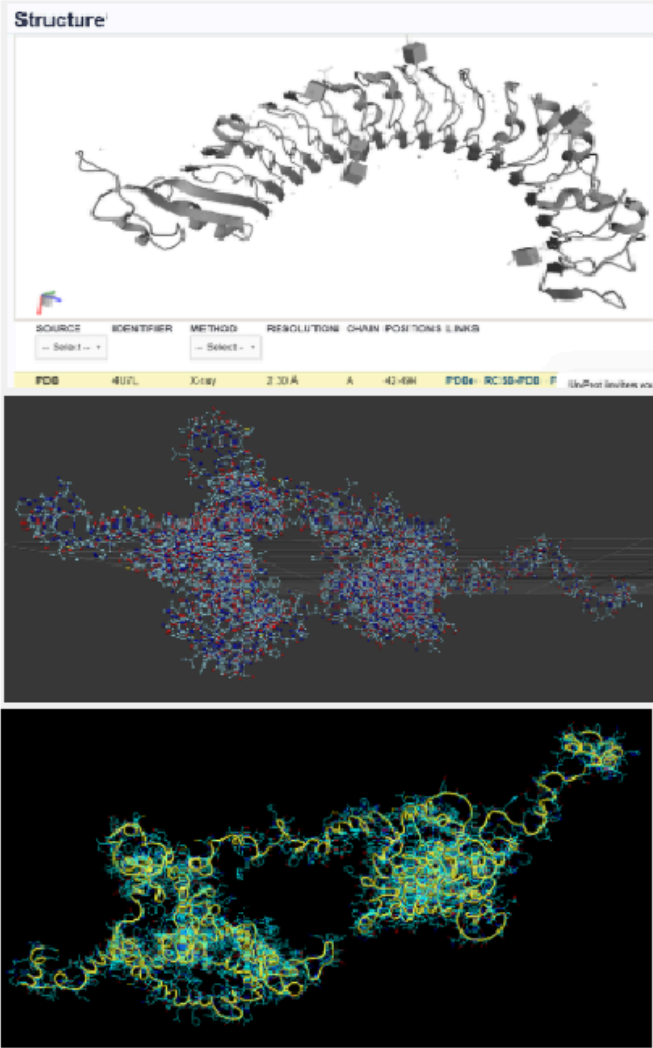
"Wet" experiments showed... fractal helices (helices of helices)

