

F Element Project: Annotation Report

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College/university: California Polytechnic State University – San Luis Obispo

Course number: CHEM 441

Course name: Bioinformatics Applications

Authorship Information for GEP Scientific Publications

Initials: By entering my/our initials, I/we grant permission for the Genomics

Education Partnership (GEP) to use the annotation data produced in

RS/VY this report in future scientific publications.

Note: Please skip the rest of this section if <u>more than three students</u> contribute to this annotation report. When more than three students contribute to an annotation project, the class as a whole will be acknowledged in future GEP scientific publications.

Co-authors Responsibilities

In order to be a co-author on a GEP publication, you must review, critique, and approve the final gene models and manuscript, responding promptly to requests to read and approve. As part of the preparations for the microPublication article, co-authors are required to validate specific data within the manuscript, supplemental materials, and GenBank submission (the specific details will depend on each annotation project). In most cases, the manuscript preparation process will take approximately 3–5 hours of your time.

The above requirements mean that we must be able to contact you when the GEP microPublication, and later, the scientific paper with meta-analysis, is ready for your review and approval. If we cannot reach you at that time, you will not be a co-author on our GEP scientific publications, as scientific journals require all co-authors to have read and approved the manuscript.

Please provide your contact information below. Note that your name and contact information will be publicly available through the scientific publication and the GenBank record (this is standard for all scientific publications.). Please list the authors in ascending alphabetical order by last name. (The actual order of the student co-authors in the scientific publication will be determined by a random number generator.)

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Project Details

Project name: Contig60

Project species: D. Bipectinata
Date of submission: June 2, 2022
Size of project in base pairs: 49,290
Number of genes in project: 5

Does this report cover all of the genes or is it a partial report? Full Report

Note: For each gene described in this annotation report, you should also prepare the corresponding **GFF**, **transcript and peptide sequence files** as part of your submission.

Complete the following Gene Report Form for each gene in your project. Copy and paste the sections below to create as many copies as needed within this report. Be sure to create enough Isoform Report Forms within your Gene Report Form for all isoforms. If you cannot find evidence for any protein-coding genes in the project, jump to the "REF_Ref91436230 \h Check for additional features in your project" section.

Gene Report Form

Gene name (e.g., D. bipectinata eyeless): D. bipectinate CG13293 Gene symbol (e.g., dbip ey): <u>dbip</u> CG13293

Approximate location in project (from 5' end to 3' end): 4185-6917

Number of isoforms in D. melanogaster: 4

Number of isoforms in this project: 4

Complete the following table, including all of the isoforms in this project:

Name(s) of unique isoform(s) based	List of isoforms with identical coding sequences		
on coding sequence			
CG13293 – PC			
CG13293 – PB	CG13293 - PE		
CG13293 - PD			

Names of the isoforms with unique coding sequences in *D. melanogaster* that are absent in this species: N/A (Error Report Form removed from doc as there is no suggested change)

Isoform Report Form

Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

Gene-isoform symbol: dbip 13293-PB

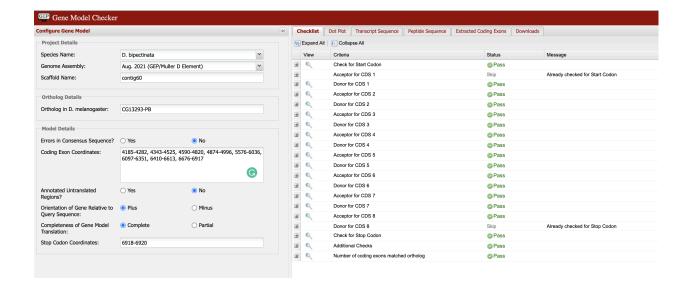
Names of any additional isoforms with identical coding sequences: <u>dbip 13293-PE</u>

Is the 5' end of this isoform missing from the end of the project? <u>NO</u> Is the 3' end of this isoform missing from the end of the project? <u>NO</u>

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.

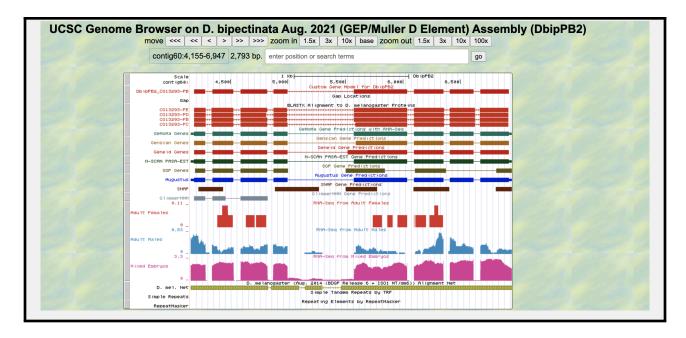


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the *GEP UCSC Genome Browser*. Zoom in so that <u>only this isoform is in</u> <u>the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

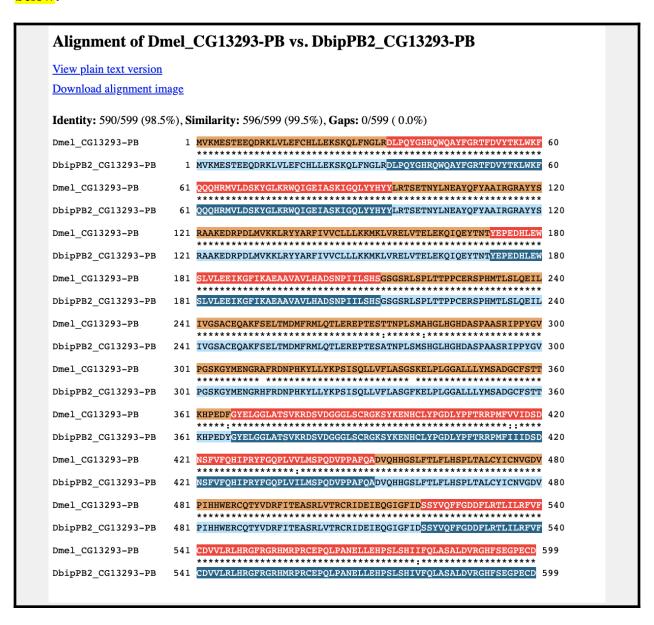
- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 3. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

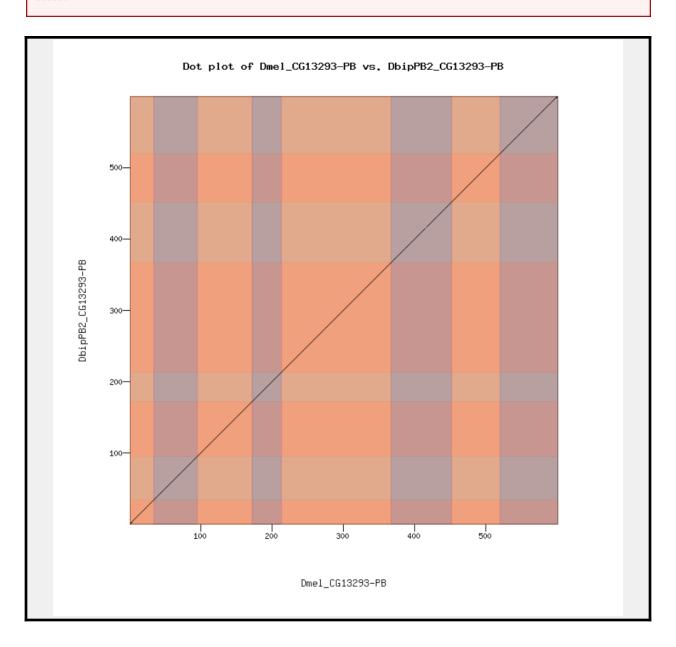
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Isoform Report Form

Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

Gene-isoform symbol: dbip 13293-PC

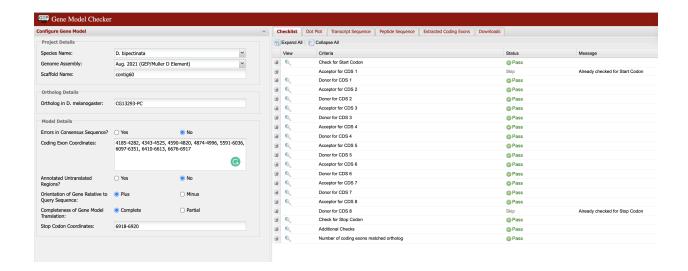
Names of any additional isoforms with identical coding sequences:

Is the 5' end of this isoform missing from the end of the project? <u>NO</u> Is the 3' end of this isoform missing from the end of the project? <u>NO</u>

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.



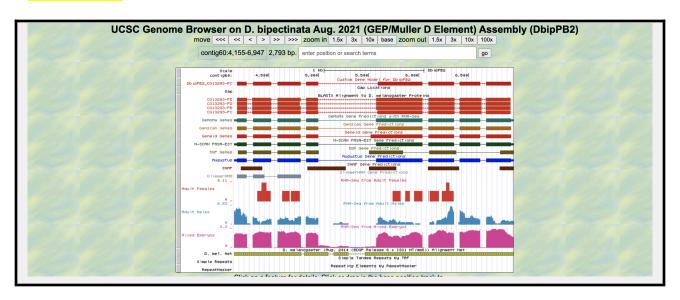
2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the *GEP UCSC Genome Browser*. Zoom in so that <u>only this isoform is in</u>

the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:

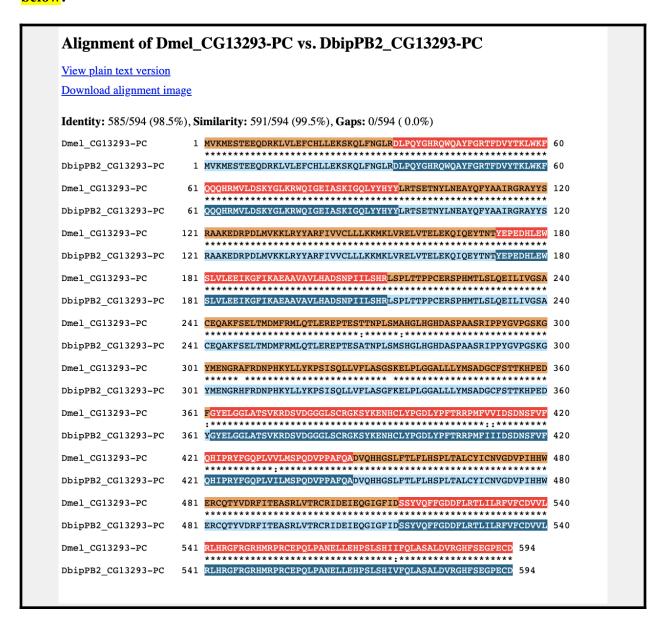
- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 3. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

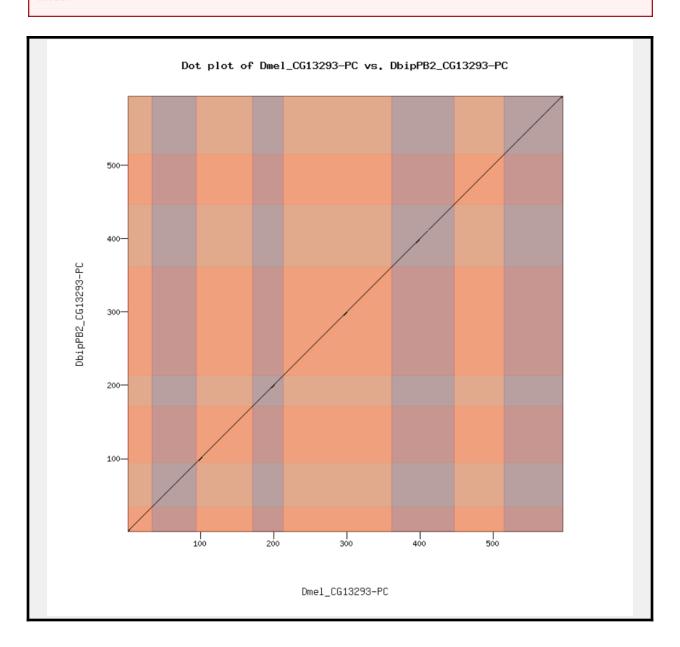
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Isoform Report Form

Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

Gene-isoform symbol: dbip 13293-PD

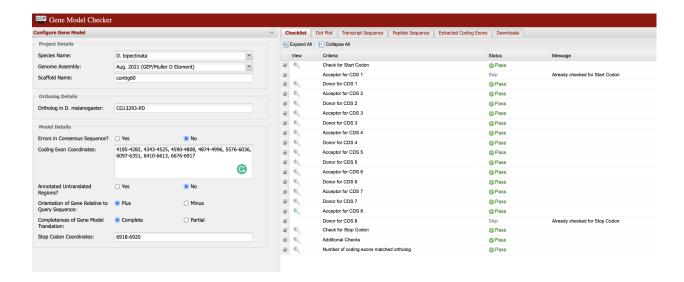
Names of any additional isoforms with identical coding sequences:

Is the 5' end of this isoform missing from the end of the project? <u>NO</u> Is the 3' end of this isoform missing from the end of the project? <u>NO</u>

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.



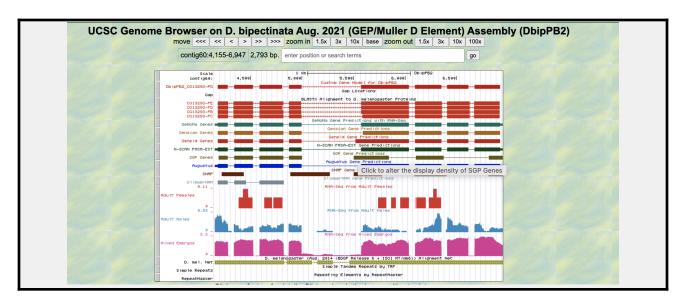
2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the *GEP UCSC Genome Browser*. Zoom in so that <u>only this isoform is in</u>

the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:

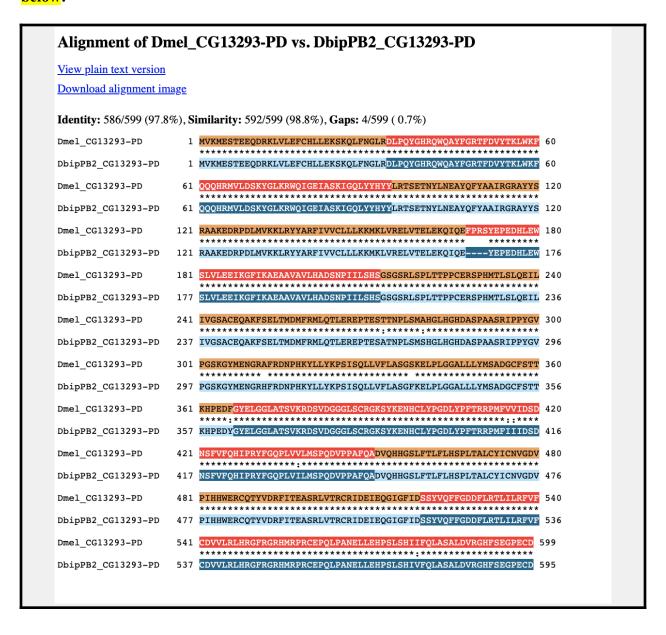
- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 3. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

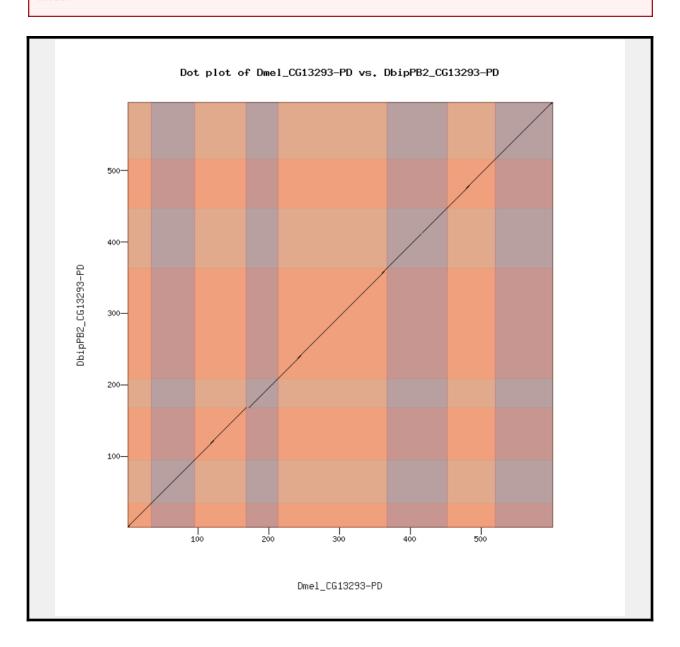
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Transcription Start Sites (TSS) Report Form (Optional)

Note: Complete this section if you have annotated the TSS for the gene above. This section is **optional** and you do not need to complete this section to submit the project.

Gene name: D. bipectinata CG13293

Gene symbol: dbip CG13293_

Name(s) of isoform(s) with unique TSS	List of isoforms with identical TSS
CG13293 - PB	CG13293 - PE
	CG13293 - PD
	CG13293 - PC

Names of the isoforms with unique TSS in D. melanogaster that are absent in this species: N/A

Isoform TSS Report

Complete an Isoform TSS report (through page PAGEREF _Ref91436230 \h 17) for each unique TSS listed in the table above. Copy and paste this form to create as many copies as needed.

Gene-isoform name: dbip CG13293 - PB

Names of the isoforms with the same TSS as this isoform: dbip CG13293 – PE, dbip CG13293 – PD, dbip CG13293 - PC

Type of core promoter in *D. melanogaster* (see table below): (Peaked / Intermediate / Broad / Insufficient Evidence)

<u>Peaked</u>

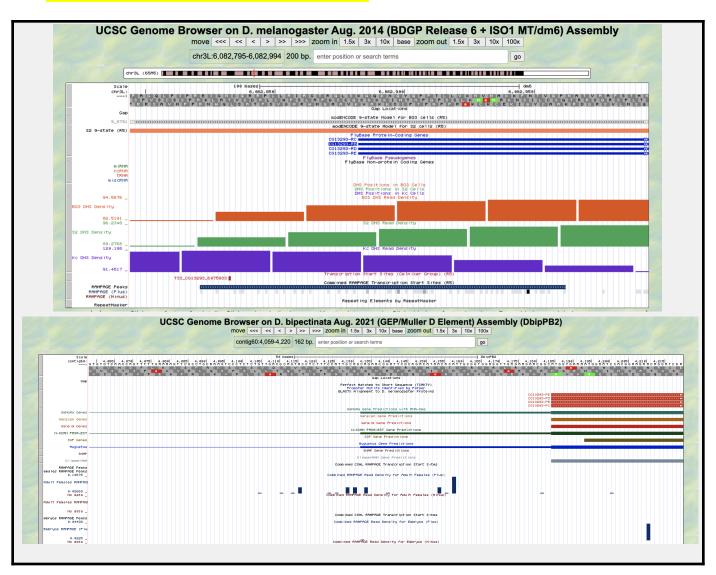
The type of core promoter is defined by the number of TSS annotated by the Celniker group at modENCODE and the number of DHS positions:

1. Turn on RAMPAGE evidence tracks (Only applies to projects with these tracks)

Coordinates of the TSS position based on position with the highest RAMPAGE read density Bipectinata: 4105-4182

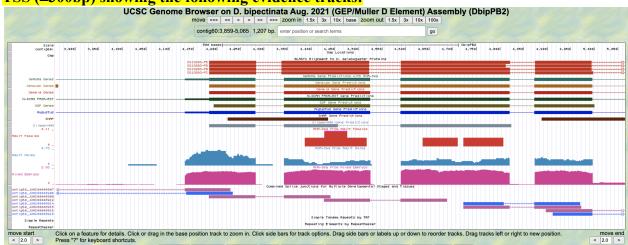
Coordinates of the narrow TSS search region based on RAMPAGE peaks Bipectinata: 3,767

If the TSS position and narrow TSS search region are supported by RAMPAGE data, paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the Combined RAMPAGE TSS evidence track:



2. Turn on RNA-Seq evidence tracks (N/A for this Gene)

If the TSS annotation is supported by RNA-Seq read coverage or splice junction predictions (e.g., regtools), paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the following evidence tracks:



- 1. RNA-Seq Coverage or RNA-Seq Alignment Summary
- 2. Combined Splice Junctions or RNA-Seq TopHat

If the RNA-Seq evidence tracks indicate a TSS search region, list it here: 4133-4184

3. Annotate the first transcribed exon

Coordinates of the first transcribed exon based on BLASTN alignment: 279-406 (suggests TSS at 279 which makes no sense = refute)

Does the BLASTN alignment cover the entire D. melanogaster first transcribed exon? \underline{NO}

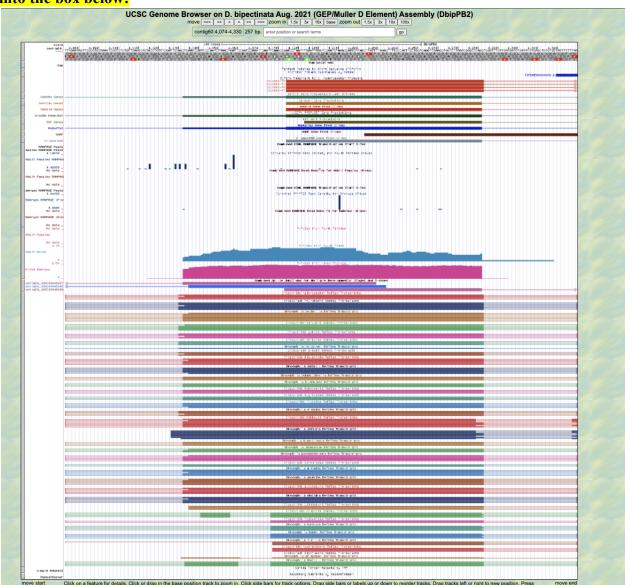
If not, specify the parts of the *D. melanogaster* exon that are missing from the *BLASTN* alignment.

Query length of the entire exon is 471. BLATN covers 1-128

If the TSS annotation is supported by *BLASTN* alignment of the initial transcribed exon against the contig sequence, paste a screenshot of the *BLASTN* alignment into the box below:

4. Turn on comparative genomics tracks

If the TSS annotation is supported by sequence conservation with other *Drosophila* species, paste a screenshot of the multiple sequence alignment (e.g., from Clustal Omega, ROAST) into the box below:



5. Summarize the evidence that supports the TSS annotation postulated above

Coordinate(s) of the TSS position(s):

Based on RAMPAGE data (if applicable): <u>4105-4182</u> (<u>also corresponds to the same region in respect to first exon in Melanogaster RAMPAGE</u>)

Based on RNA-Seq data: **4133-4184** Based on *BLASTN* alignment: 279

Note: If the *BLASTN* alignment for the initial transcribed exon is a partial alignment, you can **extrapolate the TSS position** based on the number of nucleotides that are missing from the beginning of the exon. (Enter "Insufficient evidence" if you cannot determine the TSS position based on the available evidence.)

Were you able to define a TSS search region based on the available evidence? YES, 4105-4184 with most evidence pointing to 4133 as the TSS

If so, indicate in the table below the evidence that supports this TSS position

For each evidence type, enter an "X" in the cell to indicate whether the line of evidence supports, refutes, or neither supports nor refutes the TSS annotation:

Evidence type	Support	Refute	Neither
RAMPAGE peaks and read density	X		
RNA-Seq coverage and splice junctions	X		
BLASTN alignment of the initial exon from D. melanogaster		Х	
Sequence conservation with other <i>Drosophila</i> species (<i>e.g.</i> , "Conservation" track on the Genome Browser)	X		

Note: The evidence type refutes the TSS annotation only if it suggests an alternate TSS position. For example, the presence of RNA-Seq read coverage upstream of the annotated TSS indicates that the TSS is located further upstream and it would be considered to be evidence against the annotated TSS; check "Refute." In contrast, the lack of RNA-Seq read coverage is a negative result that neither supports nor refutes the TSS annotation; check "Neither."

Provide an explanation if the TSS annotation is inconsistent with at least one of the evidence types specified above:

Gene Report Form

Gene symbol: _dbip_CG10469_

Approximate location in project (from 5' end to 3' end): 9014-9871

Number of isoforms in *D. melanogaster*: 1

Number of isoforms in this project: 1

Complete the following table, including all of the isoforms in this project:

Name(s) of unique isoform(s) based on coding sequence	List of isoforms with identical coding sequences		
CGG10469 - PA			

Names of the isoforms with unique coding sequences in *D. melanogaster* that are absent in this species: N/A (report form deleted from this document because of N/A)

Isoform Report Form

Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

Gene-isoform symbol: <u>dbip_CGG10469 - PA</u>

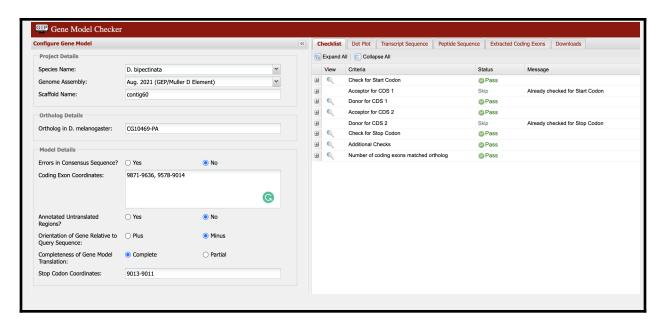
Names of any additional isoforms with identical coding sequences:

N/A

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.

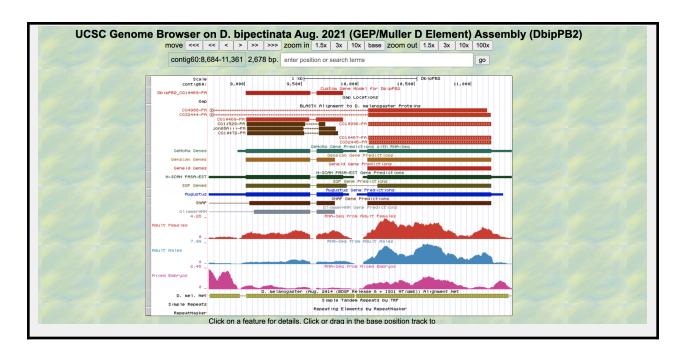


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the *GEP UCSC Genome Browser*. Zoom in so that <u>only this isoform is in</u> <u>the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

- 5. A sequence alignment track (e.g., D. mel Proteins)
- 6. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 7. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 8. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

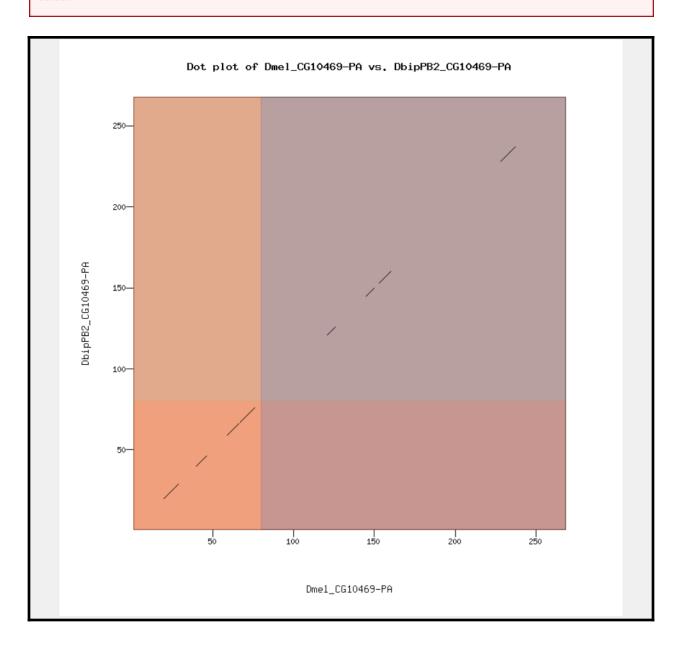
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:

Alignment of Dmel_CG10469-PA vs. DbipPB2_CG10469-PA View plain text version Download alignment image **Identity:** 160/267 (59.9%), **Similarity:** 203/267 (76.0%), **Gaps:** 0/267 (0.0%) Dmel CG10469-PA 1 MILQLVLIVQFSLVFGQETGSLRIMNGTAAKAKQLPYQVGLLCYFEGSKDEPNMCGGTIL 60 ****:.*:* ::* *.*: ******* * :*:**:.***** * ::*.:**:** DbipPB2_CG10469-PA 1 MILQFFLFVPITLGFAQDLGSLRIMNGTYAVEEQFPYEAGLLCYFAGFPNNPSLCGGAIL 60 Dmel CG10469-PA 61 SNRWIITAAHCLQDPKSNLWKVLIHVGKVKSFDDKEIVVNRSYTIVHKKFDRKTVTNDIA 120 DbipPB2 CG10469-PA 61 SNRWILTAAHCLQDPDANLTQVRVQVGSLEAPGGDDILVNGSDIIVHKNFNRKTVFNDLG 120 Dmel CG10469-PA 121 LIKLPKKLTFNKYIQPAKLPSAKKTYTGRKAIISGWGLTTKQLPSQVLQYIRAPIISNKE 180 *****::***.. :**.***: :*****..:****** :* .*: ***:*: ::***: DbipPB2_CG10469-PA 121 LIKLPRNLTFSSKVQPVKLPSSYRTYTGRSVFISGWGLTDNQTASESLQYLRTDVVSNKQ 180 Dmel CG10469-PA 181 CERQWNKQLGGKSKKVVHNGFICIDSKKGLPCRGDSGGPMVLDDGSRTLVGIVSHGFDGE 240 181 CQSQWNKALKGKKKKVVSWTFVCVDTQQGMPCQGDSGSPMVLADGSKTLVGIVSHGLDPE 240 DbipPB2 CG10469-PA Dmel CG10469-PA 241 CKLKLPDVSTRVSSYLKWIKYYSGGLK 267 ** *:**:* ***::*:*: :*** DbipPB2 CG10469-PA 241 CKRKVPDISMRVSAFLRWINSNTGGLK 267

4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Transcription Start Sites (TSS) Report Form (Optional)

Note: Complete this section if you have annotated the TSS for the gene above. This section is **optional** and you do not need to complete this section to submit the project.

Gene name: D. bipectinate CG10469

Gene symbol: <u>dbip CG10469</u>

Name(s) of isoform(s) with unique TSS List of isoforms with identical TSS CG10469-PA

Names of the isoforms with unique TSS in *D. melanogaster* that are absent in this species: N/A

Provide the evidence (text and figures) which support the hypothesis that these isoforms are absent in this species (*e.g.*, changes in canonical splice sites, gene structure, etc.):

Isoform TSS Report

Complete an Isoform TSS report (through page PAGEREF _Ref91436230 \h 17) for each unique TSS listed in the table above. Copy and paste this form to create as many copies as needed.