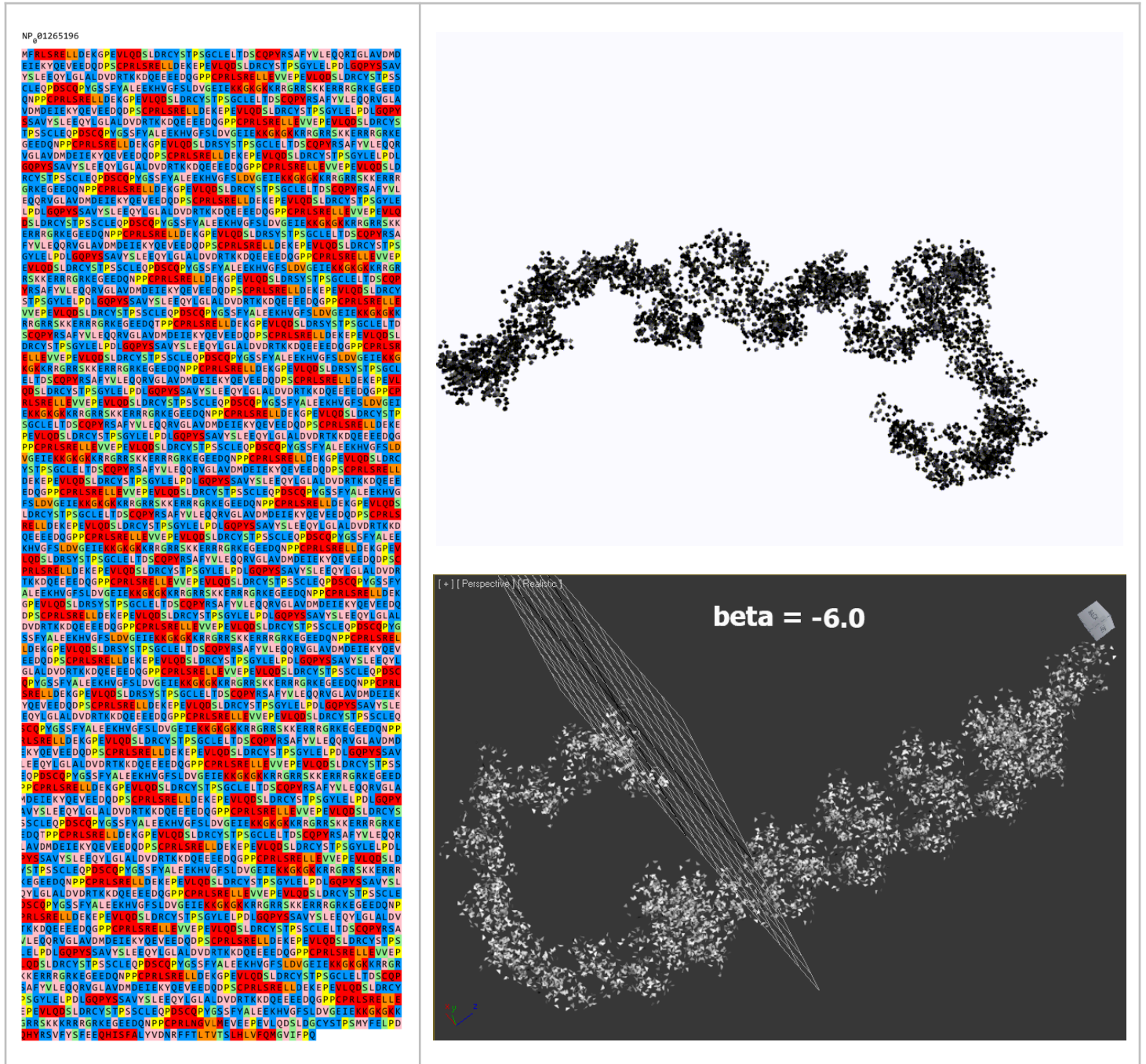
1



2

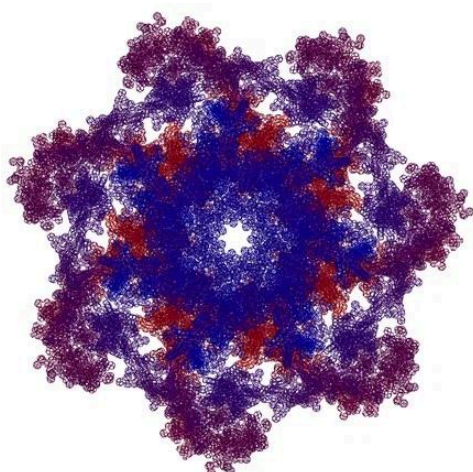
3





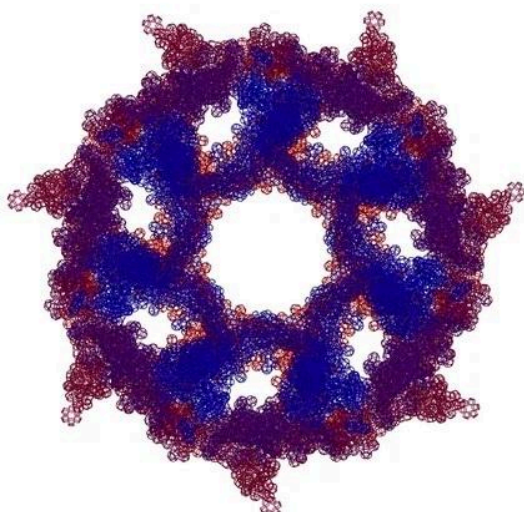
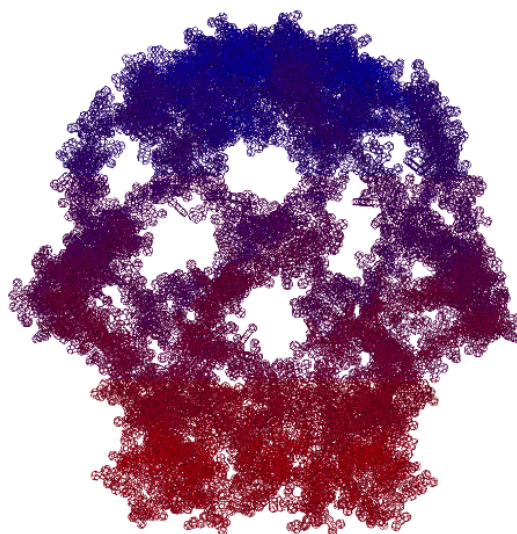