Gene-isoform name: <u>dbip CGG10469 - PA</u>

Names of the isoforms with the same TSS as this isoform: N/A

Type of core promoter in *D. melanogaster* (see table below): (Peaked / Intermediate / Broad / Insufficient Evidence)

<u>Peaked</u>

The type of core promoter is defined by the number of TSS annotated by the Celniker group at modENCODE and the number of DHS positions:

1. Turn on RAMPAGE evidence tracks (Only applies to projects with these tracks)

Coordinates of the TSS position based on position with the highest RAMPAGE read density Bipectinata: 10023-9877

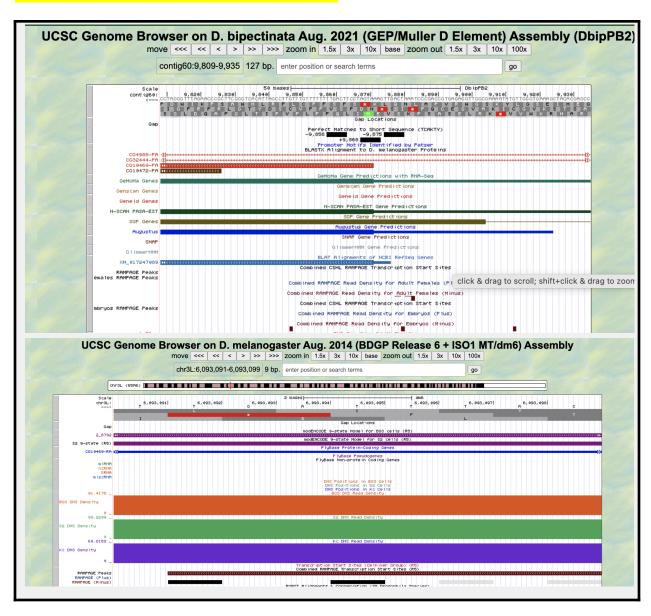
Melanogaster: 6,093,098-6,093,091

Coordinates of the narrow TSS search region based on RAMPAGE peaks

Bipectinata: 9880

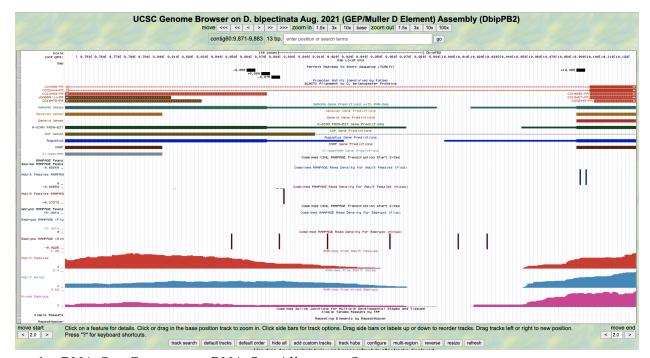
Melanogaster: 6,093,095 and 6,093,092

If the TSS position and narrow TSS search region are supported by RAMPAGE data, paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the Combined RAMPAGE TSS evidence track:



2. Turn on RNA-Seq evidence tracks

If the TSS annotation is supported by RNA-Seq read coverage or splice junction predictions (e.g., regtools), paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the following evidence tracks:



- 1. RNA-Seq Coverage or RNA-Seq Alignment Summary
- 2. Combined Splice Junctions or RNA-Seq *TopHat*

If the RNA-Seq evidence tracks indicate a TSS search region, list it here: downstream of 9950

3. Annotate the first transcribed exon

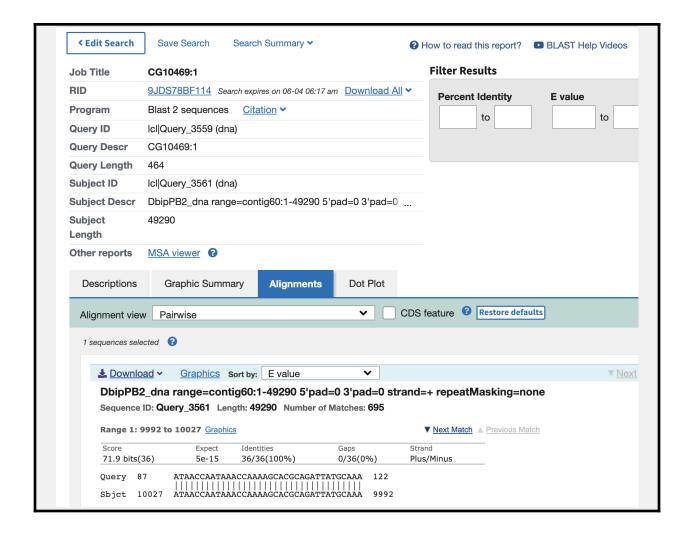
Coordinates of the first transcribed exon based on *BLASTN* alignment: 10027-9992

Does the *BLASTN* alignment cover the entire *D. melanogaster* first transcribed exon? No

If not, specify the parts of the *D. melanogaster* exon that are missing from the *BLASTN* alignment.

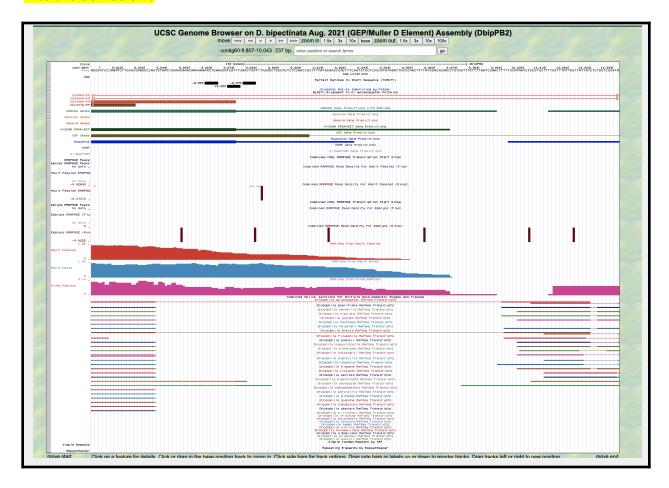
Covers 87-122. Missing 1-187 and 88-464

If the TSS annotation is supported by *BLASTN* alignment of the initial transcribed exon against the contig sequence, **paste a screenshot of the** *BLASTN* **alignment into the box below:**



4. Turn on comparative genomics tracks

If the TSS annotation is supported by sequence conservation with other *Drosophila* species, paste a screenshot of the multiple sequence alignment (e.g., from Clustal Omega, ROAST) into the box below:



5. Summarize the evidence that supports the TSS annotation postulated above

Coordinate(s) of the TSS position(s):

Based on RAMPAGE data (if applicable): <u>10023-9877</u>

Based on RNA-Seq data: N/A (unable to get RNA Seq alignment)

Based on *BLASTN* alignment: 10027-9992

Based on other evidence (ShortMatch (TCAKTY): 9873

Note: If the *BLASTN* alignment for the initial transcribed exon is a partial alignment, you can **extrapolate the TSS position** based on the number of nucleotides that are missing from the beginning of the exon. (Enter "Insufficient evidence" if you cannot determine the TSS position based on the available evidence.)

Were you able to define a TSS position based on the available evidence? NO

If not, were you able to define a TSS search region? YES, 10023-9873 If so, indicate in the table below the evidence that supports the TSS search region(s)

For each evidence type, enter an "X" in the cell to indicate whether the line of evidence supports,

refutes, or neither supports nor refutes the TSS annotation:

Evidence type	Support	Refute	Neither
RAMPAGE peaks and read density	X		
RNA-Seq coverage and splice junctions			X
BLASTN alignment of the initial exon from D. melanogaster	Х		
Sequence conservation with other <i>Drosophila</i> species (e.g., "Conservation" track on the Genome Browser)	Х		
Other (ShortMatch (TCAKTY))	X		

Note: The evidence type refutes the TSS annotation only if it **suggests an alternate TSS position**. For example, the presence of RNA-Seq read coverage upstream of the annotated TSS indicates that the TSS is located further upstream and it would be considered to be evidence against the annotated TSS; check "Refute." In contrast, the lack of RNA-Seq read coverage is a negative result that neither supports nor refutes the TSS annotation; check "Neither."

Provide an explanation if the TSS annotation is inconsistent with at least one of the evidence types specified above:

Gene Report Form

Gene name: D. bipectinata CG10467

Gene symbol: <u>dbip_CG10467</u>

Approximate location in project (from 5' end to 3' end): 10094-11179

Number of isoforms in D. melanogaster: 1

Number of isoforms in this project: 1

Complete the following table, including all of the isoforms in this project:

Name(s) of unique isoform(s) based on coding sequence	d List of isoforms with identical coding sequences	
CG10467-PA		

Names of the isoforms with unique coding sequences in *D. melanogaster* that are absent in this species: N/A (removed report form from document due to N/A)

Isoform Report Form

Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

Gene-isoform symbol: dbip CG10467 - PA

Names of any additional isoforms with identical coding sequences:

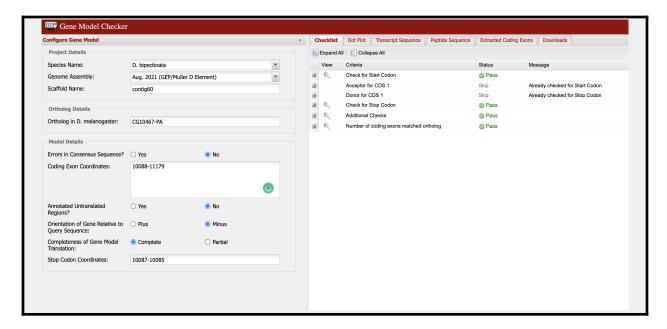
N/A

Is the 5' end of this isoform missing from the end of the project? NO Is the 3' end of this isoform missing from the end of the project? NO

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.

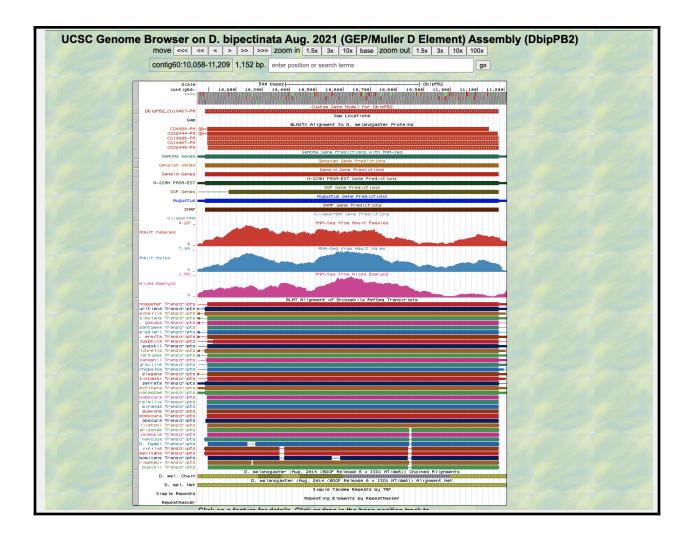


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the <u>GEP UCSC Genome Browser</u>. Zoom in so that <u>only this isoform is in</u> <u>the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

- 3. A sequence alignment track (e.g., D. mel Proteins)
- 4. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 5. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 6. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

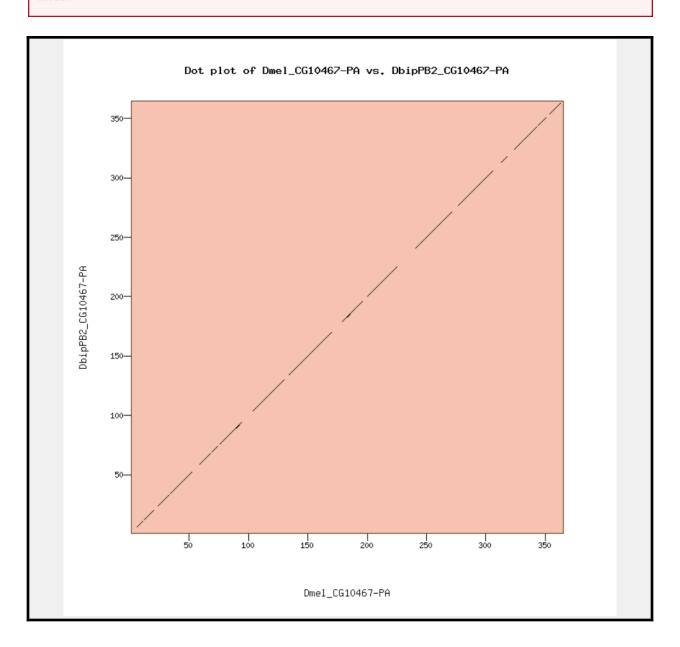
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:

Alignment of Dmel_CG10467-PA vs. DbipPB2_CG10467-PA View plain text version Download alignment image **Identity:** 313/364 (86.0%), **Similarity:** 336/364 (92.3%), **Gaps:** 0/364 (0.0%) Dmel CG10467-PA 1 MVNVTEDVFANGAVNPFTKSKDTIKRFTLTNGAGMSVQLITRGATITSIKTPDASGQIDD 60 ****.********************************* DbipPB2_CG10467-PA 1 MVNVKEDVFATGAVNPFTKATEDIKRFTLTNGYGMSVQLITRGATITSIKTMDSSGKVDD 60 61 VTLGFDDLAGYQSERNPYFGATIGRVCNRIANGSFYLDGKLVQVSKNRDNKFQLHGGFVG 120 Dmel_CG10467-PA *******:****************** DbipPB2_CG10467-PA 61 VTLGFDDVAGYQSDKNPYFGATIGRVCNRIANGRFQLNGKWIEVSKNRNNKFQLHGGFVG 120 Dmel_CG10467-PA 121 FDKAHWEVVEVRVDGVTLSHTNPDGHEGYPGKVTATASFTLSEDNCLHVQMSALADKTTP 180 121 FDKAHWEVVAVRRDGVTLSHTNPDGHEGYPGKVTATASFTLSEDNCLHVLMTAETDKVTP 180 DbipPB2 CG10467-PA Dmel_CG10467-PA 181 VNLTNHSYFNLAGHKSGANGLYEHTIEINAYGITETDQSSIPTGRITPVEGTGFDLRVSS 240 DbipPB2_CG10467-PA 181 VNLTNHSYFNLAGHKTGASGLYEHAIEINAYGITETDQDSIPTGKITPVNGTPYDLRVAG 240 Dmel_CG10467-PA 241 NLGERLKALQPARGYDDNFCVTFSPPQPLAKVARATHPPSGRWLEVVSNQPGVQFYTSNF 300 ******************* DbipPB2_CG10467-PA 241 NLGERLKALQPARGYDDNFCVTFSPPQPLAMVARASHPPSGRWLEVVSNQPGVQFYTSNF 300 Dmel_CG10467-PA 301 MPDVERGESPIPGKDGAAYAKHCAFCLETQKFPDSVNHSNFPSTILRPGESYQHEVIYKF 360 301 MPDVENGEAPISGKDGASYGKHGAFCLETQKFPDSVNHSNFPTTILRPGERYSHEVIYKF 360 DbipPB2_CG10467-PA 361 GVSH 364 Dmel_CG10467-PA ** : DbipPB2 CG10467-PA 361 GVFN 364

4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Transcription Start Sites (TSS) Report Form (Optional)

Note: Complete this section if you have annotated the TSS for the gene above. This section is **optional** and you do not need to complete this section to submit the project.

Gene name: D. Bipectinata CG10467

Gene symbol: dbip CG10467

Name(s) of isoform(s) with unique TSS List of isoforms with identical TSS CG10467-PA

Names of the isoforms with unique TSS in *D. melanogaster* that are absent in this species: N/A

Provide the evidence (text and figures) which support the hypothesis that these isoforms are absent in this species (*e.g.*, changes in canonical splice sites, gene structure, etc.):

Isoform TSS Report

Complete an Isoform TSS report (through page PAGEREF _Ref91436230 \h 17) for each unique TSS listed in the table above. Copy and paste this form to create as many copies as needed.

Gene-isoform name : dbip CG10467-pA

Names of the isoforms with the same TSS as this isoform:

N/A

Type of core promoter in *D. melanogaster* (see table below): (Peaked / Intermediate / Broad / Insufficient Evidence)

<u>Peaked</u>

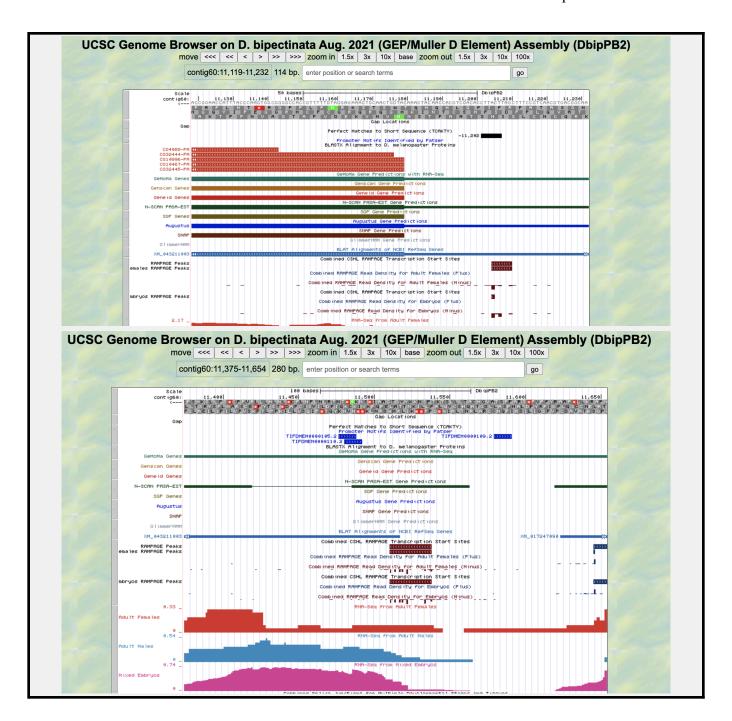
The type of core promoter is defined by the number of TSS annotated by the Celniker group at modENCODE and the number of DHS positions:

1. Turn on RAMPAGE evidence tracks (Only applies to projects with these tracks)

Coordinates of the TSS position based on position with the highest RAMPAGE read density <u>11598-11161</u>

Coordinates of the narrow TSS search region based on RAMPAGE peaks 11538-11510 and 11210-11204

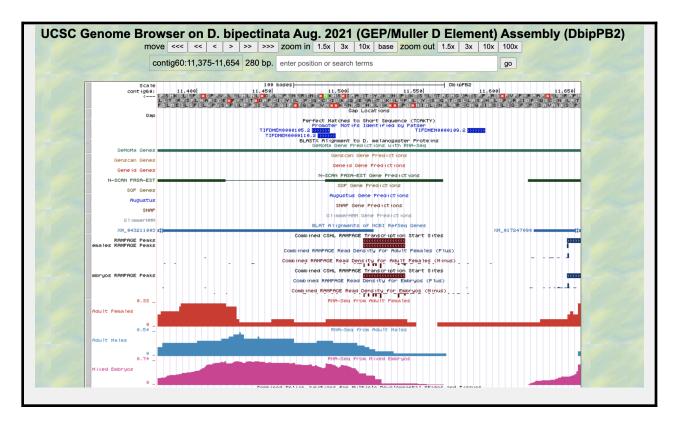
If the TSS position and narrow TSS search region are supported by RAMPAGE data, paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the Combined RAMPAGE TSS evidence track:



2. Turn on RNA-Seq evidence tracks

If the TSS annotation is supported by RNA-Seq read coverage or splice junction predictions (e.g., regtools), paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the following evidence tracks:

- 7. RNA-Seq Coverage or RNA-Seq Alignment Summary
- 8. Combined Splice Junctions or RNA-Seq *TopHat*



If the RNA-Seq evidence tracks indicate a TSS search region, list it here: Adult Females/Males and mixed embryos because they are starting near the RAMPAGE site showing that TSS could be starting in this region.

3. Annotate the first transcribed exon

Coordinates of the first transcribed exon based on *BLASTN* alignment: 10101-11279

Does the BLASTN alignment cover the entire D. melanogaster first transcribed exon? NO

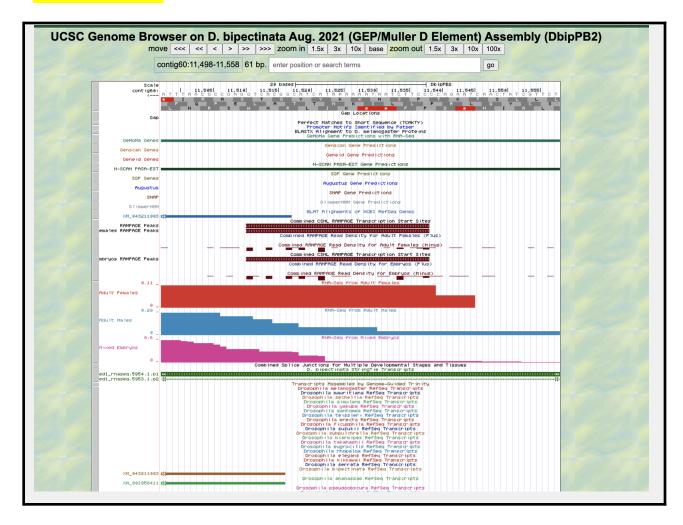
If not, specify the parts of the *D. melanogaster* exon that are missing from the *BLASTN* alignment.

Covers 253-1431, missing 1-253 and 1431-1544

If the TSS annotation is supported by *BLASTN* alignment of the initial transcribed exon against the contig sequence, **paste a screenshot of the** *BLASTN* **alignment into the box below:**

4. Turn on comparative genomics tracks

If the TSS annotation is supported by sequence conservation with other *Drosophila* species, paste a screenshot of the multiple sequence alignment (e.g., from Clustal Omega, ROAST) into the box below:



5. Summarize the evidence that supports the TSS annotation postulated above

Coordinate(s) of the TSS position(s):

Based on RAMPAGE data (if applicable): range: 11598-11161 peak: 11538-11510

Based on RNA-Seq data: near 11545

Based on *BLASTN* alignment: 10101-11279

Based on other evidence (Shortmatch(TCAKTY))): 11,200

Note: If the *BLASTN* alignment for the initial transcribed exon is a partial alignment, you can **extrapolate the TSS position** based on the number of nucleotides that are missing from the beginning of the exon. (Enter "Insufficient evidence" if you cannot determine the TSS position based on the available evidence.)

Were you able to define a TSS position based on the available evidence? NO If so, indicate in the table below the evidence that supports this TSS position

If not, were you able to define a TSS search region?11545- 11538 If so, indicate in the table below the evidence that supports the TSS search region(s)

For each evidence type, enter an "X" in the cell to indicate whether the line of evidence supports, refutes, or neither supports nor refutes the TSS annotation:

Evidence type	Support	Refute	Neither
RAMPAGE peaks and read density	X		
RNA-Seq coverage and splice junctions	X		
BLASTN alignment of the initial exon from D. melanogaster		X	
Sequence conservation with other <i>Drosophila</i> species (e.g., "Conservation" track on the Genome Browser)	X		
Other (ShortMatch)		X	

Note: The evidence type refutes the TSS annotation only if it suggests an alternate TSS position. For example, the presence of RNA-Seq read coverage upstream of the annotated TSS indicates that the TSS is located further upstream and it would be considered to be evidence against the annotated TSS; check "Refute." In contrast, the lack of RNA-Seq read coverage is a negative result that neither supports nor refutes the TSS annotation; check "Neither."

Provide an explanation if the TSS annotation is inconsistent with at least one of the evidence types specified above:

Gene Report Form

Gene name (e.g., D. bipectinata eyeless):D. bipectinata logjam_ Gene symbol (e.g., dbip_ey):dbip_loj			
Approximate location in project (from 5 Number of isoforms in <i>D. melanogaster</i> Number of isoforms in this project: 3	:3		
Complete the following table, including all of the isoforms in this project:			
Name(s) of unique isoform(s) based on coding sequence	List of isoforms with identical coding sequences		
loj-PD	loj-PE, loj-PF		
	which support the hypothesis that these isoforms are anonical splice sites, gene structure, etc.):		
Report Form for one of these isoforms column of the table above). However,	ng sequence, you only need to complete the Isoform (i.e. using the name of the isoform listed in the left you should generate GFF , transcript , and peptide spective of whether their coding sequence is identical to		
Isoform Report Form			
Complete this report form for each unit this form to create as many copies of the	que isoform listed in the table above. Copy and paste nis Isoform Report Form as needed.		
Gene-isoform symbol (e.g., dbip_ey-PA)): <u>loj-PD</u>		
Names of any additional isoforms with i	dentical coding sequences:		

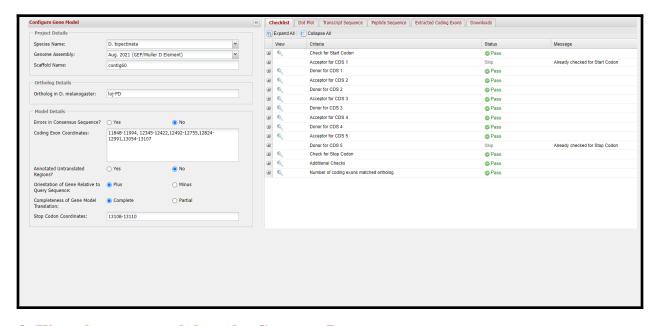
Is the 5' end of this isoform missing from the end of the project? No	
If so, how many putative exons are missing from the 5' end:	
Is the 3' end of this isoform missing from the end of the project? No	
If so, how many putative exons are missing from the 3' end:	

(Define "putative exons" based on the exons present in the *D. melanogaster* ortholog)

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.

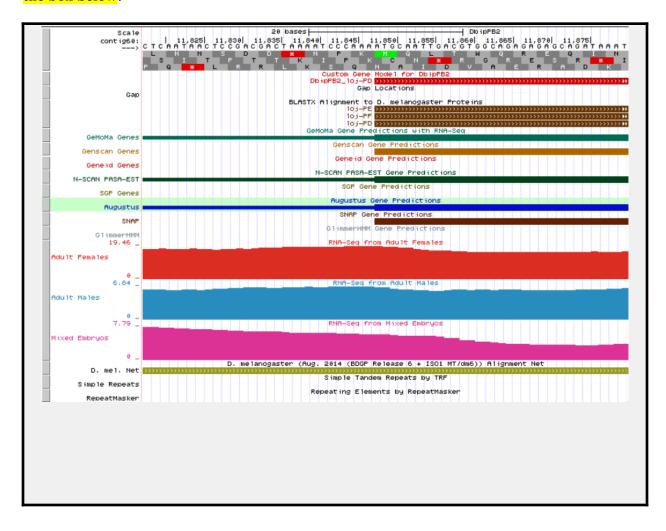


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the <u>GEP UCSC Genome Browser</u>. Zoom in so that <u>only this isoform is in</u> <u>the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

- 5. A sequence alignment track (e.g., D. mel Proteins)
- 6. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 7. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 8. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

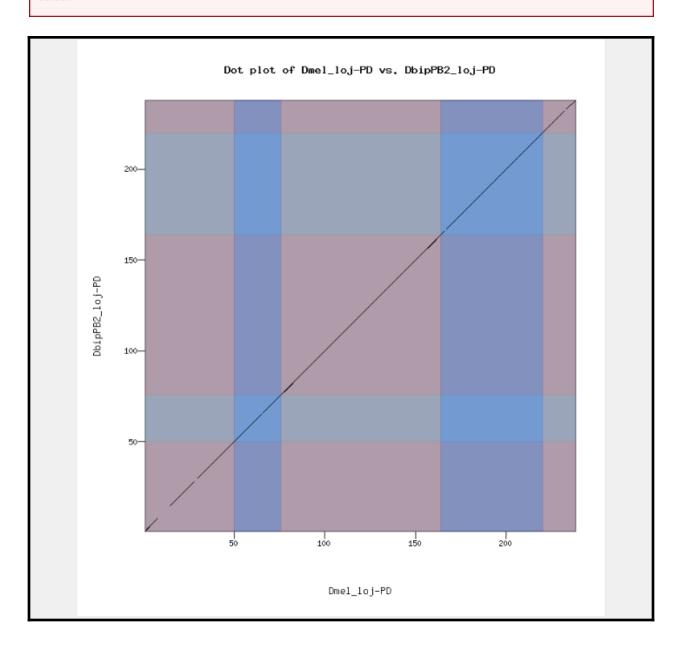
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the <u>Gene Model Checker</u> (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:

Alignment of Dmel loj-PD vs. DbipPB2 loj-PD View plain text version Download alignment image Identity: 229/237 (96.6%), Similarity: 235/237 (99.2%), Gaps: 0/237 (0.0%) Dmel loj-PD 1 MQLTWQRDQINLVLPIALICCCLLIDVTSAQEAQQPWYENLPAVAMDYKVHIDAGKEDCY 60 ************************************ 1 MQLTWQREQINLLFPIALICCCLLIDVATAQEAQQPWYENLPAVAMDYKVHIDAGKEDCY 60 DbipPB2 loj-PD Dmel_loj-PD 61 HQYVKAGATFYVSFSVVRGGDGMAGFAVRNPAGEVVKPYQWQATADYTDQVSPGGYYSVC 120 61 HQYVQAGATFYVSFSVVRGGDGMAGFAVRNPAGEVVKPYQWQATADYTDQVSPGGYYSVC 120 DbipPB2 loj-PD 121 IDNOFSRFAGKLVNIYITVVKYDAWDKYAKEIEQLQLNMQNFTATVGTVERNINDMMGYQ 180 Dmel loj-PD ********** 121 IDNQFSRFAGKLVNIYITVVKYDAWDKYAKEIEQLQLNMQNFTATIGTVERNINDMMGYQ 180 DbipPB2 loj-PD Dmel loj-PD 181 AHSRHRESRDYALLLDNNAYIQTFSISQIVVILITCSVQVFFVRKLFEVKSSSKSRI 237 ************************* 181 AHSRHRESRDYALLLDNNAYIQTFSISQIVVILITCSVQVFFVRKLFEVKSNSKSRI 237 DbipPB2 loj-PD

4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Isoform Report Form

Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

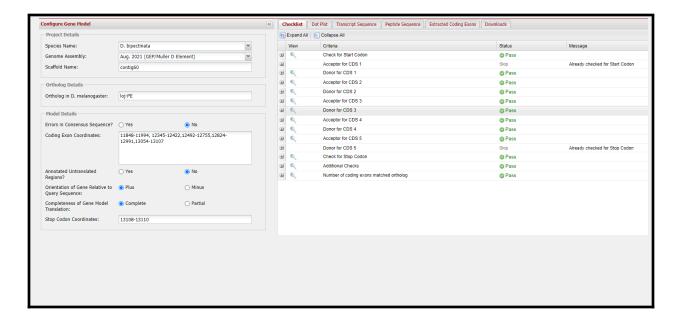
Gene-isoform symbol (e.g., dbip ey-PA): loj-PE
Names of any additional isoforms with identical coding sequences: loj-PD and loj-PF
Is the 5' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 5' end:
Is the 3' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 3' end:

(Define "putative exons" based on the exons present in the *D. melanogaster* ortholog)

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.

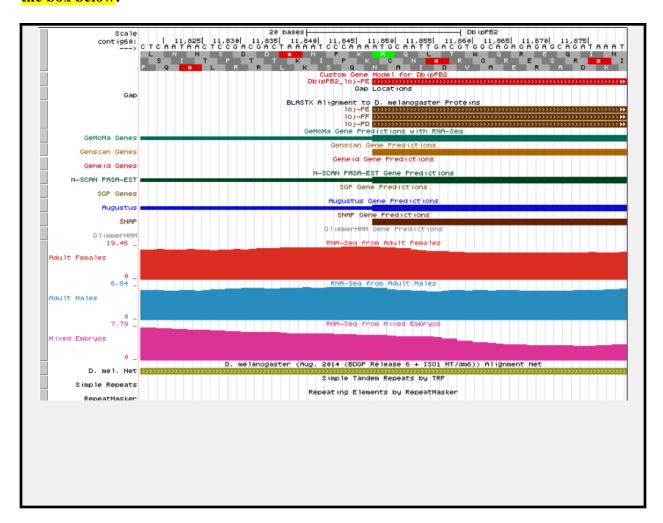


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the *GEP UCSC Genome Browser*. Zoom in so that <u>only this isoform is in</u> <u>the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

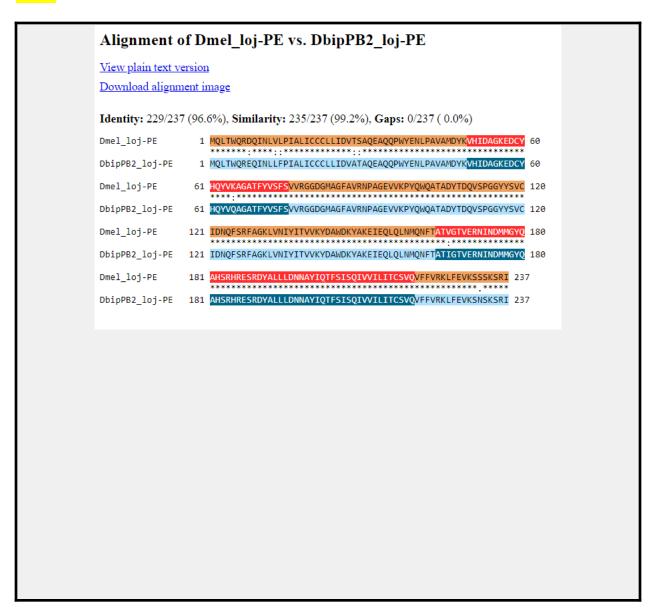
- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 3. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

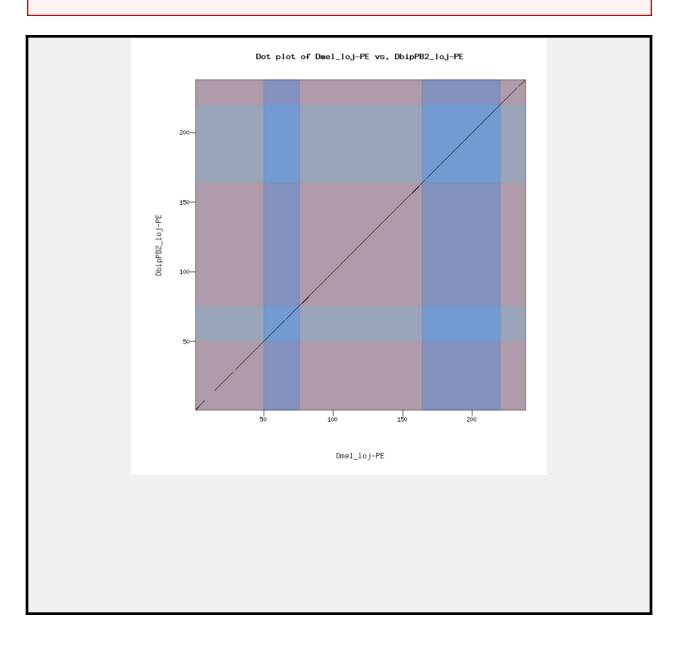
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Isoform Report Form

Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

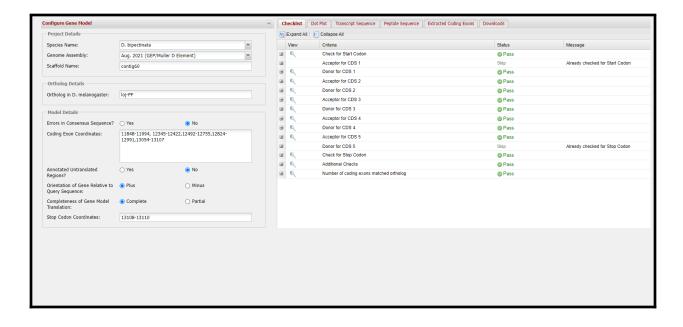
Gene-isoform symbol (e.g., dbip_ey-PA): loj-PF	
Names of any additional isoforms with identical coding sequences:	
Is the 5' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 5' end:	-
Is the 3' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 3' end:	

(Define "putative exons" based on the exons present in the *D. melanogaster* ortholog)

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.