3D Picotech models of chaperones



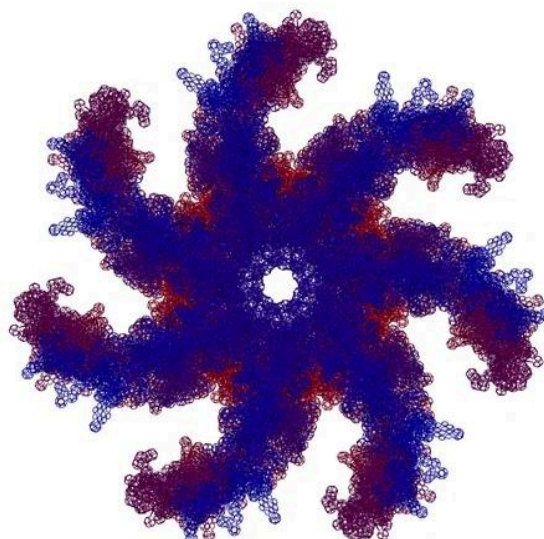
Пикотехнологическая модель
шаперона / chaperone A0KFQ4

Александр Кушелев, лаборатория Нанонир 2010(С)



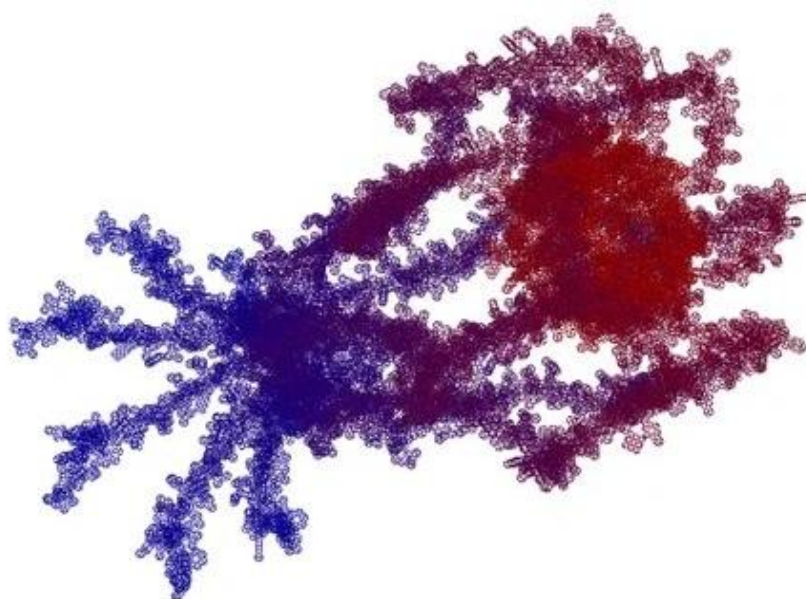
Пикотехнологическая модель
шаперона / chaperone B8GY03