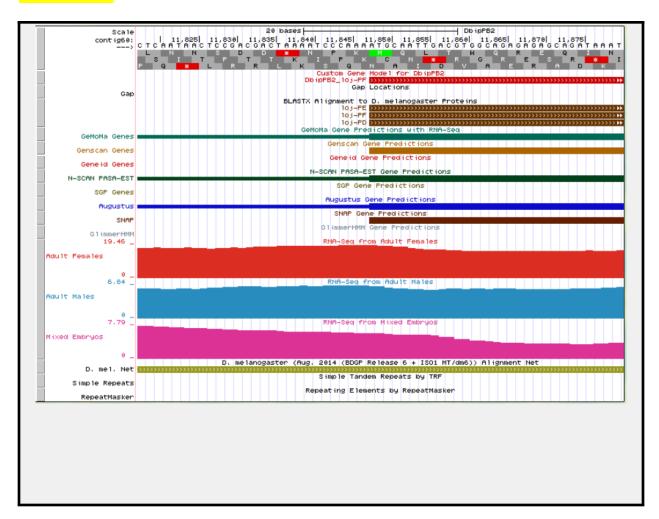


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the *GEP UCSC Genome Browser*. Zoom in so that <u>only this isoform is in</u> <u>the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

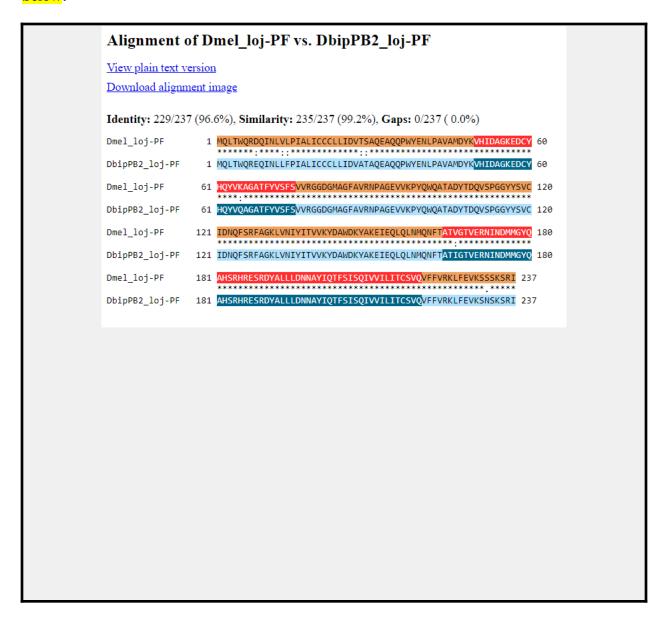
- 5. A sequence alignment track (e.g., D. mel Proteins)
- 6. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 7. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 8. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

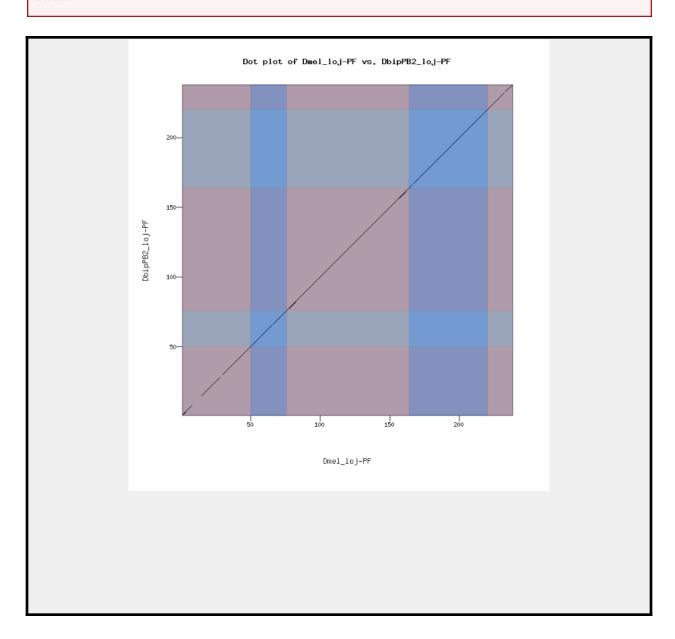
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Transcription Start Sites (TSS) Report Form (Optional)

Note: Complete this section if you have annotated the TSS for the gene above. This section is **optional** and you do not need to complete this section to submit the project.

Gene name (e.g., D. bipectinata	eyeless):logjam
Gene symbol (e.g., dbip_ey):	log

Name(s) of isoform(s) with unique TSS	List of isoforms with identical TSS
log-PD	log-PE and log-PF

Names of the isoforms with unique TSS in D. melanogaster that are absent in this species:

Provide the evidence (text and figures) which support the hypothesis that these isoforms are absent in this species (*e.g.*, changes in canonical splice sites, gene structure, etc.):

Isoform TSS Report

unique TSS listed in the table above. Copy and paste this form to create as many copies as needed.
Gene-isoform name (e.g., dbip_ey-RA):loj-PD
Names of the isoforms with the same TSS as this isoform: loj-PE and loj-PF
Type of core promoter in <i>D. melanogaster</i> (see table below): (Peaked / Intermediate / Broad / Insufficient Evidence) peaked
The type of core promoter is defined by the number of TSS annotated by the Celniker group at modENCODE and the number of DHS positions:

Complete an Isoform TSS report (through page PAGEREF _Ref91436230 \h 17) for each

1. Turn on RAMPAGE evidence tracks (Only applies to projects with these tracks)

Coordinates of the TSS position based on position with the highest RAMPAGE read density D. melanogaster: 6,094,971-6,095,063

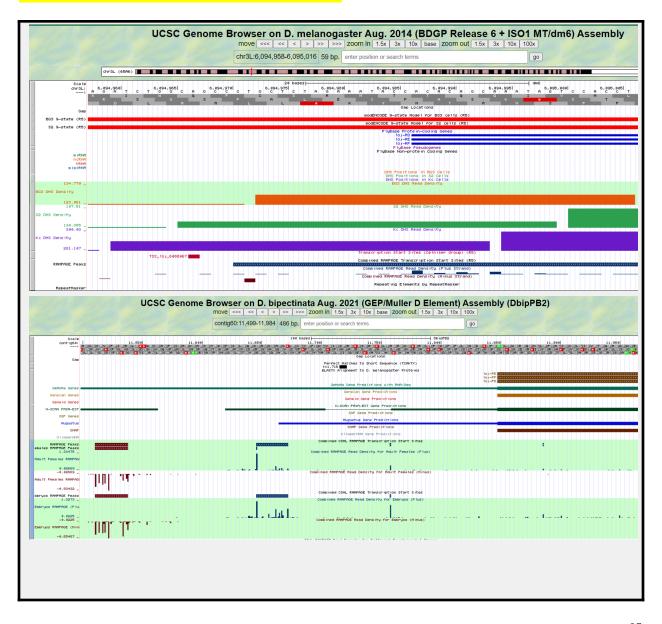
D. bipectinata: 11, 646-11,672

Coordinates of the narrow TSS search region based on RAMPAGE peaks

D. bipectinate: 11,646

D. melanogaster: 6,094,987

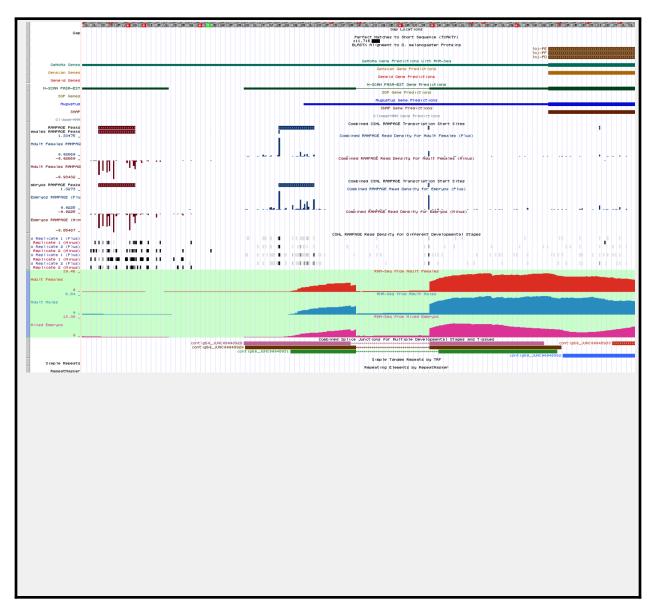
If the TSS position and narrow TSS search region are supported by RAMPAGE data, paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the Combined RAMPAGE TSS evidence track:



2. Turn on RNA-Seq evidence tracks

If the TSS annotation is supported by RNA-Seq read coverage or splice junction predictions (e.g., regtools), paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the following evidence tracks:

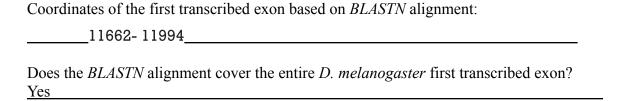
- 3. RNA-Seq Coverage or RNA-Seq Alignment Summary
- 4. Combined Splice Junctions or RNA-Seq TopHat



If the RNA-Seq evidence tracks indicate the TSS position, list it here: N/A

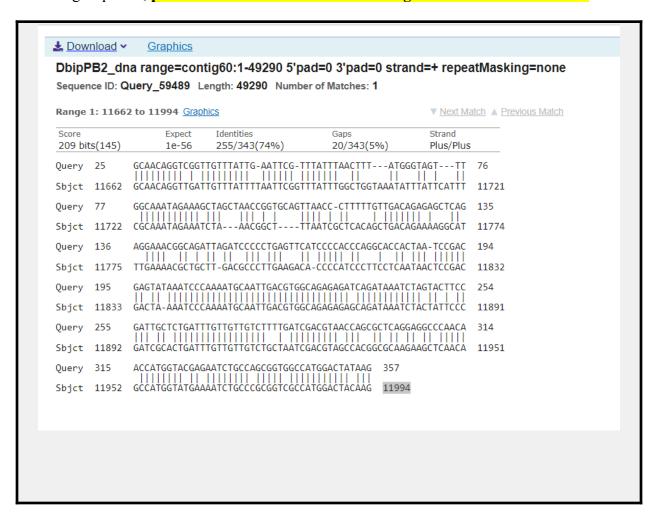
If the RNA-Seq evidence tracks indicate a TSS search region, list it here: 11,655-11,703_____

3. Annotate the first transcribed exon



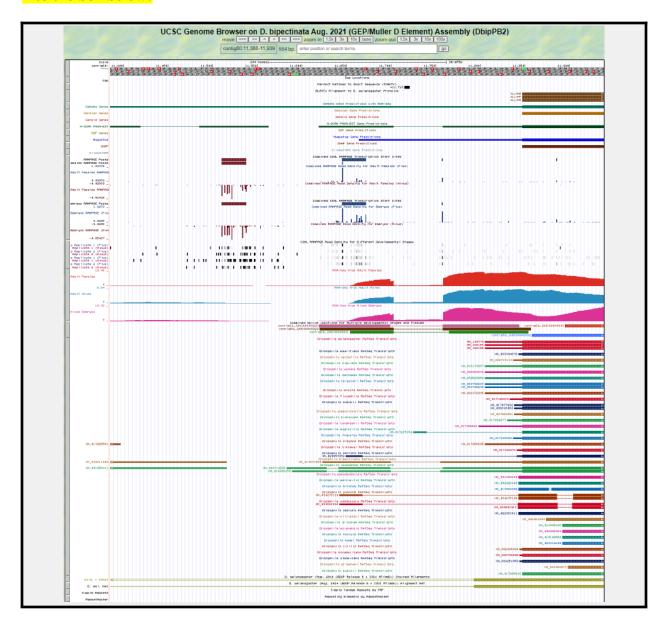
If not, specify the parts of the *D. melanogaster* exon that are missing from the *BLASTN* alignment.

If the TSS annotation is supported by *BLASTN* alignment of the initial transcribed exon against the contig sequence, paste a screenshot of the *BLASTN* alignment into the box below:



4. Turn on comparative genomics tracks

If the TSS annotation is supported by sequence conservation with other *Drosophila* species, paste a screenshot of the multiple sequence alignment (e.g., from Clustal Omega, ROAST) into the box below:



5. Summarize the evidence that supports the TSS annotation postulated above

Coordinate(s) of the TSS position(s):

Based on RAMPAGE data (if applicable):11,646 Based on RNA-Seq data:11,655-11,703 Based on BLASTN alignment:11,662-11,672 Based on other evidence (please specify):11,656-11,667 based on comparative genomics tracks	
Note: If the <i>BLASTN</i> alignment for the initial transcribed exon is a partial alignment, you can extrapolate the TSS position based on the number of nucleotides that are missing from the beginning of the exon. (Enter "Insufficient evidence" if you cannot determine the TSS position based on the available evidence.)	
Were you able to define a TSS position based on the available evidence?yes; 11,662 If so, indicate in the table below the evidence that supports this TSS position	
If not, were you able to define a TSS search region?	

For each evidence type, enter an "X" in the cell to indicate whether the line of evidence supports, refutes, or neither supports nor refutes the TSS annotation:

If so, indicate in the table below the evidence that supports the TSS search region(s)

Evidence type	Support	Refute	Neither
RAMPAGE peaks and read density	yes		
RNA-Seq coverage and splice junctions	yes		
BLASTN alignment of the initial exon from D. melanogaster	yes		
Sequence conservation with other <i>Drosophila</i> species (e.g., "Conservation" track on the Genome Browser)	yes		
Other (please specify) [e.g., RefSeq Genes, N-SCAN PASA-EST, Augustus TSS predictions; histone modifications (ChIP-Seq data)].			

Note: The evidence type refutes the TSS annotation only if it suggests an alternate TSS position. For example, the presence of RNA-Seq read coverage upstream of the annotated TSS indicates that the TSS is located further upstream and it would be considered to be evidence against the annotated TSS; check "Refute." In contrast, the lack of RNA-Seq read coverage is a negative result that neither supports nor refutes the TSS annotation; check "Neither."

Gene Report Form

Ets65A-PB

Names o	f the isoforms with	unique coding sequence	es in D. meland	ogaster that are	absent in this
species:	N/A				

Provide the evidence (text and figures) which support the hypothesis that these isoforms are absent in this species (*e.g.*, changes in canonical splice sites, gene structure, etc.):

Note: For isoforms with identical coding sequence, you only need to complete the Isoform Report Form for one of these isoforms (i.e. using the name of the isoform listed in the left column of the table above). However, you should **generate GFF**, **transcript**, **and peptide sequence files for <u>ALL</u> isoforms**, irrespective of whether their coding sequence is identical to that of another isoform.

Consensus Sequence Errors Report Form

Complete this section if you have identified errors in the project consensus sequence that affect the annotation of the gene described above.

All of the coordinates reported in this section should be <u>relative to the coordinates of the original project sequence</u>.

Location(s) in the project sequence with consensus errors: Ets65A-PB: exon 2 is missing from D. bipectinata, but present in D. melanogaster. Coordinates: 35133-35183
1. Evidence that supports the consensus errors postulated above
Note: Evidence that could be used to support the hypothesis of errors within the consensus sequence includes a CDS alignment with frame shifts or in-frame stop codons, and RNA-Seq reads with discrepant alignments compared to the project sequence.
2. Generate a VCF file which describes the changes to the consensus sequence
Use the <u>Sequencer Updater</u> to create a Variant Call Format (VCF) file that describes the changes to the consensus sequence you have identified above. Paste a screenshot with the list of sequence changes into the box below:
Isoform Report Form
Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.
Gene-isoform symbol (e.g., dbip_ey-PA):dbip_Ets65A-PA
Names of any additional isoforms with identical coding sequences:

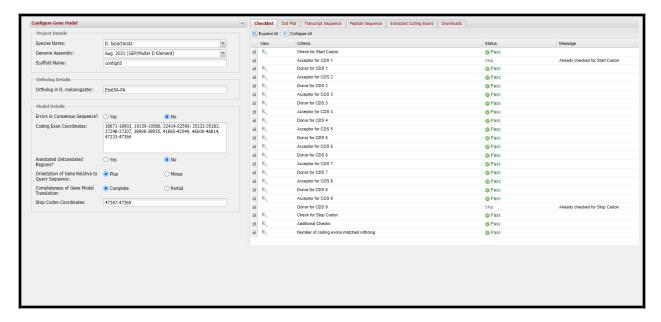
Is the 5' end of this isoform missing from the end of the project? No	
If so, how many putative exons are missing from the 5' end:	
Is the 3' end of this isoform missing from the end of the project? No	
If so, how many putative exons are missing from the 3' end:	

(Define "putative exons" based on the exons present in the *D. melanogaster* ortholog)

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.



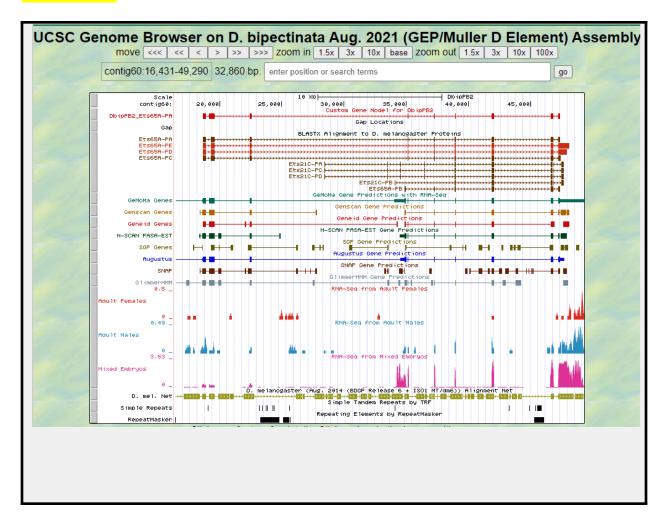
2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the *GEP UCSC Genome Browser*. Zoom in so that <u>only this isoform is in</u> <u>the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

- 9. A sequence alignment track (e.g., D. mel Proteins)
- 10. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 11. At least one RNA-Seq track (e.g., RNA-Seq Coverage)

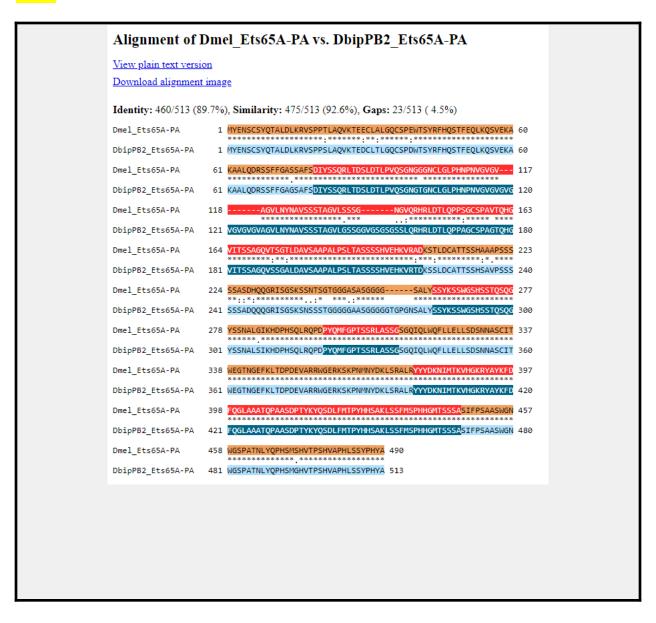
12. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

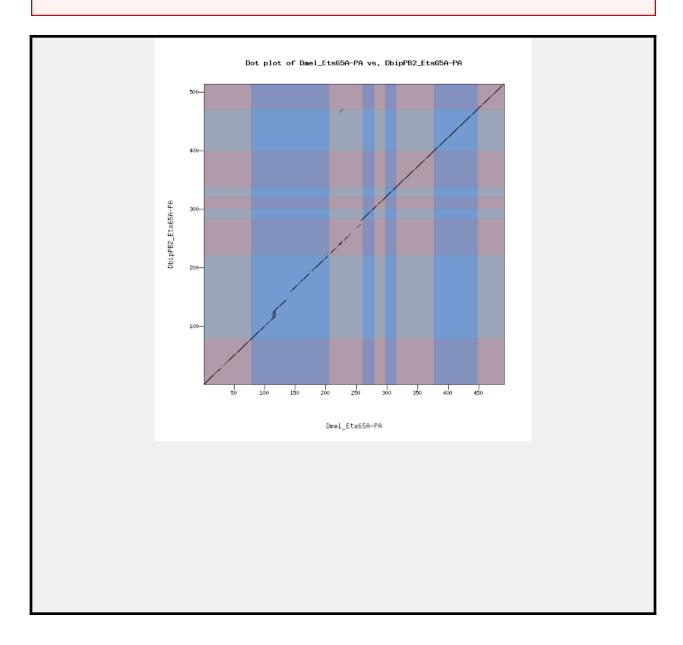
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Isoform Report Form

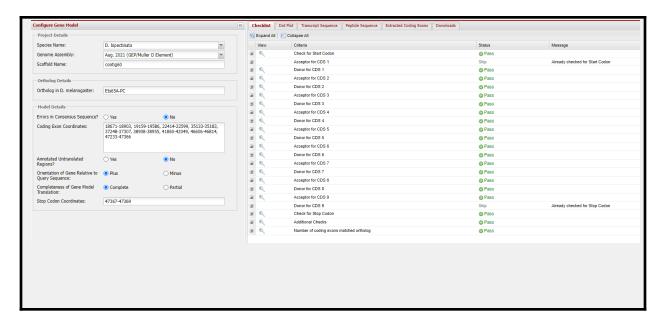
Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

Gene-isoform symbol (e.g., dbip_ey-PA):dbip_Ets65A-PC	
Names of any additional isoforms with identical coding sequences: Dbip Ets65A-PA	
	
Is the 5' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 5' end:	
Is the 3' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 3' end:	
(Define "putative exons" based on the exons present in the <i>D. melanogaster</i> ortholog)	

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.

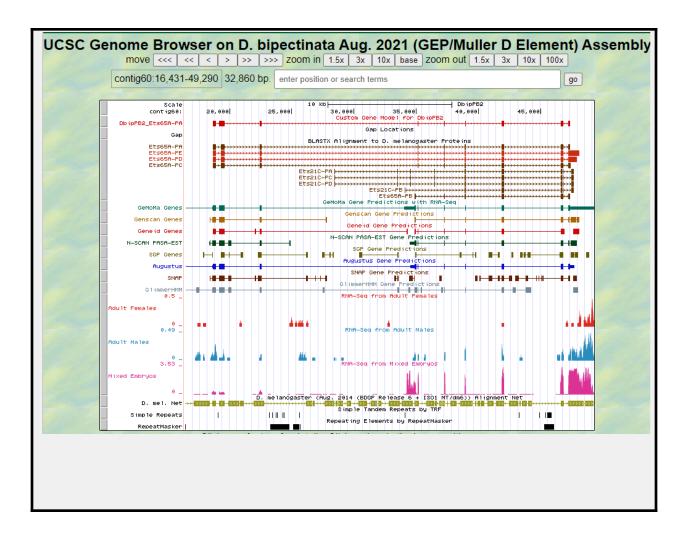


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the *GEP UCSC Genome Browser*. Zoom in so that <u>only this isoform is in</u> <u>the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

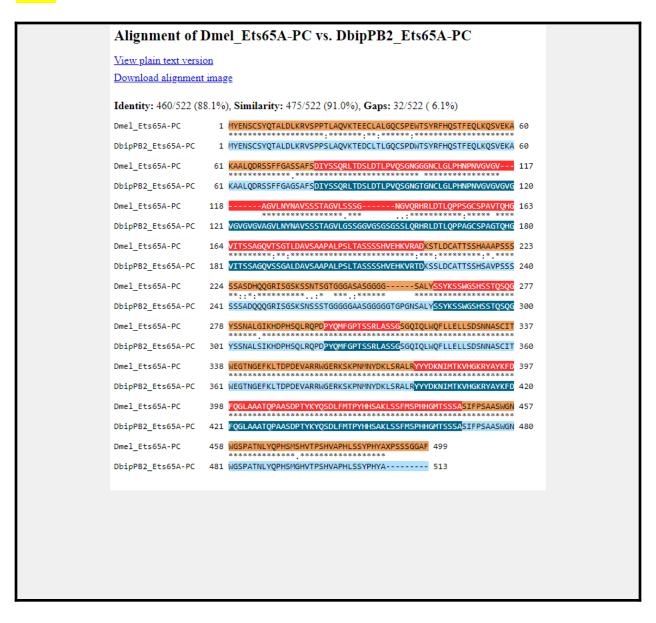
- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 3. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

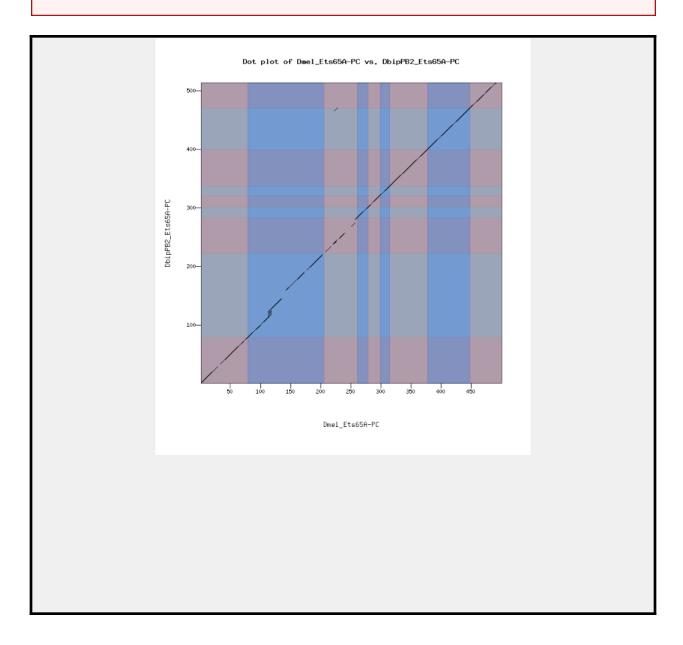
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Isoform Report Form

Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

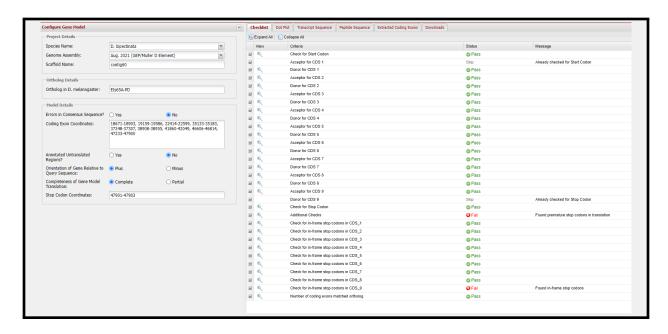
Gene-isoform symbol (e.g., dbip ey-PA): dbip Ets65A-PD
Names of any additional isoforms with identical coding sequences: N/A
Is the 5' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 5' end:
Is the 3' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 3' end:

(Define "putative exons" based on the exons present in the *D. melanogaster* ortholog)

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.

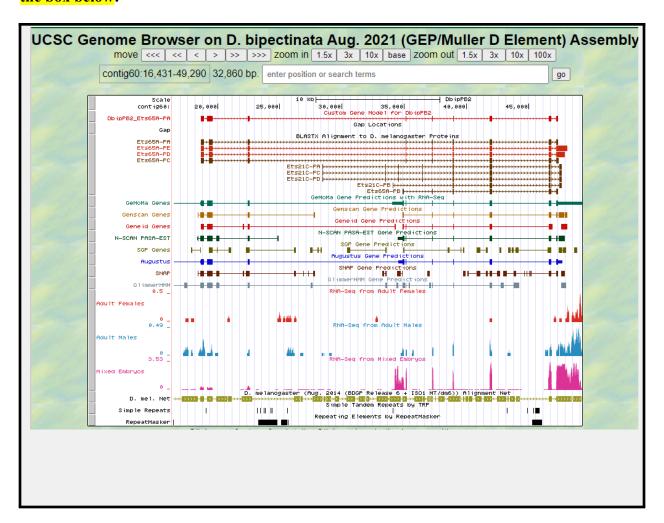


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the GEP UCSC Genome Browser. Zoom in so that <u>only this isoform is in</u> the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:

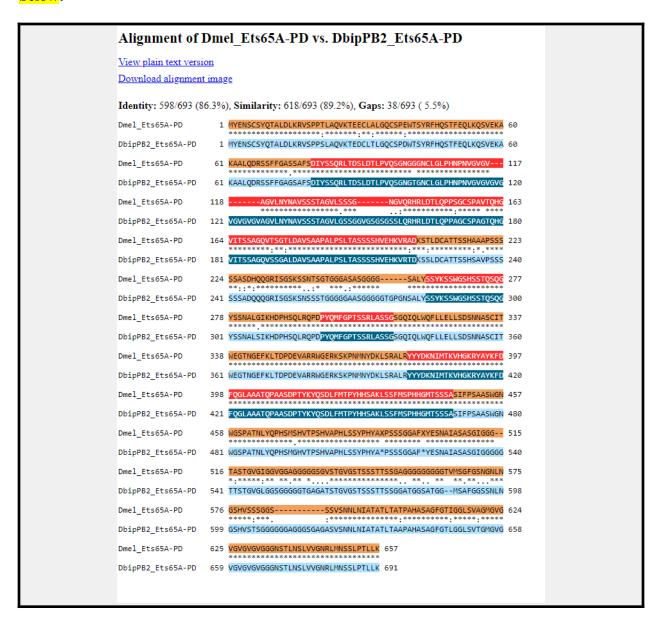
- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 3. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

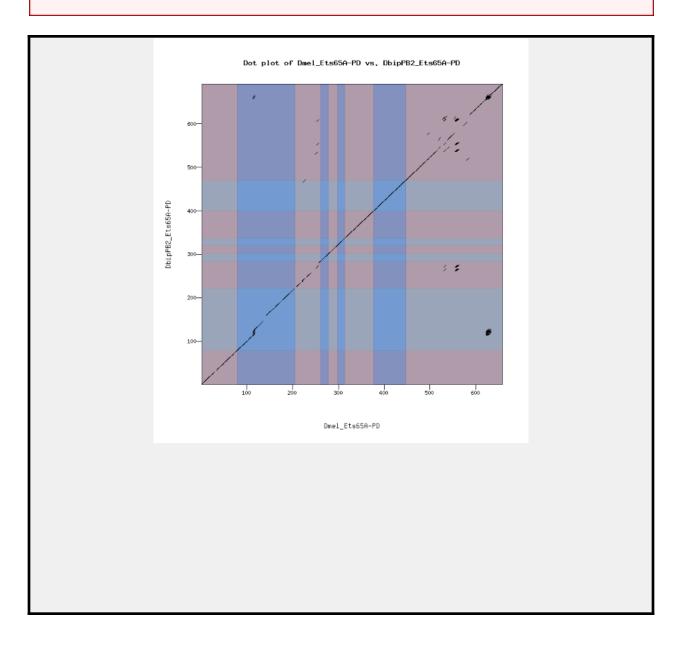
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Isoform Report Form

Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

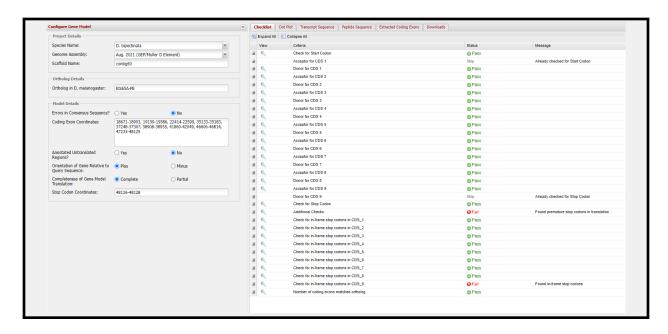
Gene-isoform symbol (e.g., dbip ey-PA): dbip Ets65A-PE	
Names of any additional isoforms with identical coding sequences: N/A	
Is the 5' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 5' end:	
Is the 3' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 3' end:	

(Define "putative exons" based on the exons present in the *D. melanogaster* ortholog)

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.