Александр Кушелев, лаборатория Нанонир (С)2010

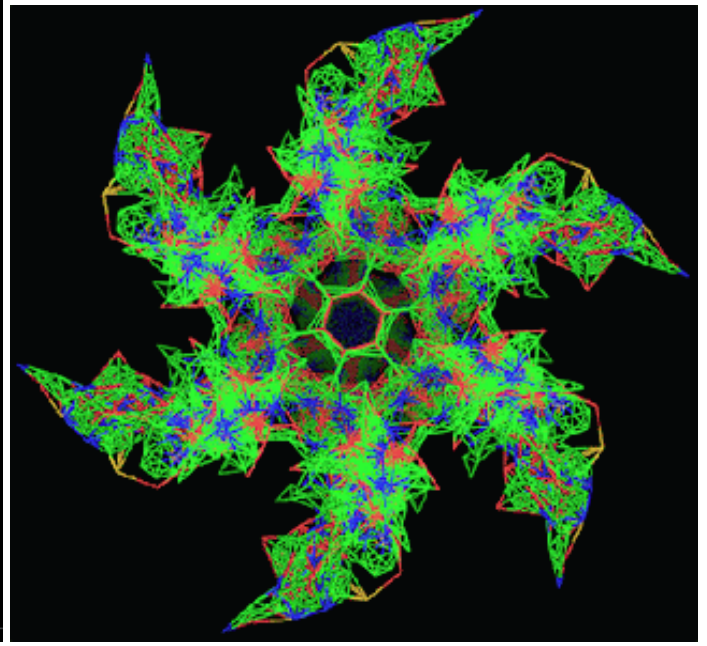
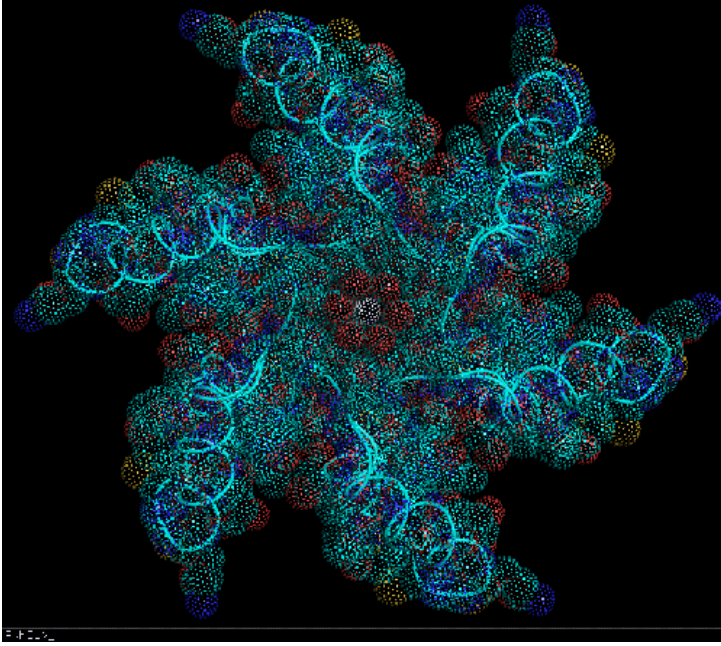


Пикотехнологическая модель
шаперона / chaperone B8H333

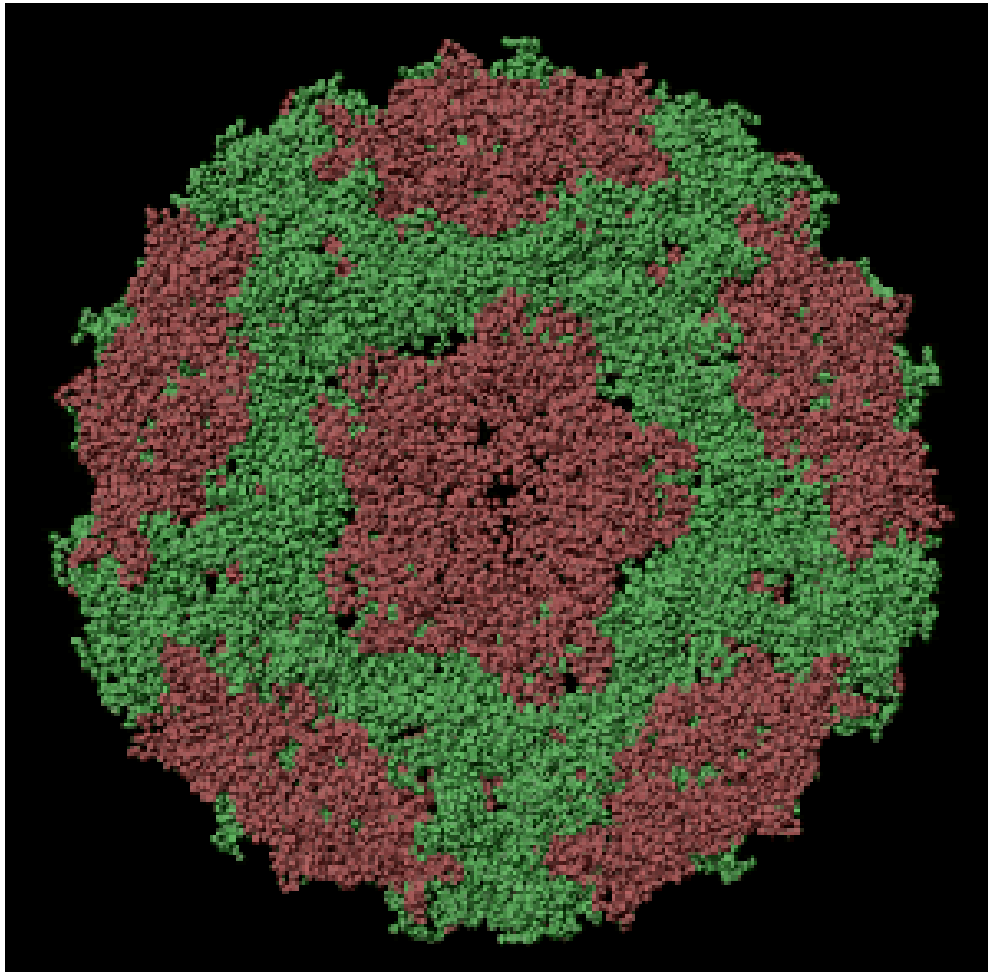
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3D model of insulin molecule