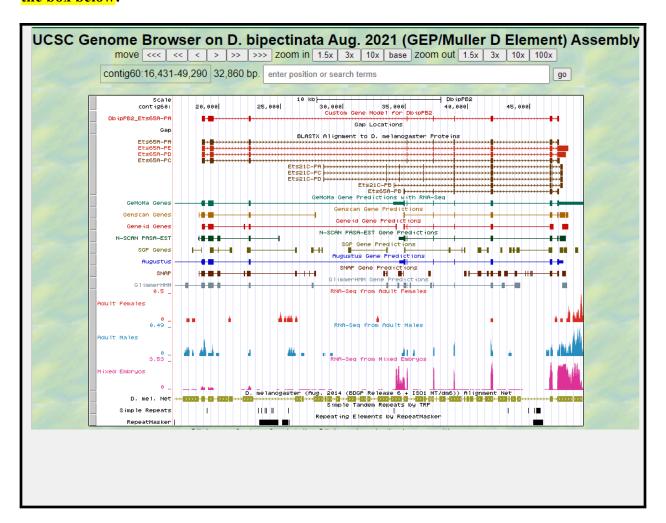


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the GEP UCSC Genome Browser. Zoom in so that <u>only this isoform is in</u> the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:

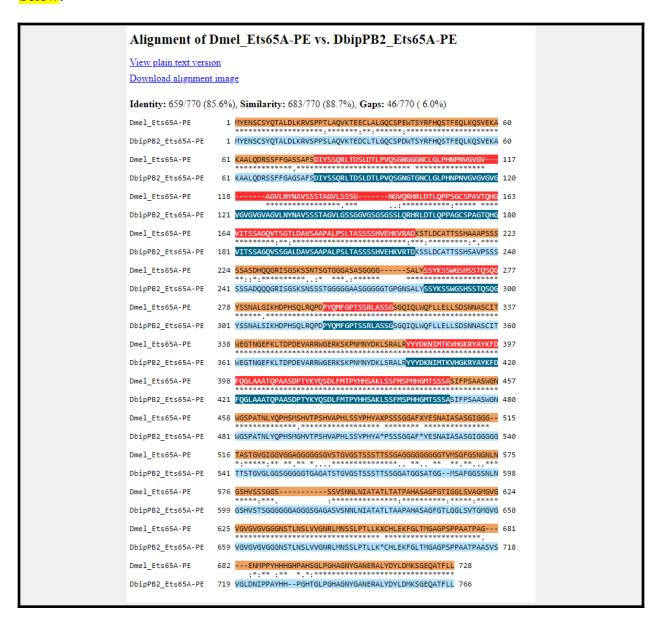
- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 3. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

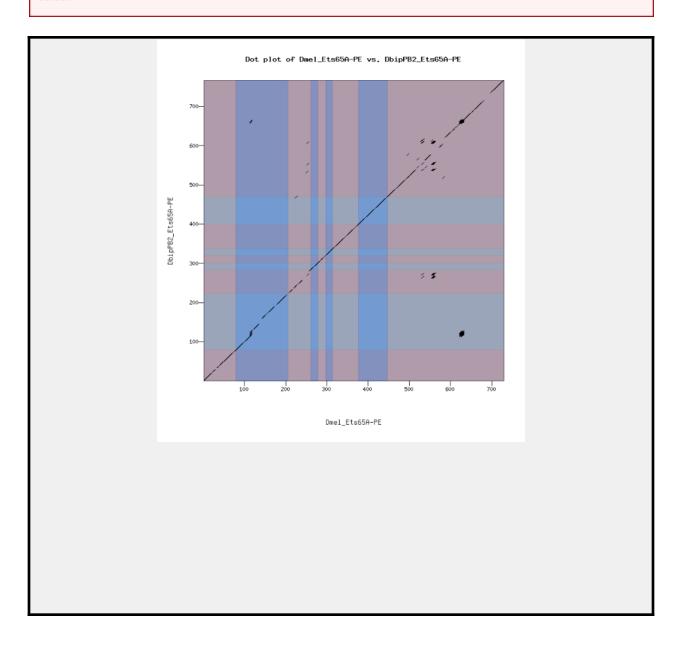
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Isoform Report Form

Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

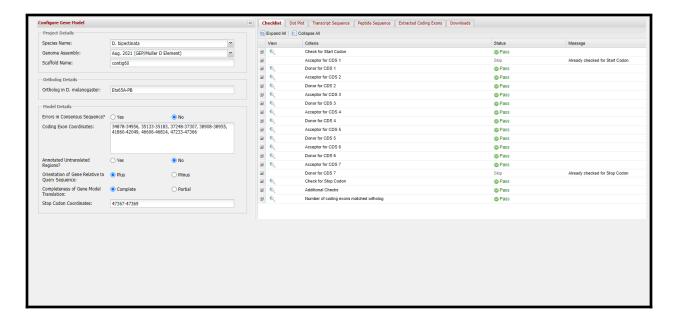
Gene-isoform symbol (e.g., dbip_ey-PA):dbip_Ets65A-PB	
Names of any additional isoforms with identical coding sequences: N/A	
Is the 5' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 5' end:	
Is the 3' end of this isoform missing from the end of the project? No If so, how many putative exons are missing from the 3' end:	

(Define "putative exons" based on the exons present in the *D. melanogaster* ortholog)

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the <u>Gene Model Checker</u> and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the *Gene Model Checker*. The *Gene Model Checker* will use this VCF file to automatically revise the submitted exon coordinates.

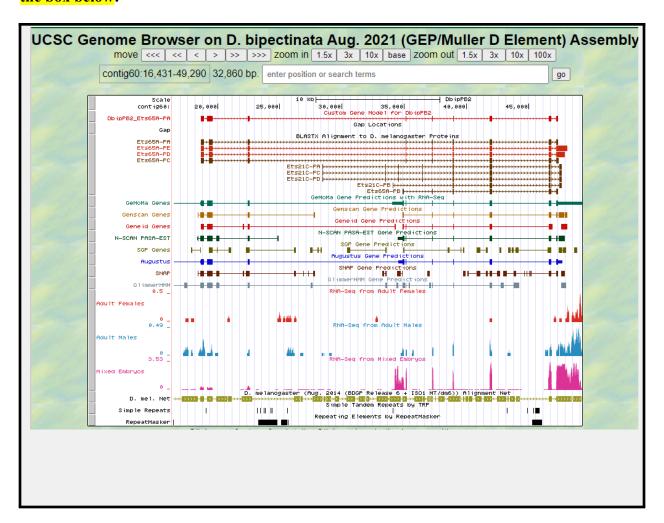


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the GEP UCSC Genome Browser. Zoom in so that <u>only this isoform is in</u> the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:

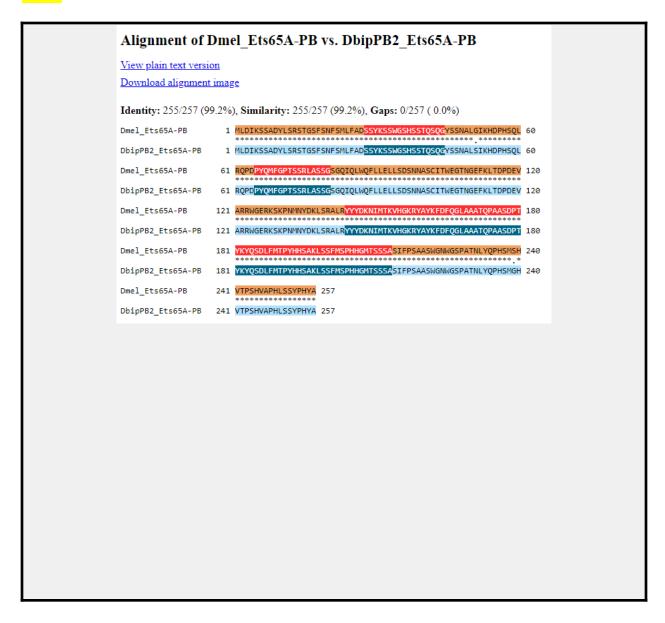
- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan, GeMoMa)
- 3. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

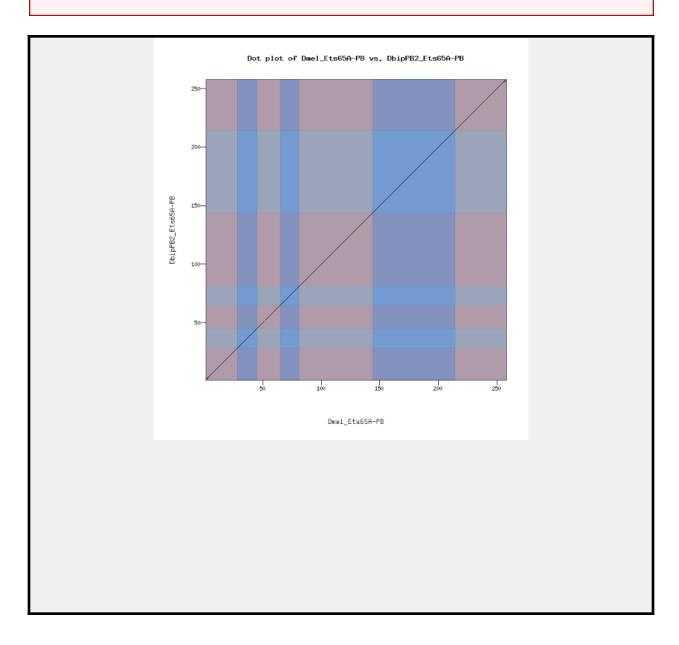
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the *Gene Model Checker* (available through the "View protein alignment" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST website. Paste a screenshot of the protein alignment into the box below:



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the <u>Gene Model Checker</u>) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, which would appear as kinks in the diagonal line; regions with no sequence similarity; indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Transcription Start Sites (TSS) Report Form 2 (Optional)

Note: Complete this section if you have annotated the TSS for the gene above	. This section is
optional and you do not need to complete this section to submit the project.	

Gene name (e.g., D. bipectinata eyeless):	D. bipectinate Ets at 65A
Gene symbol (e.g., dbip ey): $\underline{\qquad}$	bip Ets65A

Name(s) of isoform(s) with unique TSS	List of isoforms with identical TSS
Ets65A-PA	Ets65A-PC, Ets65A-PD, Ets65A-PE
Ets65A-PB	

Names of the isoforms with unique TSS in D. melanogaster that are absent in this species:

Provide the evidence (text and figures) which support the hypothesis that these isoforms are absent in this species (*e.g.*, changes in canonical splice sites, gene structure, etc.):

Isoform TSS Report

Complete an Isoform TSS report (through page PAGEREF _Ref91436230 \h 17) for each unique TSS listed in the table above. Copy and paste this form to create as many copies as needed.

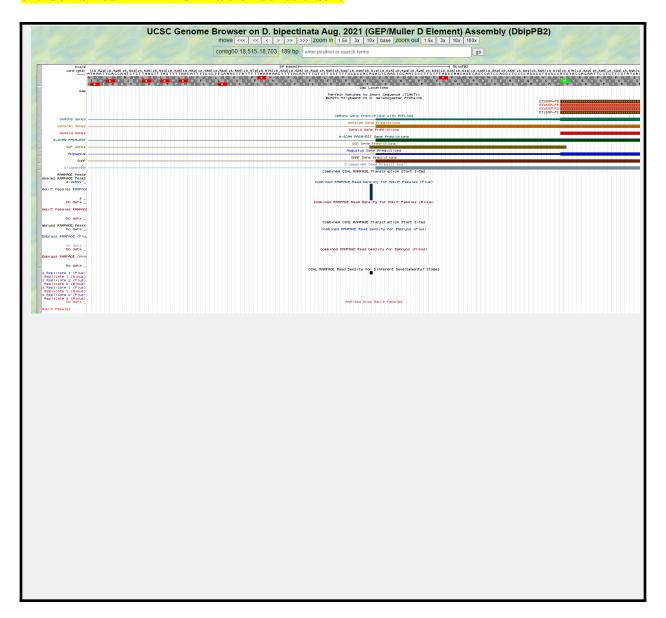
Gene-isoform name (e.g., dbip ey-RA):dbip Ets65A-PA
Names of the isoforms with the same TSS as this isoform: <u>dbip_Ets65A-PC</u> , <u>dbip_Ets65A-PD</u> , <u>dbip_Ets65A-PE</u>
Type of core promoter in <i>D. melanogaster</i> (see table below): (Peaked / Intermediate / Broad / Insufficient Evidence) peaked
The type of core promoter is defined by the number of TSS annotated by the Celniker group at modENCODE and the number of DHS positions:

1. Turn on RAMPAGE evidence tracks (Only applies to projects with these tracks)

Coordinates of the TSS position based on position with the highest RAMPAGE read density 18,608

Coordinates of the narrow TSS search region based on RAMPAGE peaks 18,608

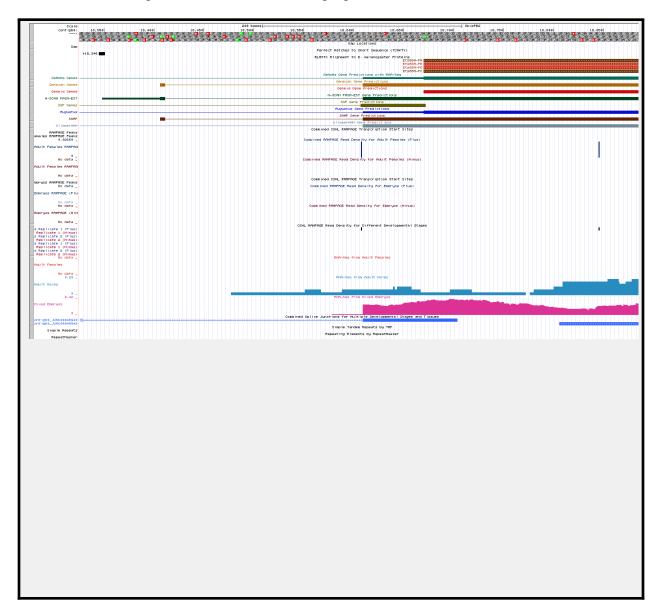
If the TSS position and narrow TSS search region are supported by RAMPAGE data, paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the Combined RAMPAGE TSS evidence track:



2. Turn on RNA-Seq evidence tracks

If the TSS annotation is supported by RNA-Seq read coverage or splice junction predictions (e.g., regtools), paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the following evidence tracks:

- 5. RNA-Seq Coverage or RNA-Seq Alignment Summary
- 6. Combined Splice Junctions or RNA-Seq TopHat



If the RNA-Seq evidence tracks indicate the TSS position, list it here:

If the RNA-Seq evidence tracks indicate a TSS search region, list it here: 18,600-18,670

3. Annotate the first transcribed exon

Coordinates of the first transcribed exon based on *BLASTN* alignment: 14473 to 15229

Does the *BLASTN* alignment cover the entire *D. melanogaster* first transcribed exon? No

If not, specify the parts of the *D. melanogaster* exon that are missing from the *BLASTN* alignment.

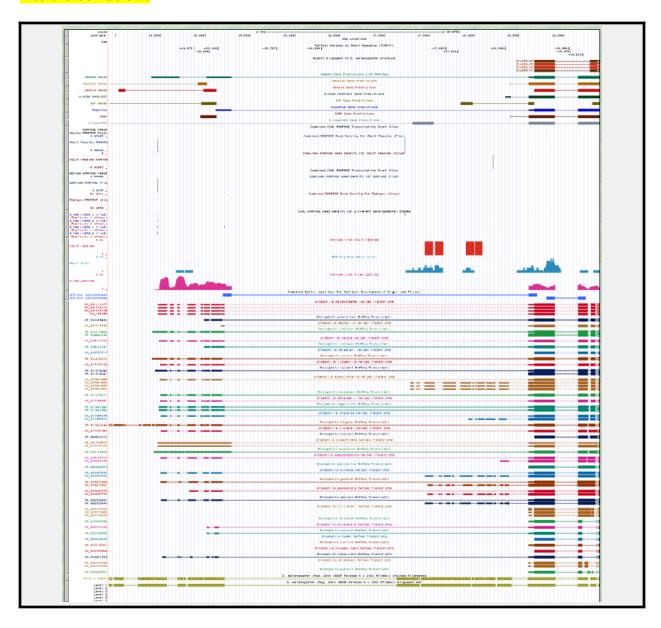
Missing the entire first exon. Coordinates: 18671-18903

If the TSS annotation is supported by *BLASTN* alignment of the initial transcribed exon against the contig sequence, paste a screenshot of the *BLASTN* alignment into the box below:

_	na range=contig60:1-49290 5 Query_2027		repeatMasking=none
Range 1: 144	73 to 15229 Graphics	•	Next Match A Previous Match
Score 479 bits(334)	Expect Identities 2e-137 591/810(73%)		Strand Plus/Plus
Query 1 Sbjct 14473			
Query 61 Sbjct 14533	TTTGCGGTTTAGATTCGTCGGTCACACGT		H
Query 113 Sbjct 14593		GCAAAAAGGAAAACAAACG	 GCAGCA 14645
Query 172 Sbjct 14646			
Query 232 Sbjct 14695			GAAAAA 14754
Query 281 Sbjct 14755		CCATACAATAGTAACAATTCACATG	 GTCCT 14814
Query 322 Sbjct 14815	ACCAGTGTGTCGCGGAAATTTTTGTGCGG		 TTCAT 14874
Query 382 Sbjct 14875		CTGCACTCCT-GCAGTCCTTCAGCGC	
Query 438 Sbjct 14934	ŤŤŤĊĠŤĊĊĊĠŦŦAŦŤĠŤĠĊÀ		CCTTGG 14984
Query 492 Sbjct 14985		ACTCGTCCTTGCACTC	 GACAAA 15026
Query 552 Sbjct 15027		 GAACGAAGGCTTGCGAAACCGTT	 ATTTG 15080
Query 612 Sbjct 15081			 AATTT 15140
Query 672 Sbjct 15141		GAGTTGAGTGATTAACTGCACCCGGG	
Query 731 Sbjct 15201	CGATTGTGAAGTGTTTAAGATTACTAAGT 		

4. Turn on comparative genomics tracks

If the TSS annotation is supported by sequence conservation with other *Drosophila* species, paste a screenshot of the multiple sequence alignment (e.g., from Clustal Omega, ROAST) into the box below:



5. Summarize the evidence that supports the TSS annotation postulated above

Coordinate(s) of the TSS position(s):
Based on RAMPAGE data (if applicable): 14,472; 14,474
Based on RNA-Seq data: <u>14,470-15,200</u>
Based on <i>BLASTN</i> alignment: 14,474
Based on other evidence (please specify):
Note: If the <i>BLASTN</i> alignment for the initial transcribed exon is a partial alignment, you can extrapolate the TSS position based on the number of nucleotides that are missing from the
beginning of the exon. (Enter "Insufficient evidence" if you cannot determine the TSS

Vere you able to define a TSS position based on the available evidence?
yes
f so, indicate in the table below the evidence that supports this TSS position
f not, were you able to define a TSS search region?
f so, indicate in the table below the evidence that supports the TSS search region(s)

position based on the available evidence.)

For each evidence type, enter an "X" in the cell to indicate whether the line of evidence supports, refutes, or neither supports nor refutes the TSS annotation:

Evidence type	Support	Refute	Neither
RAMPAGE peaks and read density	yes		
RNA-Seq coverage and splice junctions	yes		
BLASTN alignment of the initial exon from D. melanogaster	yes		
Sequence conservation with other <i>Drosophila</i> species (<i>e.g.</i> , "Conservation" track on the Genome Browser)	yes		
Other (please specify) [e.g., RefSeq Genes, N-SCAN PASA-EST, Augustus TSS predictions; histone modifications (ChIP-Seq data)].			

Note: The evidence type refutes the TSS annotation only if it <u>suggests an alternate TSS</u> <u>position</u>. For example, the presence of RNA-Seq read coverage upstream of the annotated TSS indicates that the TSS is located further upstream and it would be considered to be evidence against the annotated TSS; check "Refute." In contrast, the lack of RNA-Seq read coverage is a negative result that neither supports nor refutes the TSS annotation; check "Neither."

Provide an explanation if the TSS annotation is inconsistent with at least one of the evidence types specified above:

Isoform TSS Report

Complete an Isoform TSS report (through page PAGEREF _Ref91436230 \h 17) for each unique TSS listed in the table above. Copy and paste this form to create as many copies as needed.

Gene-isoform name (e.g., dbip_ey-RA):dbip_Ets65A-PB	
Names of the isoforms with the same TSS as this isoform: N/A	
Type of core promoter in <i>D. melanogaster</i> (see table below): (Peaked / Intermediate / Broad / Insufficient Evidence) peaked	
The type of core promoter is defined by the number of TSS annotated by the Celniker group modENCODE and the number of DHS positions:	o at

1. Turn on RAMPAGE evidence tracks (Only applies to projects with these tracks)

Coordinates of the TSS position based on position with the highest RAMPAGE read density 34,400-34,865

Coordinates of the narrow TSS search region based on RAMPAGE peaks 34,438; 34,444; 34,445; 34,865

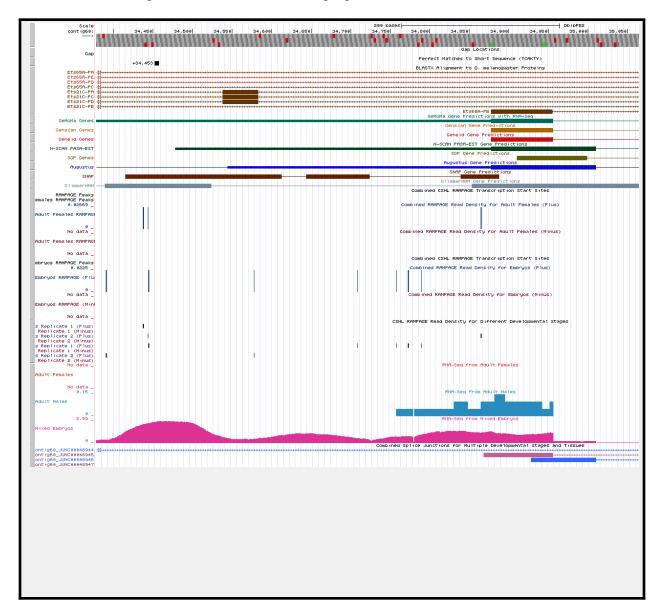
If the TSS position and narrow TSS search region are supported by RAMPAGE data, paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the Combined RAMPAGE TSS evidence track:



2. Turn on RNA-Seq evidence tracks

If the TSS annotation is supported by RNA-Seq read coverage or splice junction predictions (e.g., regtools), paste a Genome Browser screenshot of the region surrounding the putative TSS (±300bp) showing the following evidence tracks:

- 1. RNA-Seq Coverage or RNA-Seq Alignment Summary
- 2. Combined Splice Junctions or RNA-Seq TopHat



If the RNA-Seq evidence tracks indicate the TSS position, list it here:

If the RNA-Seq evidence tracks indicate a TSS search region, list it here: _34,756-34,865____

3. Annotate the first transcribed exon

Coordinates of the first transcribed exon based on *BLASTN* alignment: 34214 to 34956

Does the *BLASTN* alignment cover the entire *D. melanogaster* first transcribed exon? Yes

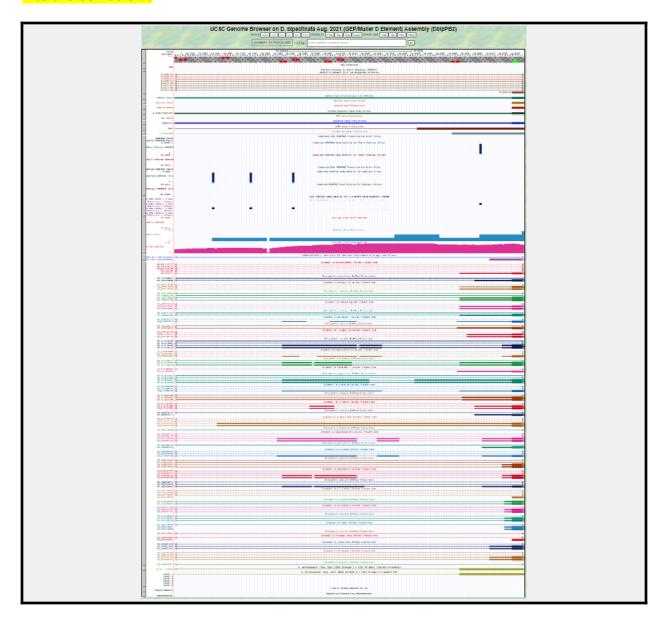
If not, specify the parts of the D. melanogaster exon that are missing from the BLASTN alignment.

If the TSS annotation is supported by *BLASTN* alignment of the initial transcribed exon against the contig sequence, paste a screenshot of the *BLASTN* alignment into the box below:

Range 1: 342	14 to 34956 Graph	nics		▼ Next Ma	atch A Previous Match
Score 562 bits(392	Expect 2e-162	Identities 599/774(77%)	Gaps 73/774(9%)	Strand Plus/Plus	5
Query 1		CGTGCCACAGATCGGACGC			60
Sbjct 3421					34273
Query 61	GTC	CTAGCCAGGAGC		CAGTGATTT	103
Sbjct 3427	4 GTCGTCTAGCTCG	 ATATCCTCGCCAGGAGAGC		AGTGATTT	34329
Query 104		TGGGCCTACCCACACATAT			155
Sbjct 3433		TGGGCCTACCCGCAAATAC			34387
Query 156		AGTGCTGAAAAAATATCAC		ATTGACACAT	210
Sbjct 3438		AACACAACCAAAAAATCAC			34447
Query 211		ACACGCGAACGGCAGGCAG			270
Sbjct 3444		ACACGCGAACGGCAGGCAG			34507
Query 271		TCCGGTAAACGGTAACCGG			330
Sbjct 3450	3 ĠĠĀĀĠŤĊĠĀĠĠŤĠ	TCCGGTAAACGGTAACCGG	ttcccatcctcactttc	ccgttccccgt	34567
Query 331		CCGCTGCGCGTCGCTGCTC	CACCGCTGCCCATTTGCC		390
Sbjct 3456		CCGCTGAGCGTCGCTGC			34614
Query 391		TCAATTGCGGACACCACTT 	11111111 11111		444
Sbjct 3461		TCAATTGTGGACACCACTT			34674
Query 445		CCTCCGGCGACCAATTTAC			504
Sbjct 3467		GCTCCGGCGACCAATTTAC			34734
Query 505 Sbjct 3473		GACAAGGACAAAG 			558 34792
Query 559		ACTAATTGCTGTGTGTGTG			618
Sbict 3479				111	34842
Query 619		TACCCAACCGACGGCGGCC			678
Sbjct 3484		AACCCATCCGCCGGCGGTC			34902
Query 679	ATTACTTGTCACG	CTCCACGGGCAGCTTCAGC	AACTTCTCGATGCTCTTC	GCAG 732	
Sbjct 3490					j

4. Turn on comparative genomics tracks

If the TSS annotation is supported by sequence conservation with other *Drosophila* species, paste a screenshot of the multiple sequence alignment (e.g., from Clustal Omega, ROAST) into the box below:



5. Summarize the evidence that supports the TSS annotation postulated above

Coordinate(s) of the TSS position(s):	
Based on RAMPAGE data (if applicable): 34,865	
Based on RNA-Seq data: 34,865	
Based on <i>BLASTN</i> alignment: 34,865	
Based on other evidence (please specify):	
· · · · · · · · · · · · · · · · · · ·	

Note: If the *BLASTN* alignment for the initial transcribed exon is a partial alignment, you can **extrapolate the TSS position** based on the number of nucleotides that are missing from the beginning of the exon. (Enter "Insufficient evidence" if you cannot determine the TSS position based on the available evidence.)

Vere you able to define a TSS position based on the available evidence?	
yes;34,865	
f so, indicate in the table below the evidence that supports this TSS position	
f not, were you able to define a TSS search region?	
f so, indicate in the table below the evidence that supports the TSS search region(s)

For each evidence type, enter an "X" in the cell to indicate whether the line of evidence supports, refutes, or neither supports nor refutes the TSS annotation:

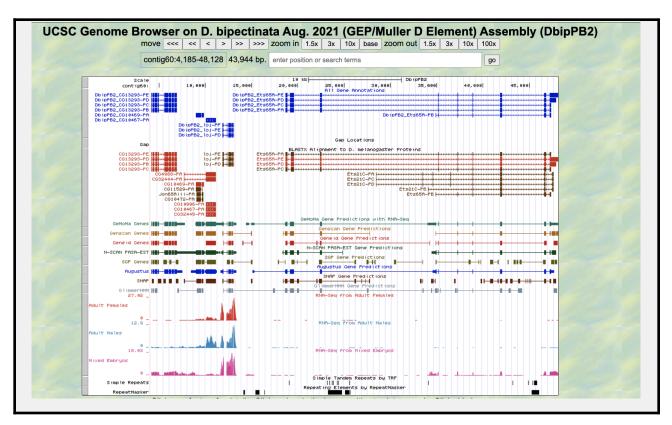
Evidence type	Support	Refute	Neither
RAMPAGE peaks and read density	yes		
RNA-Seq coverage and splice junctions	yes		
BLASTN alignment of the initial exon from D. melanogaster	yes		
Sequence conservation with other <i>Drosophila</i> species (<i>e.g.</i> , "Conservation" track on the Genome Browser)	yes		
Other (please specify) [e.g., RefSeq Genes, N-SCAN PASA-EST, Augustus TSS predictions; histone modifications (ChIP-Seq data)].			

Note: The evidence type refutes the TSS annotation only if it <u>suggests an alternate TSS</u> <u>position</u>. For example, the presence of RNA-Seq read coverage upstream of the annotated TSS indicates that the TSS is located further upstream and it would be considered to be evidence against the annotated TSS; check "Refute." In contrast, the lack of RNA-Seq read coverage is a negative result that neither supports nor refutes the TSS annotation; check "Neither."

Preparing the Project for Submission

For each project, you should prepare the project GFF, transcript, and peptide sequence files for **ALL** isoforms along with this report. You can combine the individual files generated by the *Gene Model Checker* into a single file using the *Annotation Files Merger*. Once you have combined the GFF files into a single file, click on the "**Show Track**" button to view all the gene models in the combined GFF file within the Genome Browser.

Paste a screenshot (generated by the *Annotation Files Merger*) with all the gene models you have annotated in this project into the box below.



Thank you for your submission, and congratulations on completing your analysis of this region of this genome. Our planned GEP meta-analysis of the genes and genomes in this study depends on the high quality annotations accomplished by GEP students.