3D model of the virus shell



How exactly does this service work?

You send us the coding nucleotide sequence (ideally - in fasta format) of the protein you're interested in via email: nanoworld_laboratory@mail.ru.

The key advantages are:

1. Reliability and accuracy of protein secondary structure.

Below the 3D genetic code table, you can see a transcript showing that each triplet of the genetic code [corresponds to a specific structure](#).

2. The size of the protein is irrelevant.

3. The ability of the protein to crystallize is irrelevant.

4. The presence/absence of homologues is irrelevant.

5. The degree of study is irrelevant.

The disadvantages of this method include:

1. It does not account for protein post-processing after ribosome synthesis. The method only identifies the structure resulting from ribosome synthesis. If the protein is subsequently processed by chaperones or enzymes, meaning its secondary structure is altered (fragments are excised), I can only account for this manually by removing the enzyme-excised fragments of the amino acid sequence from the processing results. The client can do this themselves or provide me with data on the excised protein fragments.

2. Only secondary structures are automated. Tertiary structure can be automatically determined only for a narrow class of proteins, such as those consisting of a straight section of any type of helix. Therefore, only the secondary structure is guaranteed.

Tertiary structure and higher structures are "manual work," which can not only take longer but also does not guarantee its determination, for example, if the amino acid sequence contains prolines, which transform the rigid protein structure into a mechanism that can significantly change its conformation. In this case, the client can be provided with partial information on the tertiary and higher structures.

However, there is a high probability of determining both the tertiary and quaternary structures, [as well as the protein dynamics](#)

What exactly do clients receive when they contact us?

In what format are the results provided?

1. The client is guaranteed to receive a 2D Picotech diagram of the protein secondary structure.

2. With a certain degree of probability, the client will be able to obtain a 3D model from the joint-rod constructor.

3. If the 3D model is generated automatically or manually, the client will also be able to obtain a standard .pdb file with the coordinates of all protein atoms.

Unlike X-ray diffraction, the new technology has no "blind spots," meaning the PDB file will contain the coordinates of all atoms, from the first to the last amino acid residue.

Also, the client can obtain a [dynamic protein model](#), which provides insight into how the protein performs its functions.

Additionally, clients can receive a consultation in which we demonstrate the correlation between different analysis methods and our data. For example, X-ray diffraction (XRD) or alpha-fold analysis reveal the presence of helical regions, but do not specify their type. We can demonstrate where these methods correctly identified the beginning and end of a helical region, and where they erred, and why. In particular, interpreters often mistake hybrid (alpha-310-) regions of proteins for beta sheets. Picotechnological models can help clients understand why this occurs.

Moreover, we can search for protein active sites using the natural frequencies of amino acid side chains [1](#), [2](#), [3](#) (pp. 29-42). If any are detected, we can attempt to model the mechanism of these active sites at the picomechanical level, i.e., with picometric accuracy.

Thank you
very much
for your lenient
attention !